

SYNOPSIS OF THE MORPHOLOGY AND TAXONOMY OF *CAREX* SECTION  
*GLAUCESCENTES* IN NORTH AMERICA

A Thesis

by

DIANE COSTON MCLAUGHLIN

Submitted to the Office of Graduate Studies of  
Texas A&M University  
in partial fulfillment of the requirements for the degree of

MASTER OF SCIENCE

August 2004

Major Subject: Rangeland Ecology and Management

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## ABSTRACT

The Morphology and Taxonomy of *Carex* Section

*Glaucescences* in North America. (August 2004)

Diane Coston McLaughlin, B.S., Texas A&M University

Chair of Advisory Committee: Dr. Stephan L. Hatch

Field studies were used to characterize habitat and evaluate morphological characters of *Carex glaucescens*, *C. jorii* and *C. verrucosa*. Morphometric analysis of herbarium specimens along with field studies, Environmental Scanning Electron Microscopy (ESEM), pollen viability and phenology show *C. glaucescens*, *C. jorii* and *C. verrucosa* to be taxonomically distinguishable at the species level. The taxonomy of *Carex* section *Glaucescences* is presented in an artificial dichotomous key to the taxa and in species descriptions.

## DEDICATION

To the beginnings of every Botanist.

## ACKNOWLEDGEMENTS

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## INTRODUCTION

The Cyperaceae (sedge) family consists of 4500 to 5000 species (Reznicek 1990). This large cosmopolitan family contains 70 to 100 genera that are characterized as monocotyledons with linear, three-ranked, alternate or basal, flat or terete or triangular or absent leaves; basal sheaths with connate margins, ligulate or eligulate (Walters & Keil 1996). The family generally has bisexual flowers with highly reduced or absent perianth parts that are subtended by a bract. The two- to three-carpellate ovary matures into an achene (Standley 1981). The flowers are generally wind pollinated, with the exception of the few entomophilous taxa in *Carex*, *Cymophyllus* and *Rhynchospora* in North America.

Four subfamilies in the Cyperaceae are the Mapanioideae C.B. Clarke, Cyperoideae, Sclerioideae C.B. Clarke, and Caricoideae Pax (Jones 1994). Caricoideae is characterized by strictly unisexual florets (Goetghebeur 1985), which are subtended by partially or wholly closed perigynium of prophyllar origin (Blaser 1944). Caricoideae contains one tribe, the Cariceae Kunth ex Dumortier, which is comprised of 5 genera, including *Carex* (Reznicek 1990).

*Carex* comprises the majority of species within the Caricoideae subfamily (Reznicek 1990). *Carex* is separated from other genera in the Caricoideae by a completely closed perigynia and a reduced to absent rachillas. *Carex* species are often dominant in wetlands, pastures, prairies, tundra and the herb layer of temperate forests (Catling et al. 1990). *Carex* contains about 500 (Catling et al. 1990) to 600 (Hermann 1954) species in North America and as many as 2000 worldwide (Catling et al. 1990). The number of North American hybrid taxa reported in *Carex* is 253

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This thesis follows the format of Sida, Contributions to Botany.

(Cayouette & Catling 1992), however 70 hybrid taxa are commonly recognized.

Hybridization within the family has been ignored, partially due to numerous, sometimes erroneous, reports of hybrids in old monographs and floristic lists as well as the difficulty of delimiting hybrid taxa and the differing views of botanists such as Mackenzie and Fernald (Cayouette & Catling 1992). The majority of recent taxonomic research in the Cyperaceae has been in the genus *Carex* (personal communication Jones 2001). Current research indicates the highest propensity for hybridization is in the subgenus *Carex* (Cayouette & Catling 1992).

The three *Carex* subgenera recognized by Reznicek (1990) are *Vignea*, *Carex* and *Indocarex*. Kükenthal (1909) included a fourth subgenus, *Primocarex*, which is considered an artificial grouping (Reznicek 1990). *Vignea* is separated from other subgenera based on the absence of cladoprophylls, presence of an abaxial false suture and lateral inflorescence subunits that are generally sessile (Reznicek 1990). *Carex* and *Indocarex* subgenera are separated from each other based on the presence or absence of inflorescence prophylls and lateral inflorescence subunits (Reznicek 1990). Reznicek (1990) depicts the *Carex* inflorescence with lateral inflorescence units that depending on the subgenus contain spikes (lateral inflorescence subunits). Reznicek (1990) also refers to the single flowered perigynia as a spikelet. Within this manuscript lateral inflorescence subunits will not be further discussed and inflorescence units will be referred to as spikes.

*Carex glaucescens* S. Elliott, *C. jorii* L.H. Bailey, and *C. verrucosa* G.H. Muhlenberg are in section *Glaucescentes* of the genus *Carex* (Reznicek 2001). Based on subgenera characters listed by Reznicek (1990), these taxa are placed in the subgenus *Carex*. *Carex brasiliensis* St. Hilaire is also included in section

*Glaucescens*; however, it is a South American species and is excluded from this study due to logistics and a clear morphological separation from the other three taxa. *Carex brasiliensis* is reported to differ from the North American taxa by having more lateral spikes (9) and flattened perigynia (Boott 1860). These North American taxa are distributed from East Texas to Maryland along the southeast coast of the United States and inland from Arkansas to Tennessee. They occur in cypress-gum ponds and depressions, seasonally wet pine savannas, flatwood ponds, branch bays, adjacent ditches, swales, floodplain forests, bottomland woodlands, marshy shores and often in shallow water (Godfrey & Wooten 1979). Reznicek (2001) described the section *Glaucescens* as follows:

Cespitose in large, leafy clumps, rhizomes short and stout; basal sheaths coriaceous, brown to dull reddish purple. Culms central, 50-150 cm tall. Leaves 5 to 15, M-shaped in cross section, glaucous and  $\pm$  papillose beneath; sheath fronts brownish hyaline, finely brown-dotted. Inflorescences 10-40 cm, with 3 to 8 spikes, terminal spikes staminate, lateral spikes pistillate or the distal 1 or 2 staminate or androgynous, peduncled, erect or pendent; proximal bracts leaf-like, exceeding the inflorescence, sheathless or short-sheathing. Pistillate scales acute to retuse, awned. Perigynia ascending or spreading, narrowly ovoid to broadly obovoid, rounded-trigonal to flattened-elliptic in cross section, strongly to weakly nerved, papillose, tapering or contracted to a short, entire or weakly bidentulate beak, stigmas 3, anthers 3, reddish.

Elliott (1824) described *C. glaucescens* and did not designate a type specimen.

*Carex glaucescens* material examined by Elliott was not located in the literature or herbarium material. Bailey (1886) described *C. jorii* from a Joor collection [Holotype: Louisiana, Comite Swamp, near Baton Rouge, 5 Aug 1885, J. F. Joor s.n. (BH)]. *Carex verrucosa* was listed by Muhlenberg (1813) in a checklist of North America.

Muhlenberg (1817) provided a description for *C. verrucosa*, thus validly publishing the name, but did not designate a type specimen.

Boott (1860) was the first to group *C. brasiliensis* with *C. glaucescens* and *C. verrucosa* at the sectional level. Boott (1860) acknowledged Drejer's 1844 placement of these taxa in *Carex* section *Aeorostachyae*. Kükenthal (1909) placed *C. jorii* in section *Maximae* (Ascherson) Kükenthal and *C. glaucescens*, *C. verrucosa* and *C. brasiliensis* in section *Paludosae* G. Don. Reznicek (2001) disagreed with Kükenthal's placement of the taxa based on their sheathless or short sheathing bracts compared to section *Maximae* and cespitose habit compared to most species in the *Paludosae*. Mackenzie's (1935) North American treatment placed *C. glaucescens*, *C. jorii* and *C. verrucosa* in *Carex* section *Pendulineae*. Mackenzie (1935) included *Carex flacca* in his treatment of the section *Pendulineae*, however Reznicek (2001) reported *C. flacca* as unrelated based on it being smaller, having elongate creeping rhizomes, scabrous hispid perigynia and awnless pistillate scales. Reznicek (2001) placed *C. glaucescens*, *C. jorii*, *C. verrucosa* and *C. brasiliensis* in section *Glaucescentes* because of their papillose perigynia and basal staminate flowers of the pistillate spikes.

Differentiation between species in the *Glaucescentes* section is difficult due to similarities and indistinct delimitations that contribute to the confusion concerning species identification and their distribution. Until recent, the distribution of *C. verrucosa* in Texas was unclear. In Texas, Correll and Johnston (1970) excluded *C. verrucosa*, citing it as erroneously being reported in Texas. Gould (1975) included *C. verrucosa* as occurring in Texas. Godfrey and Wooten (1979) included *C. verrucosa* in their work, but as not occurring in Texas. Bridges & Orzell (1989) reported *C. verrucosa* from several southeastern Texas Counties. Jones et al. (1997) considered *C. verrucosa* as present in Texas from the examination and collection of specimens (personal communication Jones 2001). Standley (2002) also mapped *C. verrucosa* as being

present in Texas. Clarification of the characters that delimit these taxa will increase accuracy in identifications and make distributions easier to define.

*Carex jorii* is separated from *C. glaucescens* and *C. verrucosa* by a pistillate scale that tapers distally into an acuminate to aristate apex and strong veins on the perigynia (Godfrey & Wooten 1979). These characters distinguish *Carex jorii* as a separate species; however it is important to include *C. jorii* in the study because it is in the section *Glaucescentes*. *Carex glaucescens* and *C. verrucosa* are differentiated, in several keys (e.g., Godfrey & Wooten 1979, personal communication Jones 2001) by drooping or erect peduncles, peduncle length, and frequently androgynous versus frequently pistillate spikelets. The degree to which a peduncle is erect or drooping varies and leads to subjective interpretation, thus increasing the difficulty of obtaining accurate identifications. These characters are indistinct between taxa are not desirable for taxonomic keys. Quantitative characters which more effectively separate taxa are preferable.

Objectives of this study were to define character and character states that help delimit the taxa, produce a dichotomous key for identification supported by statistical analysis of morphological data that will increase accuracy in identification of *C. glaucescens*, *C. jorii*, and *C. verrucosa*, determine the nomenclatural rank of the three North American taxa within *Carex* section *Glaucescentes*, examine herbarium, field and type specimens, add to the taxonomic knowledge pertaining to *Carex* and review pertinent taxonomic literature.



## METHODS

Methods for evaluation of taxonomic ranking within *Carex* section

*Glaucescences* concentrated on gross plant morphology. Other methods used to support morphological findings included cytology, micromorphology, phenology and evaluation of taxa distributions. Results from these areas were used to characterize the taxonomy of North American taxa within *Carex* section *Glaucescences*.

Fourteen herbaria loaned a total of 1,495 specimens of *C. glaucescens*, *C. jorii*, and *C. verrucosa* (Table 1) to the S.M. Tracy Herbarium (TAES) for this study. The selected herbaria represent the collective geographic distributions of these taxa. To provide a complete treatment of the North American taxa in *Carex* section *Glaucescences*, it is necessary to locate or designate type specimens for valid taxa. Type specimens were located, when possible, for comparison with type descriptions and borrowed specimens. Taxon descriptions were evaluated for holotype designation, so that an exact specimen could be requested. When no type specimen was designated in the original protologue, taxonomic literature was searched to locate repositories of the publishing author. Requests to those herbaria for material were made.

If no type specimen for a valid taxon was designated at the time of publication, specimens were considered for typification. Selection of specimens for typification were based on rules from Article 9 of the International Code of Botanical Nomenclature (ICBN) (Greuter et al. 2000). Any specimen chosen to serve as a type specimen within this manuscript is not validly published here and must be validly published in accordance with the ICBN (Greuter et al. 2000). If a specimen is selected from original material available to the author at the time of publication, the type specimen would be

TABLE 1. Herbaria lending specimens and specimen images to TAES for use in this study.

LENDING INSTITUTION	ACRONYM <sup>1</sup>	LOANED SHEETS	IMAGED SHEETS	STATES
L.H. Bailey Hortorium	BH*	1	0	NY
Herbarium of Charles T. Bryson	CTB <sup>1</sup>	146	0	MS
Duke University	DUKE	147	0	NC
University of Florida	FLAS	100	0	FL
Florida State University	FSU	45	0	FL
University of Michigan	MICH	142	0	MI
North Carolina State University	NCSC	52	0	NC
University of North Carolina	NCU	215	0	NC
University of Louisiana at Monroe	NLU	241	0	LA
New York Botanical Garden	NY***	11	1	NY
The Academy of Natural Sciences Herbarium	PH**	0	2	PA
Southern Forest Research Program	SFRP	23	0	LA
Southern Weed Science Research Center	SWSL	6	0	MS
S. M. Tracy Herbarium	TAES	55	0	TX
University of Texas - Austin	TEX/LL	101	0	TX
University of Alabama	UNA	50	0	AL
U.S. National Herbarium	US	160	0	DC
TOTALS		1495	3	

\* Holotype of *Carex jorii* Bailey.

\*\* Elliott 215, *Carex verrucosa* Muhl.

\*\*\* Imaged sheet is included in loaned sheets.

<sup>1</sup> Acronyms follow Holmgren et al. (1990), excluding the personal herbarium of Charles T. Bryson (CTB).

considered a lectotype (Greuter et al. 2000). A neotype would be designated, if all of the original material on which the name of the taxon was based was missing (Greuter et al. 2000). Recommendations from the ICBN for typification include considering not all material in the author's herbarium was used to describe the taxon, exclusion of unscientific methods of purely selecting specimens based on collector or citation in the protologue and a critical knowledge of the taxon.

Literature concerning the family, genus, section and species was assessed for information with reference to taxon description, synonyms and characters. Synonyms for the currently accepted taxa were identified and their descriptions evaluated for distinguishing characters, taxon validity and character overlap. Specifically, characters were selected from taxa descriptions, keys and papers on *Carex* morphology including Bailey (1886), Boot (1860), Elliott (1824), Godfrey & Wooten (1979), Ford & Ball (1992), Mackenzie (1935), Muhlenberg (1817), Reznicek (2001), and Toivonen (1980). Subsequently, this led to a compilation of quantitative and qualitative characters (Table 2) for measurement and quantification from herbarium specimens. The data compiled from the 59 characters and selected herbarium specimens was used in the univariate and multivariate analysis.

The study initially included 60 herbarium specimens from each taxon. Random selection was constrained by ensuring that the selected specimens represented the geographic distribution for each taxon. Selected specimens were not measured if spikes were missing or inaccessibly mounted. Misidentified specimens were recoded prior to or during Principal Component Analysis (PCA) as the accepted taxon based on Jones (personal communication). Measured immature specimens were removed from the PCA. The coherence to these methods resulted in an unequal number of

TABLE 2. Characters used in the analysis. Character states are: continuous quantitative (CQ), two-state quantitative (TS), two-state qualitative (TQ) and multiple-state qualitative (MS).

CHARACTERS	SYMBOL	UNIT OF MEASURE	CHARACTER STATE
INFLORESCENCE			
Inflorescence sex	IS	—	MS
Inflorescence length from base of lowermost sheath to apex of terminal spike	IL	cm	CQ
(Terminal spike)			
Length of spike <sup>a</sup>	SLS	cm	CQ
Length of peduncle from base of uppermost terminal spike to base of sheath of uppermost pistillate spike	SLP	cm	CQ
Length from base of scale to awn apex, removed from the upper portion of the lower fourth of terminal spike	SLSC	mm	CQ
Widest point of scale removed from the upper portion of the lower fourth of the terminal spike	SWSC	mm	CQ
Width of spike at middle point	SWSP	mm	CQ
Spike sex	SS	—	MS
Length of staminate portions of terminal spike	SLSP	mm	CQ
Length of pistillate portions of terminal spike	SLPP	mm	CQ
(Lateral pistillate spikes)			
Length of second lowermost pistillate spike peduncle <sup>b</sup>	PLLP	cm	CQ
Length of uppermost pistillate spike peduncle <sup>b</sup>	PLUP	cm	CQ
Ratio of PLLP to PLUP	PRP	—	CQ
Length of second lowermost pistillate spike <sup>a</sup>	PLLS	cm	CQ
Second lowermost pistillate spike staminate portion; located proximally, distally, both or absent	PSLS	—	MS
Length of uppermost pistillate spike peduncle <sup>b</sup>	PLUS	cm	CQ
Uppermost pistillate spike staminate; portion; located proximally, distally, both or absent	PSUS	—	MS
Ratio PLLS to PLUS	PRS	—	CQ
Average length of uppermost and second lowermost peduncles	PALP	cm	CQ
Average length of uppermost and second lowermost spikes	PALS	cm	CQ
Number of lateral spikes, excluding reduced spikes	PN	—	CQ
Number of lateral spikes with staminate spikelets	PNS	—	CQ
Pistillate scale apex shape	PHC	—	MS
Length from base of scale to awn apex, randomly removed from lateral spikes	PLC	mm	CQ
Widest point of pistillate scale, randomly removed from lateral spikes	PWC	mm	CQ
Length of pistillate scale awn, randomly removed from lateral	PLCW	mm	CQ

TABLE 2. Continued.

CHARACTERS	SYMBOL	UNIT OF MEASURE	CHARACTER STATE
(Low ermost lateral spike)			
Length of spike <sup>a</sup>	LLS	cm	CQ
Length of peduncle <sup>b</sup>	LLP	cm	CQ
Peduncle pendulous or erect	LDE	—	TQ
Length of distal staminate portion	LLSD	mm	CQ
Staminate spikelets present or absent	LS	—	TQ
Staminate spikelets located proximally, distally, both or absent	LSPDB	—	MS
(Reduced spike) <sup>c</sup>			
Number of achenes	RNA	—	CQ
Spike sex	RSS	—	MS
Number of reduced spikes	RNS	—	CQ
Length of reduced spike <sup>a</sup>	RLS	mm	CQ
Spike pedunculate or sessile	RPSS	—	TS
Length to base of terminal spike	RLBS	mm	CQ
(Achene)			
Length of achene from base above possible residual stipitate portion of perigynia to apex of style	ALWS	mm	CQ
Length of achene from base above possible residual stipitate portion of perigynia to base of style	ALWOS	mm	CQ
Widest point of achene	AW	mm	MS
Ratio of ALWOS to AW	ARLW	—	CQ
Length of style	ALS	mm	CQ
Achene surface texture	AT	—	MS
Achene shape	AS	—	MS
Location of widest point of achene	AWP	—	MS
(Perigynia)			
Length of perigynia from base of stipitate base to apex of beak	GL	mm	CQ
Length of perigynia beak from abrupt angle change between beak and body of perigynia to apex of beak	GLBK	mm	CQ
Basal shape of perigynia	GHBS	—	MS
Number of nerves on perigynia faces	GNN	—	CQ
Nerves conspicuous or inconspicuous	GCI	—	TQ
Perigynia shape	GH	—	MS
Width of perigynia stipitate base	GWB	mm	CQ
Length of perigynia stipitate base	GLB	mm	CQ
Perigynia texture	GT	—	MS
Widest point of perigynia	GW	mm	CQ
Location of widest point of perigynia	GWP	—	MS
Perigynia color	GC	—	MS

TABLE 2. Continued.

CHARACTERS	SYMBOL	UNIT OF MEASURE	CHARACTER STATE
PHENOLOGY			
Collection date	FD	—	CQ
VEGETATIVE			
Culm length from apex of rootstock to apex of inflorescence	HP	cm	CQ
Sheath auricle shape	AURI	—	MS

- a spike length was measured from the apex of the uppermost scale to the base of the lowest scale  
b peduncle length is measured from base of spike, denoted by lowest scale, to base of spike sheath  
c reduced spikes were considered those at the base of the terminal spike with less than 5 achenes

specimens available for study for each taxon. The 117 herbarium specimens or Operational Taxonomic Units (OTU's) used for data analysis are listed in Table 3.

#### Field Work

Location data from herbarium labels aided in the location of populations for field studies. Specimens were collected from 16 populations (Table 4). Research in the field centered on plant habit and taxon variation. Characters evaluated during field collection included exposure to the sun, orientation of lowermost pistillate spike peduncle and soil type.

A representative soil description was recorded at one site for each taxon (Table 5). Soil descriptions were recorded to aid in habitat description. Soil characteristics and sampling procedures were taken from Schoeneberger et al. (2002). The characters of depth, matrix color, texture, structure, redoximorphic depletion characters, iron or manganese concentration characters and iron and manganese concretions within upper horizons were recorded.

#### Cytology

##### *Chromosome Numbers*

Mitotic and meiotic chromosome counts were attempted. Mitotic counts were made from greenhouse grown specimens. Seeds obtained from field collections successfully germinated after removing a portion of the seed coat. Root tips collected from the greenhouse grown specimens were pretreated with alpha-bromonaphthalene to increase the frequency of cell divisions in metaphase and to aid in accurately counting chromosomes. Specimens were rinsed with distilled water (d. H<sub>2</sub>O) and fixed

TABLE 3. Specimens used in the numerical analysis and scanning electron microscopy.

TAXA CODE	HERBARIUM AND ACCESSION NUMBER	COLLECTOR	COLLECTOR NUMBER	STATE	COUNTY	COLLECTION DATE	
g01	CTB	—	<u>J.R. MacDonald</u>	<u>2673</u>	AL	Houston	23 Jun 1991
g02	CTB	—	<u>J.R. MacDonald</u>	<u>2959</u>	AL	Houston	3 Aug 1991
g03	CTB	—	<u>R.F.C. Naczi</u>	<u>3305</u>	AL	De Kalb	2 Aug 1993
g04	CTB	—	<u>R. Kral</u>	<u>51209</u>	AL	Pike	10 Jun 1955
g05	CTB	—	<u>R. Kral</u>	<u>81288</u>	AL	Bibb	15 Aug 1994
g06	CTB	—	<u>C.T. Bryson</u>	<u>10147</u>	MS	Low ndes	12 Aug 1990
g07	CTB	—	<u>C.T. Bryson</u>	<u>14943</u>	MS	Greene	25 Jul 1995
g08	CTB	—	<u>S. McDaniel</u>	<u>26438</u>	MS	Lamar	28 Aug 1982
g09	CTB	—	<u>S. McDaniel</u>	<u>28031</u>	MS	Hancock	25 Aug 1984
g10	DUKE	365024	<u>J.W. Horn</u>	<u>135</u>	GA	McIntosh	20 Aug 1997
g11	DUKE	164377	<u>D. Demaree</u>	<u>32583</u>	MS	Stone	5 Aug 1952
g12	DUKE	41857	<u>H.T. O'Neil</u>	<u>8506</u>	MS	Jackson	15 Sep 1937
g13	DUKE	99627	<u>A.B. Seymour</u>	<u>91828</u>	MS	Jackson	14 Aug 1989
g14	DUKE	363728	<u>J.W. Horn</u>	<u>1400</u>	NC	Moore	21 Aug 1998
g15	DUKE	14116	<u>W.B. Fox</u>	<u>2812</u>	NC	Bertie	23 Jul 1949
g18	FLAS	131823	<u>K.D. Perkins</u>	<u>360</u>	FL	Okaloosa	24 Jul 1977
g22	MICH	—	<u>R. Kral</u>	<u>85572</u>	AL	Grenshaw	20 Jul 1990
g23	MICH	—	<u>H.H. Bartlett</u>	<u>1031</u>	GA	McDuffe	25 Aug 1907
g24	MICH	—	<u>J.B. Nelson</u>	<u>13281</u>	SC	Clarendon	21 Jul 1992
g25	MICH	—	<u>J.B. Nelson</u>	<u>14748</u>	SC	Sumter	20 Aug 1993
g26	MICH	—	<u>S.D. Jones</u>	<u>10393</u>	TX	New ton	4 Nov 1993
g27	NCU	6575	<u>J.R. Bozeman</u>	<u>5812</u>	GA	Tattnall	12 Jul 1966
g30	NCU	6565	<u>J.R. Bozeman</u>	<u>10549</u>	GA	Charlton	26 Jun 1967
g31	NCU	6578	<u>J.W. Thieret</u>	<u>36143</u>	LA	Livingston	5 Aug 1958
g32	NCU	6584	<u>K.E. Rogers</u>	<u>1372</u>	MS	Forrest	29 Aug 1969
g34	NCU	6393	<u>T.D. Nifong</u>	<u>597</u>	NC	Robeson	23 Sep 1994
g35	NCU	6458	<u>H.E. Ahles</u>	<u>33293</u>	NC	Bladen	7 Aug 1957
g36	NCU	6469	<u>H.E. Ahles</u>	<u>35790</u>	NC	Duplin	7 Nov 1971
g37	NCU	6504	<u>A.E. Radford</u>	<u>38840</u>	NC	Washington	6 Aug 1958
g38	NCU	6474	<u>H.E. Ahles</u>	<u>48285</u>	NC	Gates	21 May 1949
g42	NLU	207716	<u>K.H. Kessler</u>	<u>2381</u>	LA	Winn	6 Aug 1982
g44	NLU	377787	<u>C. Allen</u>	<u>18200</u>	LA	Washington	14 Aug 1995
g45	NLU	377785	<u>C. Allen</u>	<u>18202</u>	LA	St. Tammany	15 Aug 1995
g46	NLU	191283	<u>D. Thomas</u>	<u>78118</u>	LA	Vernon	30 May 1992
g48	TAES	207534	<u>R. Carter</u>	<u>10347</u>	GA	Liberty	12 Aug 1992
g50	TAES	188980	<u>S.&amp;G. Jones</u>	<u>3707</u>	TX	Tyler	17 Aug 1989
g51	TAES	159349	<u>J.W. Kessler</u>	<u>4958</u>	TX	New ton	22 Aug 1981
g52	USNAT	817421	<u>J.D.S.</u>	<u>s.n.</u>	AL	Butler	21 Aug
g53	USNAT	2310281	<u>D. Demaree</u>	<u>32512</u>	MS	Harrison	4 Aug 1952
g54	USNAT	1838039	<u>R.K. Godfrey</u>	<u>1340</u>	SC	Lexington	8 Aug 1939
g55	USNAT	3306756	<u>C.N. Horn</u>	<u>5436</u>	SC	Richland	21 Jul 1992
g58	UT	—	<u>S.L. Orzell</u>	<u>8306</u>	TX	Angelina	26 Aug 1988
g61	NCU	6418	<u>A. McCrary</u>	<u>717</u>	NC	New Hanover	27 Jul 1963
g62	TEX-LL	—	<u>S.L. Orzell</u>	<u>12484</u>	FL	Santa Rosa	21 Sep 1989
g63	TEX-LL	—	<u>S.L. Orzell</u>	<u>12502</u>	FL	Okaloosa	21 Sep 1989
g64	CTB	—	<u>R. Carter</u>	<u>10347</u>	GA	Liberty	12 Aug 1992
g66	CTB	—	<u>C.T. Bryson</u>	<u>14554</u>	MS	Jackson	16 Oct 1994
j01	DUKE	375748	<u>J.R. MacDonald</u>	<u>13294</u>	AL	Geneva	26 Jul 1999



TABLE 3. Continued.

TAXA CODE	HERBARIUM AND ACCESSION NUMBER	COLLECTOR	COLLECTOR NUMBER	STATE	COUNTY	COLLECTION DATE	
j02	DUKE	160162	<u>R. Kral</u>	<u>1638</u>	FL	Gadsden	14 Oct 1955
j03	DUKE	365151	<u>M.H. Alford</u>	<u>1470</u>	MS	Amite	12 Sep 1998
j04	FLAS	195370	<u>S.L. Orzell</u>	<u>20304</u>	AL	Houston	1 Aug 1992
j05	FLAS	156912	<u>J.R. Burkhalter</u>	<u>9724</u>	FL	Santa Rosa	17 Oct 1984
j06	FLAS	195371	<u>S.L. Orzell</u>	<u>20026</u>	FL	Lafayette	7 Jul 1992
j07	LL	—	<u>A.E. Radford</u>	<u>44342</u>	NC	Nash	8 Sep 1975
j09	LL	—	<u>F. R. Waller, Jr.</u>	<u>256</u>	TX	Nacogdoches	11 Aug 1964
j10	LL	293013	<u>R.S. Mitchell</u>	<u>3958</u>	TX	Harrison	25 Aug 1967
j11	LL	293011	<u>S.L. Orzell</u>	<u>11373</u>	TX	Wood	5 Aug 1989
j12	LL	—	<u>C. L. Lundell</u>	<u>14092</u>	TX	Newton	18 Jul 1945
j13	LL	293017	<u>B. C. Tharp</u>	<u>43268</u>	TX	Angelina	9 Sep 1961
j14	LL	—	<u>V. L. Cory</u>	<u>54744</u>	TX	Jasper	2 Aug 1961
j16	NCU	6411	<u>L. S. Beard</u>	<u>1423</u>	NC	Lee	23 Aug 1956
j17	NCU	6407	<u>L.J. Musselman</u>	<u>4113</u>	NC	Gates	17 Oct 1970
j18	NCU	6410	<u>A.E. Radford</u>	<u>27811</u>	NC	Johnston	7 Aug 1957
j20	NCU	6423	<u>A.E. Radford</u>	<u>39197</u>	NC	Tyrrell	19 May 1974
j22	NCU	6434	<u>S. W. Leonard</u>	<u>1938</u>	SC	Jasper	9 Sep 1968
j23	NCU	6438	<u>A.E. Radford</u>	<u>28365</u>	SC	Williamsburg	22 Aug 1957
j24	NLU	128639	<u>P. Laird</u>	<u>728</u>	LA	Quachita	4 Aug 1974
j25	NLU	154800	<u>S.E. Schutz</u>	<u>1767</u>	LA	Rapides	6 Oct 1978
j26	NLU	227676	<u>D.C. Moore</u>	<u>2520</u>	LA	Union	4 Sep 1983
j27	NLU	340667	<u>C.T. Bryson</u>	<u>10847</u>	LA	St. Tammany	26 Aug 1991
j29	NLU	120446	<u>R. D. Thomas</u>	<u>46104</u>	LA	Quachita	1 Aug 1958
j30	NLU	294873	<u>M.W. Palmer</u>	<u>889</u>	NC	Durham	1 Aug 2386
j31	NLU	7943	<u>R. D. Thomas</u>	<u>39724</u>	TX	Liberty	23 Sep 1958
j32	TAES	207540	<u>R. Carter</u>	<u>10268</u>	GA	Bryan	4 Aug 1992
j33**	TAES	5700	<u>S.M. Tracy</u>	<u>8475</u>	MS	—	1 Aug 1903
j34	TAES	100028	<u>F.R. Waller</u>	<u>256</u>	TX	Nacogdoches	11 Aug 1964
j35	TAES	140005	<u>D. Hartman</u>	<u>1274</u>	TX	San Jacinto	27 Aug 1973
j36	TAES	202165	<u>S.&amp;G. Jones</u>	<u>3632</u>	TX	Marion	5 Aug 1989
j37	TAES	202162	<u>S.&amp;G. Jones</u>	<u>3656</u>	TX	Harrison	5 Aug 1989
j38	TAES	159345	<u>J.W. Kessler</u>	<u>5100</u>	TX	Harris	18 Sep 1981
j39	UNA	959	<u>R.R. Haynes</u>	<u>6915</u>	AL	Macon	24 Aug 1978
j40	DUKE	15388	<u>H.L.B.</u>	<u>5474</u>	NC	Harnett	16 Aug 1932
v01	CTB	—	<u>J.R. MacDonald</u>	<u>5569</u>	AL	Houston	19 Sep 1992
v02	CTB	—	<u>J.R. MacDonald</u>	<u>5524</u>	AL	Houston	12 Sep 1992
v03	CTB	—	<u>J.R. MacDonald</u>	<u>5133</u>	AL	Houston	22 Jul 1992
v04	CTB	—	<u>J.R. MacDonald</u>	<u>4</u>	AL	Geneva	11 May 1978
v05	CTB	—	<u>R. Kral</u>	<u>80797</u>	GA	Dougherty	9 Aug 1992
v08	CTB	—	<u>C.T. Bryson</u>	<u>8021</u>	GA	Lowndes	9 Jul 1988
v10	CTB	—	<u>C.T. Bryson</u>	<u>4172</u>	MS	Jackson	15 Apr 1986
v11	DUKE	65850	<u>F.S. Earle</u>	<u>82</u>	MS	Jackson	19 Jul 1889
v13	DUKE	137839	<u>R.K. Godfrey</u>	<u>53496</u>	FL	Leon	27 Mar 1956
v15	DUKE	205118	<u>J.W. Thieret</u>	<u>25921</u>	LA	St. Tammany	29 Apr 1967
v16	FSU	166923	<u>L.C. Anderson</u>	<u>6703</u>	FL	Franklin	24 Oct 1983
v17	FSU	164600	<u>L.C. Anderson</u>	<u>6327</u>	FL	Gadsden	25 Apr 1983

TABLE 3. Continued.

TAXA CODE	HERBARIUM AND ACCESSION NUMBER	COLLECTOR	COLLECTOR NUMBER	STATE	COUNTY	COLLECTION DATE	
v19	FSU	146481	<u>S.W. Leonard</u>	<u>6256</u>	FL	Leon	7 Apr 1976
v20	FSU	182708	<u>L.C. Anderson</u>	<u>12776</u>	FL	Walton	24 May 1990
v21	FSU	176694	<u>L.C. Anderson</u>	<u>10359</u>	FL	Taylor	16 Apr 1987
v22	FSU	98176	<u>S. McDaniel</u>	<u>6146</u>	FL	Liberty	4 May 1965
v23	FSU	179695	<u>J.R. Burkhalter</u>	<u>10596</u>	FL	Escambia	8 May 1987
v24	FSU	76329	<u>G.R. Cooley</u>	<u>8240</u>	FL	Polk	3 May 1961
v25	FSU	144878	<u>D.L. Martin</u>	<u>831</u>	FL	Marion	26 Sep 1975
v26	NCU	6441	<u>J.A. Lassiter</u>	<u>25076</u>	FL	Hillsborough	20 May 1962
v27	NCU	6391	<u>A.E. Radford</u>	<u>4274</u>	NC	New Hanover	21 May 1949
v28	NCU	6395	<u>H.E. Ahles</u>	<u>12547</u>	SC	Allendale	13 May 1956
v29	NLU	351219	<u>K. Northrup</u>	<u>s.n.</u>	TX	Harris	26 Apr 1992
v30	UNA	1607	<u>B. Hansen</u>	<u>7077</u>	FL	Highlands	17 Apr 1980
v31	US	3167473	<u>J.B. McFarlin</u>	<u>5807</u>	FL	Park	15 Jun 1931
v32	US	3163427	<u>J.B. McFarlin</u>	<u>5662</u>	FL	Osceola	5 Jun 1931
v33	US	3163426	<u>D.S. Correll</u>	<u>5310</u>	SC	Georgetow n	14 Jun 1936
v35	TEX-LL	—	<u>S.L. Orzell</u>	<u>5787</u>	LA	Allen	22 Sep 1987
v36	TEX-LL	293407	<u>S.L. Orzell</u>	<u>5809</u>	TX	San Jacinto	25 Sep 1987
v37	TEX-LL	293409	<u>S.L. Orzell</u>	<u>5406</u>	TX	Chambers	1 Jun 1987
v38	TEX-LL	293408	<u>S.L. Orzell</u>	<u>8230</u>	TX	New ton	23 Aug 1988
v39	TEX-LL	—	<u>S.L. Orzell</u>	<u>8567</u>	TX	New ton	26 Sep 1988
v40	TEX-LL	—	<u>R.K. Godfrey</u>	<u>49189</u>	NC	Brunsw ick	21 May 1949
v41	TAES	222971	<u>L. Clayton</u>	<u>13</u>	TX	Montgomery	16 Oct 1998
v42	FLAS	175458	<u>W.T. Scudder</u>	<u>1687</u>	FL	Volusia	27 Apr 1985
477*	TAES	—	<u>D.C. McLaughlin</u>	<u>477</u>	TX	Hardin	18 Jul 2002
522B*	TAES	—	<u>D.C. McLaughlin</u>	<u>522B</u>	TX	New ton	28 Jul 2002
511*	TAES	—	<u>D.C. McLaughlin</u>	<u>511</u>	TX	Orange	28 Jul 2002
521*	TAES	—	<u>D.C. McLaughlin</u>	<u>521</u>	TX	New ton	28 Jul 2002
518*	TAES	—	<u>D.C. McLaughlin</u>	<u>518</u>	TX	New ton	28 Jul 2002
508*	TAES	—	<u>D.C. McLaughlin</u>	<u>508</u>	TX	New ton	28 Jul 2002
436C*	TAES	—	<u>D.C. McLaughlin</u>	<u>436C</u>	TX	Tyler	24 May 2002
416B*	TAES	—	<u>D.C. McLaughlin</u>	<u>416B</u>	TX	Montgomery	22 May 2002
492*	TAES	—	<u>D.C. McLaughlin</u>	<u>492</u>	TX	Montgomery	25 Jul 2002
512****	TAES	—	<u>D.C. McLaughlin</u>	<u>512</u>	TX	Orange	28 Jul 2002
1612***	TAES	236082	<u>D. Rosen</u>	<u>1612</u>	TX	Montgomery	21 Aug 2001
71***	TEX-LL	—	<u>S.L. Orzell</u>	<u>7104</u>	LA	Calcasieu	30 May 1988
151***	TAES	234901	<u>H. Amestoy</u>	<u>151</u>	TX	Montgomery	16 Oct 1998
KN***	TAES	209301	<u>K. Northrup</u>	<u>s.n.</u>	TX	Harris	26 Apr 1992

\* Personal collections used in both numerical analysis and environmental scanning electron microscopy.

\*\* Borrowed specimens used in both numerical analysis and environmental scanning electron microscopy.

\*\*\* Borrowed specimens used only in environmental scanning electron microscopy.

\*\*\*\* Personal collections used in both numerical analysis and environmental scanning electron microscopy.

TABLE 4. Personal collections of *Carex* section *Glaucescentes*. Specimens used in PCA and those that are possible hybrid taxa are indicated.

LOCATION	TEXAS County	TAXA AND COLLECTION NUMBERS**		
		<i>verrucosa</i>	<i>glaucescens</i>	<i>joorii</i>
1 E side of Hwy 69, 5.9 mi S of Village Creek, near Kountze	Hardin		<u>477*, 478*, 479</u>	
2 E side of Hwy 2798, N of Votaw, 3.1 mi S of intersection w/ FR 943	Hardin		<u>504*</u>	
3 E side of 69, N of Kountze, 1.8 mi N of intersection w/ 420, 0.2 mi N of Village Creek	Hardin		<u>754A*,B*,C*, D*,E*</u>	
4 Hwy 362, 1m SW of intersection w/ FR 1293	Hardin		<u>755A*,B*</u>	
5 E side of Hwy 943, 4.7 mi W of intersection w/ 1003 and 7.5 mi SE of Polk Co line	Hardin	<u>650A,B,C,D</u>		
6 W side of Hwy 62, 1.8 mi N of Orange Co line, N of Vidor, near Texla	Jasper		<u>507A*,B</u>	
7 N side of Hwy 105, 1.8 mi W of Peach Creek	Montgomery	<u>407, 834*</u>	<u>473, 475*, 491*, 492*, 493, 494, 495*</u>	<u>471<sup>h</sup>, 472<sup>h</sup>, 835<sup>h*</sup>, 836<sup>h</sup>, 837<sup>h*</sup>, 838<sup>h*</sup></u>
8 N side of Hwy 105, 1.0 mile E of Peach Creek***	Montgomery		<u>347A*,B,C*, 416A*,B*, 476, 831*, 832*, 833*</u>	
9 W side of Hwy 62, .5 mi N of Orange Co line near Texla	Newton			<u>508*</u>

TABLE 4. Continued.

LOCATION	TEXAS County	TAXA AND COLLECTION NUMBERS**		
		<i>verrucosa</i>	<i>glaucescens</i>	<i>joorii</i>
10 S side of Hwy 12, 1.9 mi N of Orange Co line	Newton		<u>517</u>	<u>518*, 519*, 520</u>
11 W side of Hwy 87, 2.2 mi S of Slaydon Creek, 4.8 mi S of intersection w/ Hwy 2829	Newton		<u>521*</u>	
12 E side of Hwy 87, 3.6 mi S of intersection w/ Hwy 2829, small depression extending from ditch, near Bleakwood	Newton		<u>522A*,B*</u>	
13 E side of Hwy 87, 0.45 mi S of intersection w/ Hwy 2829, near Bleakwood	Newton		<u>523</u>	
14 N side of Hwy 12, 3.7 mi NE of intersection w/ 62, under Myrica and Styra shrubs, 1.0 mi SW of Newton Co line	Orange		<u>511*, 513</u>	<u>510*, 512</u>
15 N side of 943, several mi W of Hardin Co line	Polk		<u>505</u>	
16 FM 92, 7.9 mi N of intersection with FM 1112	Tyler	<u>436A*,B,C*, 649A,B,C,D</u>		

\* Specimens used in PCA of personal collections and loaned herbarium specimens.

\*\* Numbering procedures were not consistent, no inferences can be made from specimen numbering.

\*\*\* Plants present, with vegetative characteristics of *C. glaucescens* and *C. joorii*, but not in flower.

h Specimens considered as probable hybrid taxa.

TABLE 5. Representative soil descriptions of *Carex* section *Glaucescens* from personal collections.

<b><i>Carex glaucescens</i></b>						
Depth (in)***	Horizon	Matrix Color	Texture, Structure	Redoximorphic Depletions %,Sz, Cn, Col, Location	Fe or Mn Concentrations %,Sz, Cn, Col, Location	FeMn Concretions
0 - 2	A	10YR4/2	L, SBK	—	—	—
2 - 8	B	10YR5/3	L, SBK	F, F, P, 10YR 6/3, w ithin peds	Fe: F, F, F, 10YR5/4, root channels & w ithin peds	Present
8 - 16	B	10YR5/3	SIL, Moderate SBK	F, —, —, 10YR6/3, clay depletions w ithin peds & on ped surfaces	Fe: M, F, F, 10YR6/6, w ithin peds	Few , Fine

Original surface horizon has been removed.

Location: Texas. Polk Co., on E side of Hwy 2798, N of Votaw , 3.1 miles S of intersection w ith FR 943.

***Carex joorii***

Depth (in)***	Horizon	Matrix Color	Texture, Structure	Redoximorphic Depletions %,Sz, Cn, Col, Location	Fe or Mn Concentrations %,Sz, Cn, Col, Location	FeMn Concretions
0 - 1 1/2	Oi*	—	—, —	—	—	—
1 1/2 - 8 1/2	A	2.5Y3/2	SICL, SBK	C, —, —, 10YR6/2, —	Fe: F, F, —, 10YR5/6, root channels & w ithin peds Fe: —, —, —, 2.5Y4/6, almost an amorphous Fe oxide	—
8 1/2 - 16	Eg	10YR5/2	SICL, SBK	—, —, —, 10YR6/2, clay depletions w ithin peds	Fe: C, F, D, 2.5Y6/6, root channels	—

Location: Texas. New ton Co., on W side of Hwy 62, 0.5 miles N of Orange Co., line near Texla.

TABLE 5. Continued.

<b><i>Carex verrucosa</i></b>						
Depth (in)***	Horizon	Matrix Color	Texture, Structure	Redoximorphic Depletions %,Sz, Cn, Col, Location	Fe or Mn Concentrations %,Sz, Cn, Col, Location	FeMn Concretions
0 - 1 1/2	Overburden	Rubbed & Moist 10YR6/1 Dry 10YR3/1	L, —	Hydrophobic, waxy material on soil particles	—	—
1 1/2 - 4	Oe**	10YR2/1	Peat, —	—	—	—
4 - 10	A	10YR2/1	SIL, SBK	C, —, —, 10YR6/1	—	—
10 - 15 +	Eg	10YR5/2	L, SBK	C, —, —, 10YR6/2, clay depletions on ped faces	Fe: F, —, —, 2.5Y6/6, root channels	—

Location: Texas. Hardin Co., on Hwy 943, E side, 4.7 miles W of intersection w / FM 1003 and 7.5 miles SE of Polk Co. line.

***C. glaucescens, C. joorii, C. verrucosa***

Depth (in)***	Horizon	Matrix Color	Texture, Structure	Redoximorphic Depletions %,Sz, Cn, Col, Location	Fe or Mn Concentrations %,Sz, Cn, Col, Location	FeMn Concretions
0 - 1	A	7.5YR6/1	SIL, —	—	—	—
1 - 5	B	7.5YR5/2	CL, —	—	Fe: F, F, F, 7.5YR6/6, root channels & within	—
5 +	B	7.5YR6/1	CL, —	—	Fe: M, M, D, 7.5YR5/6, root channels & within peds	—

Location: Texas. Montgomery Co., on Hwy 105, 1.8 miles W of Peach Creek.

% Percent of area covered.

Sz Size.

Cn Contrast.

Col Color.

\* Fimbric: poorly decomposed.

\*\* Hemic: intermediately decomposed.

\*\*\* Recorded soil characteristics and abbreviations were taken from Schoeneberger et al. (2002).

with ethanol/glacial acetic acid (3:1). Enzymatic hydrolysis was used to digest cell walls and aid in chromosome spread. Prior to enzymatic hydrolysis specimens were rinsed four times and soaked for 30 minutes with d. H<sub>2</sub>O, acidified with 0.1 N HCL for 10 minutes and rinsed twice and soaked for five minutes with d. H<sub>2</sub>O. Root tips were dissected behind the meristem and placed in 0.5 ml microfuge tubes. The tubes also included the addition of a digestion enzyme mixture of 5% cellulose (Onazuka R-10, Yakult Honshu Co. Ltd., Tokyo, Japan) and 1% pectolyase Y-23 (Seichen Corp., Tokyo, Japan) prepared in citrate buffer (0.05, pH 4.5). Cell walls were digested away for an experimental series of 20-45 minutes at 68°C. Root tips were washed with d. H<sub>2</sub>O four times, and single root tips were transferred to slides. Excess water was removed from around the root tip before adding one drop of fixative and macerating the material. Excess material was rinsed away with fixative after cells attached to the slide. Specimens were viewed either with no stain and phase contrast microscopy or stained with azure B and viewed with brightfield microscopy.

Meiotic chromosome counts were attempted from inflorescences fixed with ethanol/glacial acetic acid (3:1) in the field. Greenhouse grown specimens for mitotic counts never reached flowering stage and were not used. Anthers were removed from the fixative, washed and soaked in d. H<sub>2</sub>O and transferred to a glass slide. Before macerating the anthers, aceto-carmine stain was added. Anthers were heated to aid in stain absorption by the chromatin. Before viewing, the cover slip was pressed down to spread cells and cell contents.

#### *Pollen Stainability*

Pollen stainability is one way to evaluate sexual reproduction and potential pollen viability. Mature pollen grains from each taxon, including suspected hybrids,

were hydrated in d. H<sub>2</sub>O and stained with Iodine-Potassium Iodide (IKI). Based on a count of 100 pollen grains per slide for each specimen, percent potential pollen viability was recorded for each taxon. Specimens used for pollen counts are listed in Table 6. Only pollen with intact exines were counted. Pollen staining a dark brown color tested positive for starch, which indicated the number of mature, viable pollen grains, since mature, viable pollen grains contain starch. Specimens were compared to Toivonen's (1980) research with infertile pollen grains of hybrid *Carex* specimens.

#### Achene and Perigynia Micromorphology

Achene and perigynia micromorphology were examined using Environmental Scanning Electron Microscopy (ESEM). The five specimens from each taxon used for ESEM (Table 3) were separated into the currently accepted taxa, based on Jones (personal communication). The protocol for laboratory procedures were used by Schuyler (1971), Menapace et al. (1986), Standley (1987a), Standley (1987b), Wujek and Menapace (1986), Catling et al. (1989) and Saarela and Ford (2001). The specimens, mounted on Scanning Electron Microscopy (SEM) stubs with double sided tape, were examined using an Electroscan E-3 Environmental Scanning Electron Microscope at an accelerating voltage of 30 KV.

#### Data Analysis

##### *Univariate Analysis*

The data matrix contained measurements and observations for 59 characters and 147 OTU's. Continuous variables were evaluated based on mean, standard deviation, variance, skewness and range using Statistical Package for Social Sciences 11.0 (SPSS). Continuous and discrete variables were evaluated graphically with box



TABLE 6. Pollen stainability. Percent viability was recorded for specimens; pollen staining a dark color were recorded as viable.

TAXA	PERCENT VIABILITY	POPULATION NUMBER*	POPULATION IS KNOWN TO CONTAIN**	PERSONAL COLLECTION NUMBER*
<i>C. glaucescens</i>	60%	14	G,J	<u>511</u>
<i>C. glaucescens</i>	74%	1	G	<u>477</u>
<i>C. glaucescens</i>	75%	12	G	<u>522B</u>
<i>C. glaucescens</i>	84%	7	G, J, V	<u>492</u>
Hybrid	3%	7	G, J, V	<u>837</u>
Hybrid	25%	7	G, J, V	<u>835</u>
Hybrid	30%	7	G, J, V	<u>838</u>
Hybrid	31%	7	G, J, V	<u>473</u>
<i>C. jorii</i>	14%	9	J	<u>508</u>
<i>C. jorii</i>	80%	14	G, J	<u>510</u>
<i>C. verrucosa</i>	5%	7	G, J, V	<u>834</u>
<i>C. verrucosa</i>	9%	16	V	<u>649D</u>
<i>C. verrucosa</i>	11%	5	V	<u>650C</u>
<i>C. verrucosa</i>	91%	8	V	<u>831</u>

\* Location data are presented in Table 4.

\*\* *Carex glaucescens* (G), *C. jorii* (J), *C. verrucosa* (V).

and whisker, histograms and scatter plots. After re-examining outliers to ensure their accuracy, some specimens were removed from the analysis due to specimen immaturity and missing values in the data matrix. The resulting data matrix contained 59 characters and 117 OTU's.

#### *Multivariate Analysis of Borrowed Specimens*

Numerical Taxonomy System 2.1 (NTSYS) was used for PCA. SPSS was used for Kaiser-Meyer-Olkin Measure of Sampling Adequacy. Correlation matrices from a standardized data matrix were used to create the principal components, which were plotted in Sigma Plot 8.0.

Qualitative and poor characters were removed from the multivariate data matrix. A PCA, with a reduced data matrix of 117 OTU's (47 *C. glaucescens*, 35 *C. jorii* and 35 *C. verrucosa*) and 26 characters, was used to clarify groups for a priori taxon labeling. Next, PCA's for each a priori group pairing were used to detect taxonomically important characters. The group pairing PCA's enabled the selection of taxonomically useful characters for the final PCA. Character selections for the final PCA were based on characters that graphically separated the taxa into groups while increasing the percent variation explained among OTU's. Characters were chosen by selecting at least three characters with the greatest eigenvector coefficients and/or those characters greater than or equal to 0.8 from each PCA a priori group pairing.

Geographic phenotypic variation was examined by labeling each OTU with state locality of specimen collection. State abbreviation labels were used to create artificial populations within each taxon. The resulting PCA was evaluated for clustering of taxa by single or multiple states. The data set was the same as that used for the final PCA of 10 characters and 117 OTU's.

### *Multivariate Analysis of Collected Populations Including Suspected Hybrids*

Specimens collected were labeled by population and taxon based on Jones (personal communication). Personal collections of 19 *C. glaucescens*, 4 *C. jorii*, 9 *C. verrucosa* and 3 possible hybrid specimens were combined with the borrowed specimen data matrix of 117 OTU's and 10 characters. The subsequent data matrix contained 10 characters for 154 specimens. The resulting PCA was examined for differences between taxa, populations and phenotypic variation between regions.

### *Multivariate Analysis with Carex jorii Holotype Specimen*

The *C. jorii* holotype specimen was measured and added to the data matrix of 117 OTU's and 10 characters. The resulting matrix was then analyzed with PCA to ensure that the treatment of taxa aligned with the type. The resulting PCA was evaluated for orientation of the type specimen in relation to the other specimens of the same taxon.

### Taxa Distributions

Distribution maps for *C. glaucescens*, *C. jorii* and *C. verrucosa* were created to clarify taxon distributions. Collection location data from annotated herbarium specimens were recorded and mapped in ArcView 8.0. Distribution maps were created from all borrowed herbarium specimens and those collected for this research with county and state location information.

### Phenology

Several taxon descriptions conflict or altogether lack phenological information (e.g., Godfrey & Wooten 1979). Phenology was examined by coding specimen collection dates for all borrowed herbarium specimens 1 through 12 (univariate analysis) and 1 through 365 (multivariate analysis). Univariate analysis utilized

histograms to compare taxon flowering dates by month. Flowering dates coded by Julian calendar day were used as a quantitative variable in the multivariate data analysis.

### Species Descriptions

Most character ranges and states for the species descriptions were taken from the characters recorded for numerical analysis (Tables 2, 7-9). Other descriptive characters were measured and evaluated separately. Ranges for the descriptions are plus and minus one standard deviation (SD) from the mean (Tables 7-9). When one SD from the mean exceeded the lower or upper bound of the range of specimens measured, the minimum or maximum recorded value was used.

TABLE 7. *Carex glaucescens* univariate statistics from quantitative data used in PCA's and the taxon description.

CHARACTER <sup>a</sup>	n	UNIT OF MEASURE	RANGE	MINIMUM	MAXIMUM	MEAN	SD. DEV.	MEAN -1 SD. DEV.	MEAN +1 SD. DEV.
FD	47	—	137.0	174.0	311.0	229.9	28.2	201.7	258.1
IL	47	cm	21.1	7.9	29.0	14.6	3.8	10.8	18.3
HP	47	cm	91.8	58.4	150.2	96.4	17.4	79.0	113.8
SLS	47	cm	6.4	1.3	7.7	4.1	1.3	2.8	5.4
SLP	47	cm	39.3	0.3	39.6	5.0	6.9	0.3 *	11.9
SLSC	47	mm	7.2	1.3	8.5	6.3	1.3	5.0	7.6
SWSC	47	mm	4.0	1.0	5.0	1.5	0.6	1.0 *	2.2
SWSP	47	mm	4.8	4.1	8.9	5.9	1.2	4.7	7.1
PLL	47	cm	9.8	0.0	9.8	2.3	1.6	0.7	3.9
PLUP	47	cm	6.2	0.0	6.2	1.6	1.1	0.4	2.7
PLLS	47	cm	4.2	2.1	6.3	3.7	0.9	2.8	4.6
PLUS	47	cm	3.9	0.4	4.2	2.7	0.8	1.9	3.6
PLC	47	mm	3.6	2.5	6.1	4.2	0.7	3.5	4.8
PWC	47	mm	1.0	1.0	2.0	1.4	0.3	1.1	1.7
PLCW	47	mm	2.8	0.5	3.3	1.4	0.6	0.9	2.0
LLS	47	cm	3.4	2.2	5.6	3.6	0.9	2.7	4.4
LLP	47	cm	11.7	0.5	12.2	3.2	1.9	1.3	5.2
GL	47	mm	1.7	3.0	4.7	3.7	0.4	3.4	4.1
GLBK	47	mm	0.5	0.2	0.7	0.4	0.1	0.3	0.5
GW	47	mm	0.7	1.8	2.5	2.2	0.2	2.0	2.4
GWB	47	mm	0.1	0.1	0.2	0.1	0.0	0.1	0.2
GLB	47	mm	0.1	0.1	0.2	0.1	0.0	0.1	0.2
LWP	47	mm	0.4	0.2	0.6	0.4	0.1	0.3	0.5
ALWOS	47	mm	1.1	1.9	3.0	2.5	0.2	2.3	2.8
AW	47	mm	0.6	1.7	2.3	2.0	0.1	1.8	2.1
GNN	47	—	8.0	2.0	10.0	4.9	2.1	2.8	6.9

<sup>a</sup> Character codes are in Table 2.

\* Minimum or maximum range values are used if 1 standard deviation from the mean exceeds the range.

n Refers to the sample size.

TABLE 8. *Carex jorii* univariate statistics from quantitative data used in the PCA's and the taxon description.

CHARACTER <sup>a</sup>	n	UNIT OF MEASURE	RANGE	MINIMUM	MAXIMUM	MEAN	SD. DEV.	MEAN -1 SD. DEV.	MEAN +1 SD. DEV.
FD	35	—	151.0	139.0	290.0	231.6	29.7	201.9	261.3
IL	35	cm	21.0	7.5	28.5	18.7	5.4	13.3	24.1
HP	35	cm	95.3	23.5	118.8	80.3	18.4	61.9	98.7
SLS	35	cm	4.1	2.0	6.0	4.3	1.1	3.1	5.4
SLP	35	cm	5.9	0.3	6.2	3.0	1.5	1.5	4.6
SLSC	35	mm	4.2	4.9	9.1	6.3	1.2	5.1	7.5
SWSC	35	mm	0.7	1.1	1.8	1.4	0.2	1.3	1.6
SWSP	35	mm	7.0	2.5	9.5	4.6	1.3	3.4	5.9
PLLP	35	cm	4.7	0.6	5.3	2.4	1.2	1.2	3.6
PLUP	35	cm	2.7	0.0	2.7	0.5	0.5	0.0	1.0
PLLS	35	cm	4.4	2.3	6.7	3.9	1.0	2.9	4.9
PLUS	35	cm	4.7	0.8	5.5	2.5	0.9	1.6	3.4
PLC	35	mm	2.8	2.5	5.3	3.9	0.6	3.3	4.5
PWC	35	mm	0.9	1.2	2.1	1.5	0.2	1.3	1.7
PLCW	35	mm	1.3	0.3	1.6	1.1	0.4	0.7	1.5
LLS	35	cm	4.0	2.6	6.5	4.3	1.0	3.2	5.3
LLP	35	cm	5.8	2.0	7.8	4.4	1.8	2.6	6.2
GL	35	mm	1.5	3.4	4.9	4.0	0.3	3.6	4.3
GLBK	35	mm	0.7	0.7	1.4	1.0	0.1	0.8	1.1
GW	35	mm	1.0	2.1	3.1	2.7	0.2	2.5	2.9
GWB	35	mm	0.3	0.0	0.3	0.1	0.1	0.1	0.2
GLB	35	mm	0.2	0.0	0.2	0.1	0.0	0.0	0.1
LWP	35	mm	0.3	0.3	0.6	0.4	0.1	0.3	0.5
ALWOS	35	mm	0.6	2.3	2.9	2.7	0.2	2.5	2.8
AW	35	mm	0.9	1.9	2.8	2.4	0.2	2.2	2.6
GNN	35	—	8.0	12.0	20.0	14.8	1.4	13.4	16.2

<sup>a</sup> Character codes are in Table 2.

\* Minimum or maximum range values are used if 1 standard deviation from the mean exceeds the range.

n Refers to the sample size.

TABLE 9. *Carex verrucosa* univariate statistics from quantitative data used in the PCA's and the taxon description.

CHARACTER <sup>a</sup>	n	UNIT OF MEASURE	RANGE	MINIMUM	MAXIMUM	MEAN	SD. DEV.	MEAN -1 SD. DEV.	MEAN +1 SD. DEV.
FD	35	—	200.0	97.0	297.0	172.5	62.5	110.0	235.1
IL	35	cm	25.2	8.5	33.7	20.0	7.7	12.3	27.7
HP	35	cm	93.2	63.4	156.6	99.5	19.2	80.2	118.7
SLS	35	cm	5.8	2.0	7.8	3.8	1.1	2.7	5.0
SLP	35	cm	30.5	0.3	30.8	6.0	8.4	0.3 *	14.4
SLSC	35	mm	6.8	4.9	11.7	6.7	1.4	5.3	8.0
SWSC	35	mm	1.9	0.7	2.6	1.5	0.4	1.1	1.9
SWSP	35	mm	12.0	3.5	15.5	7.5	3.2	4.2	10.7
PLL <sub>P</sub>	35	cm	2.5	0.2	2.8	1.1	0.6	0.6	1.7
PLUP	35	cm	1.2	0.0	1.2	0.2	0.2	0.0	0.5
PLLS	35	cm	3.7	1.5	5.2	3.6	1.0	2.6	4.5
PLUS	35	cm	2.9	0.8	3.7	2.2	0.8	1.4	2.9
PLC	35	mm	3.4	3.6	7.0	4.8	0.9	3.9	5.7
PWC	35	mm	3.0	1.1	4.1	1.6	0.5	1.1	2.0
PLCW	35	mm	3.3	0.4	3.7	1.6	0.7	0.9	2.4
LLS	35	cm	4.1	2.0	6.1	3.8	0.9	2.8	4.7
LLP	35	cm	7.8	0.7	8.4	2.9	1.8	1.1	4.7
GL	35	mm	1.4	3.2	4.6	3.8	0.4	3.4	4.2
GLBK	35	mm	0.8	0.1	0.9	0.4	0.2	0.2	0.6
GW	35	mm	0.9	2.2	3.1	2.5	0.2	2.4	2.7
GWB	35	mm	0.3	0.2	0.5	0.3	0.1	0.2	0.4
GLB	35	mm	0.3	0.2	0.5	0.3	0.1	0.2	0.4
LWP	35	mm	0.4	0.3	0.7	0.5	0.1	0.4	0.6
ALWOS	35	mm	1.0	2.2	3.2	2.6	0.2	2.3	2.8
AW	35	mm	0.9	2.1	3.0	2.5	0.2	2.3	2.6
GNN	35	—	12.0	4.0	16.0	11.1	2.7	8.5	13.8

a Character codes are in Table 2.

\* Minimum or maximum range values are used if 1 standard deviation from the mean exceeds the range.

n Refers to the sample size.

## RESULTS AND DISCUSSION

### Cytology

#### *Chromosome Numbers*

Mitotic counts were not achieved because the pretreatment application was not developed adequately. Chromosomes were never viewed in metaphase, leading to the conclusion that results may be improved with a different or adjusted pretreatment method. Pretreatment methods can be adjusted by chemical type, concentration and application time, however, only application time was adjusted. The average digestion time for small roots was 35 minutes, which was increased or decreased for varying root sizes.

Definite meiotic chromosome counts were not obtained. Most field collections contained mature anthers with pollen grains, rather than those suitable for chromosome counts. Pollen grains were present in these specimens because collection was about a week too late. If specimens are collected after the majority of the staminate spikelets mature, it reduces the number of usable anthers. Within the staminate spike, the central spikelets mature first, followed by a few proximal and distal spikelets.

#### *Pollen Stainability*

Pollen viability, as estimated by pollen stainability with IKI, was consistently high for *C. glaucescens* ranging from 60-84% (Table 6). It was inconsistent for *C. jorii* ranging from 14-80% and *C. verrucosa* varied from 5-91%. Percent pollen viability was low for specimens considered to be hybrid taxa ranging from 3-31%. Low percent pollen viability was not correlated with hybrid taxa, because of the low pollen viability for some non-hybrid taxa. The varied pollen viability results cannot be used to definitively corroborate the existence of hybrid taxa.



Sexual reproduction among the taxa appears to vary, since low pollen viability was not solely aligned with hybrid taxa and high pollen viability was not always found in non-hybrid taxa. Because pollen viability results varied, further research within the section should evaluate other modes of reproduction such as apomixis. The rhizomonous habit of the taxa may be a primary mode of reproduction, if seed set is low from low pollen viability.

#### Achene and Perigynia Micromorphology

Achene micromorphology analysis yielded little useable taxonomic information because *Carex* achenes are better examined by methods that remove extraneous layers of cuticle prior to scanning electron microscopy. A misinterpretation of preliminary micrographs led to the conclusion that the cuticle and outer periclinal wall were absent and it was unnecessary to use cuticle and outer periclinal wall removal procedures. It is necessary to remove the cuticle and outer periclinal walls of *Carex* achenes because they are essentially devoid of useable taxonomic features (Menapace et al. 1986). Taxonomically pertinent features on the inner periclinal wall (lateral wall) of the epidermis (Menapace & Wujek 1987) are not detectable in the micrographs. The micrographs do exhibit central silica bodies and an alveolate cell shape in all three taxa (Figs. 1-2).

The perigynia micromorphology micrographs yield information that generally separates *C. jooirii* specimens (Figs. 1-2) from other taxa. *Carex jooirii* generally exhibits papillae that are reduced, appearing rounded over and sometimes absent with concave reticulate cells. Perigynia of most *C. jooirii* have clavate papillae on the beak and reduced papillae covering the body. Some *C. jooirii* specimens are entirely covered

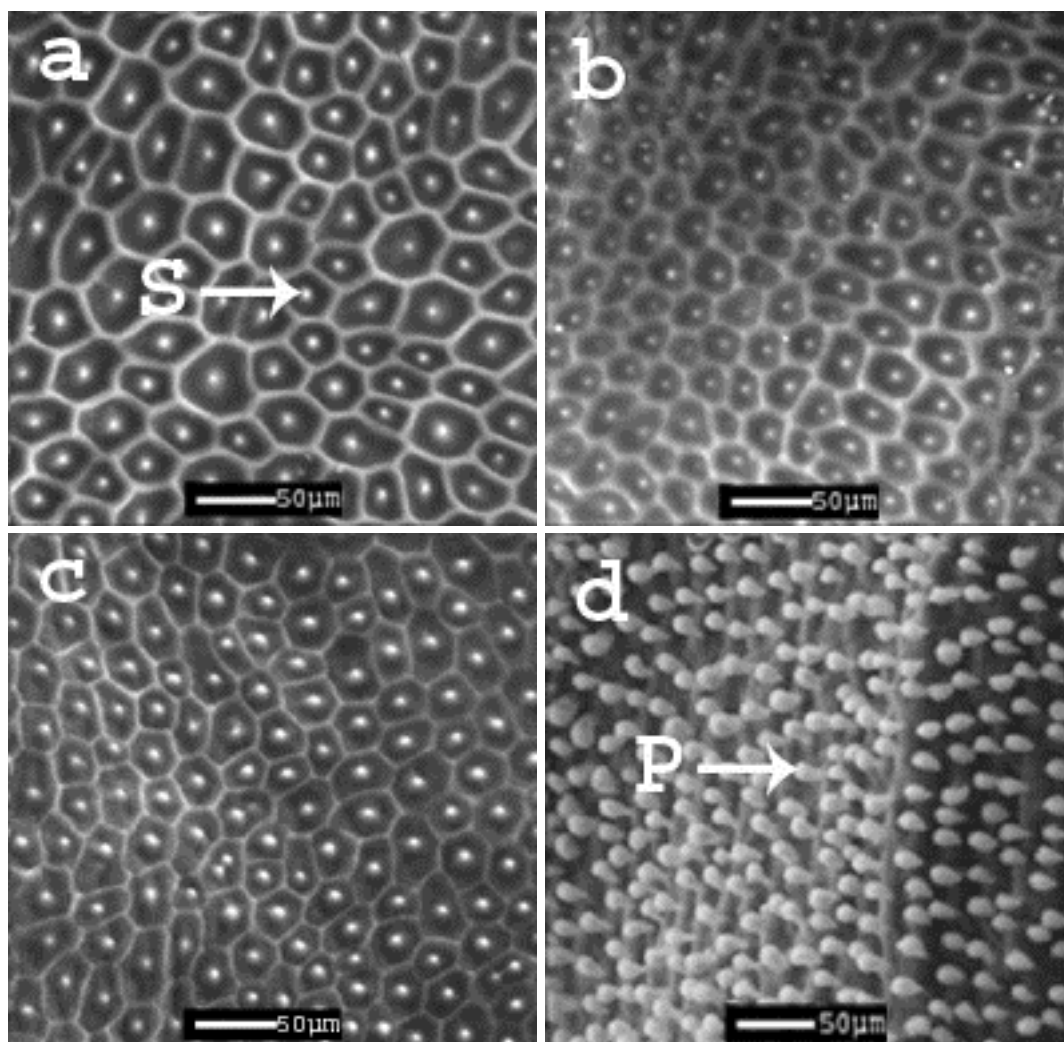


FIG. 1. Environmental scanning electron micrographs of *Carex* section *Glaucescens*. Achene epidermal cells: a. *C. glaucescens*, S. Central silica body. b. *C. jorii*. c. *C. verrucosa*. Perigynia surface: d. *C. glaucescens*, P. Clavate papillae.

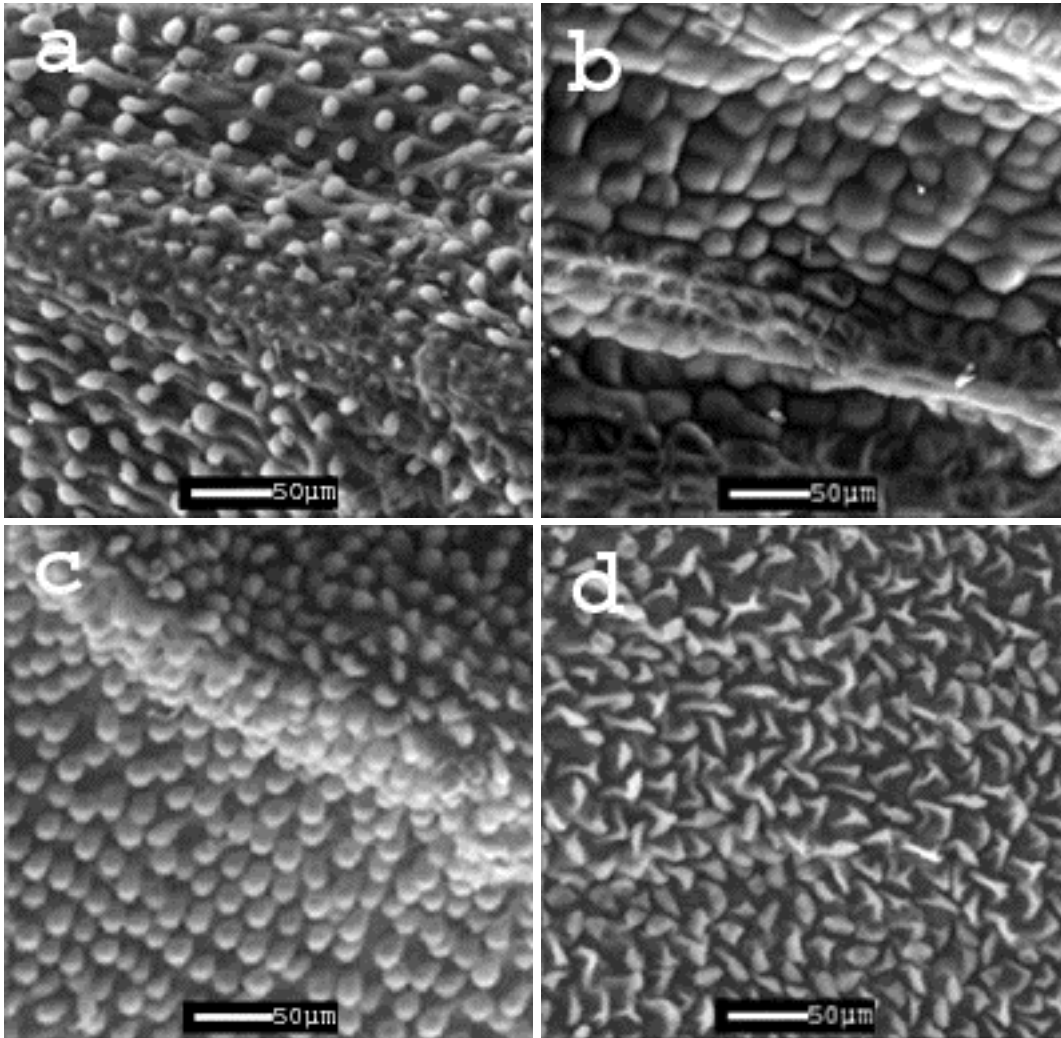


FIG. 2. Environmental scanning electron micrographs of *Carex* section *Glaucescentes*. Perigynia surface: a. *C. jorii* reduced papillae. b. *C. jorii* papillae rounded to absent with concave reticulate cells. c. *C. verrucosa* clavate papillae. d. *C. verrucosa* longitudinally troughed or compressed papillae.

with clavate papillae. The papillae on *C. glaucescens* and *C. verrucosa* are clavate to terete and sometimes compressed or troughed. Compressed papillae may be a true character state or a result of the vacuum used in ESEM. Perigynia texture is not a consistent taxonomic character to separate these taxa.

### Data Analysis

#### *Univariate Analysis*

Descriptive statistics for the continuous variables used in PCA for *C. glaucescens*, *C. jorii* and *C. verrucosa* are in Tables 7-9, respectively. The mean flowering dates for *C. verrucosa* are approximately two months earlier than *C. glaucescens* and *C. jorii*. Measurement ranges overlap for most characters, however each character mean is different for at least one taxon.

#### *Multivariate Analysis of Borrowed Specimens*

The PCA of 26 characters and all three taxa explains 48.06 percent of the total variation among OTU's (Table 10). This percent total variation explained was low, initiating steps to increase variation explained by decreasing characters used. The PCA depicts three groups, which were aligned with the accepted taxa (Fig. 3). Because the PCA depicted 3 groups, the OTU's were A Priori labeled by taxon. *Carex jorii* is rather distinctly separated from *C. glaucescens* and *C. verrucosa*. However, it is retained in the following analyses because it is suspected to hybridize with *C. verrucosa* and possibly *C. glaucescens*.

In order to create a PCA with greater percent variation explained, PCA's of taxon pairings were used to decrease the number of characters. The following taxon pairing PCA's, explain a low percent of the total variation between OTU's. The percent

TABLE 10. Eigenvector coefficients from the PCA of 26 characters used to clarify groups for a priori taxon labeling. Percent trace and cumulative variation explained for the first three principal components are presented.

CHARACTER <sup>a</sup>	PC 1	PC 2	PC 3
FD	0.114	0.471	0.116
IL	-0.753	-0.203	0.049
HP	-0.470	-0.074	-0.535
SLS	-0.761	0.250	-0.109
SLP	-0.209	-0.021	-0.275
SLSC	-0.419	-0.121	-0.239
SWSC	0.098	0.002	-0.034
SWSP	-0.071	-0.302	-0.501
PLLP	-0.510	0.661	-0.008
PLUP	-0.157	0.715	-0.393
PLLS	-0.778	0.202	-0.106
PLUS	-0.545	0.366	-0.208
PLC	-0.292	-0.313	-0.523
PWC	0.056	-0.232	0.042
PLCW	-0.107	-0.064	-0.541
LLS	-0.822	0.105	0.084
LLP	-0.638	0.374	0.140
GL	-0.302	0.021	0.355
GLBK	-0.312	0.131	0.817
GW	-0.334	-0.479	0.559
GWB	-0.096	-0.725	-0.336
GLB	-0.115	-0.723	-0.356
LWP	-0.458	-0.535	-0.198
ALWOS	-0.229	-0.005	0.259
AW	-0.372	-0.675	0.373
GNN	-0.344	-0.455	0.681
% TRACE	18.560	15.812	13.692
COMBINED % TRACE			48.064

a Character codes are in Table 2.

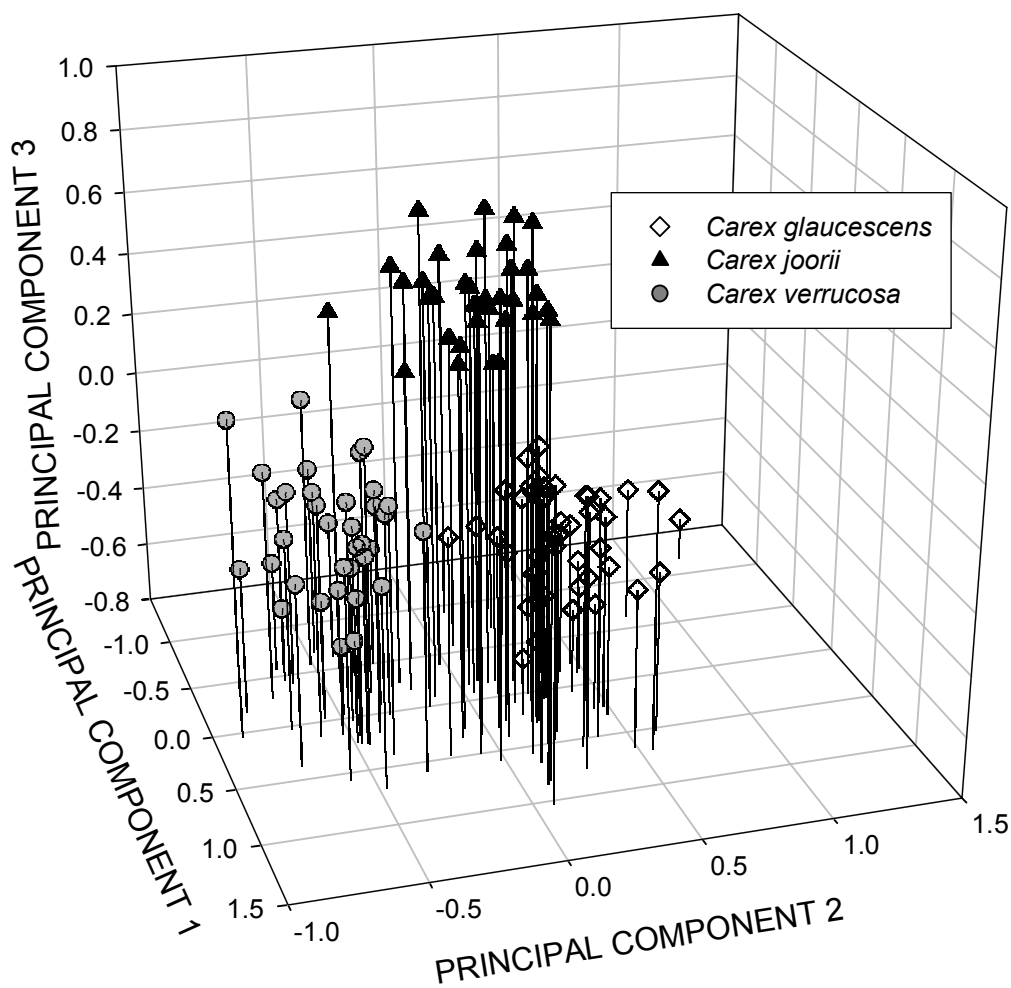


FIG. 3. PCA on *Carex glaucescens*, *C. jorii* and *C. verrucosa*, using 26 characters. The first three principal components are plotted.

of total variation explained is low because many of the 26 characters used are not taxonomically important. The use of taxon pairing PCA's enabled the selection of taxonomically important characters for a subsequent PCA.

Eigenvector coefficients of selected characters from the first taxon pairing PCA of 26 characters from *C. glaucescens* and *C. jorii* are length of lowermost lateral spike (LLS) (0.848), inflorescence length (IL) (0.813) and length of terminal spike (SLS) (0.806) (Table 11). *Carex glaucescens* and *C. jorii* are clearly separated by the first two principal components (Fig. 4). The first three principal components explain 49.071 percent of the total variation among OTU's. This PCA is an unacceptable representation of the OTU's because it explains a low amount of variation between taxa.

The second taxon pairing PCA contained 26 characters from *C. glaucescens* and *C. verrucosa*. Eigenvector coefficients of selected characters from the second PCA taxon pairing are widest point of achene (AW) (0.890), number of veins on perigynia faces (GNN) (0.844), length of perigynia stipitate base (GLB) (0.839) and width of perigynia stipitate base (GWB) (0.816) (Table 12). The first three principal components explain 49.064 percent of the total variation among OTU's. *Carex glaucescens* and *C. verrucosa* are separated by the first two principal components in Figure 5. This PCA is an unacceptable representation of the OTU's because it explains a low amount of variation between taxa.

The third taxon pairing PCA contained 26 characters of *C. jorii* and *C. verrucosa*. Eigenvector coefficients of selected characters from the third PCA taxon pairing are length of perigynia beak (GLBK) (0.841), length of second lowermost lateral pistillate spike peduncle (PLLP) (0.802), length of perigynia stipitate base (GLB) (0.690)

TABLE 11. Eigenvector coefficients from the taxon pairing PCA of 26 characters from *Carex glaucescens* and *C. jorii*. Percent trace and cumulative variation explained for the first three principal components are presented.

CHARACTER <sup>a</sup>	PC 1	PC 2	PC 3
FD	0.046	0.053	0.419
IL*	-0.813	0.159	0.061
HP	-0.422	-0.577	-0.037
SLS*	-0.806	-0.201	-0.092
SLP	-0.136	-0.323	0.070
SLSC	-0.488	-0.206	-0.339
SWSC	0.257	0.001	0.409
SWSP	-0.269	-0.581	-0.383
PLLP	-0.656	-0.261	0.504
PLUP	-0.245	-0.715	0.336
PLLS	-0.790	-0.133	-0.136
PLUS	-0.529	-0.369	-0.162
PLC	-0.358	-0.443	0.236
PWC	0.090	0.222	0.243
PLCW	-0.127	-0.544	0.480
LLS*	-0.848	0.076	-0.029
LLP	-0.700	0.016	0.445
GL	-0.379	0.304	-0.137
GLBK	-0.395	0.832	0.121
GW	-0.309	0.742	0.171
GWB	0.068	-0.196	-0.258
GLB	0.174	-0.332	-0.257
LWP	-0.535	0.054	-0.401
ALWOS	-0.360	0.225	-0.319
AW	-0.381	0.723	0.008
GNN	-0.305	0.878	0.028
% TRACE	22.020	19.221	7.830
COMBINED % TRACE			49.071

a Character codes are in Table 2.

\* Characters selected for final PCA.



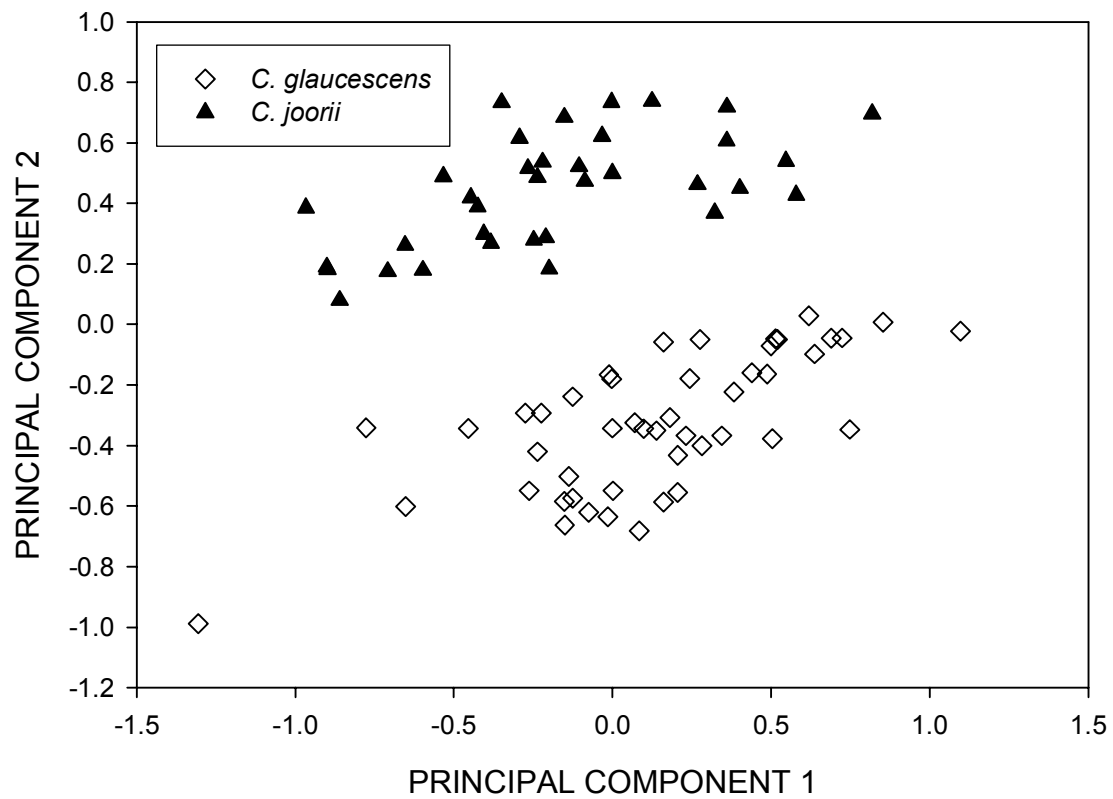


FIG. 4. PCA on *Carex glaucescens* and *C. jorii* in a taxon pairing with 26 characters. The first and second principal components are plotted.

TABLE 12. Eigenvector coefficients from the taxon pairing PCA of 26 characters from *Carex glaucescens* and *C. verrucosa*. Percent trace and cumulative variation explained for the first three principal components are presented.

CHARACTER <sup>a</sup>	PC 1	PC 2	PC 3
FD	0.521	0.002	0.646
IL	-0.540	-0.586	-0.320
HP	-0.268	-0.585	0.155
SLS	-0.047	-0.782	-0.160
SLP	-0.107	-0.267	-0.103
SLSC	-0.254	-0.354	0.259
SWSC	0.030	0.049	-0.136
SWSP	-0.285	0.002	0.434
PLLP	0.442	-0.692	0.066
PLUP	0.628	-0.543	0.077
PLLS	-0.105	-0.779	-0.195
PLUS	0.179	-0.660	-0.065
PLC	-0.426	-0.221	0.698
PWC	-0.191	0.116	0.048
PLCW	-0.121	-0.184	0.696
LLS	-0.247	-0.734	-0.163
LLP	0.077	-0.685	-0.111
GL	-0.126	-0.186	0.450
GLBK	-0.021	-0.168	0.064
GW	-0.791	0.122	0.052
GWB*	-0.816	0.120	-0.125
GLB*	-0.839	0.030	-0.036
LWP	-0.668	-0.159	-0.088
ALWOS	-0.078	-0.154	0.575
AW*	-0.890	0.003	0.078
GNN*	-0.844	0.109	-0.004
% TRACE	21.907	17.577	9.579
COMBINED % TRACE			49.064

a Character codes are in Table 2.

\* Characters selected for final PCA.

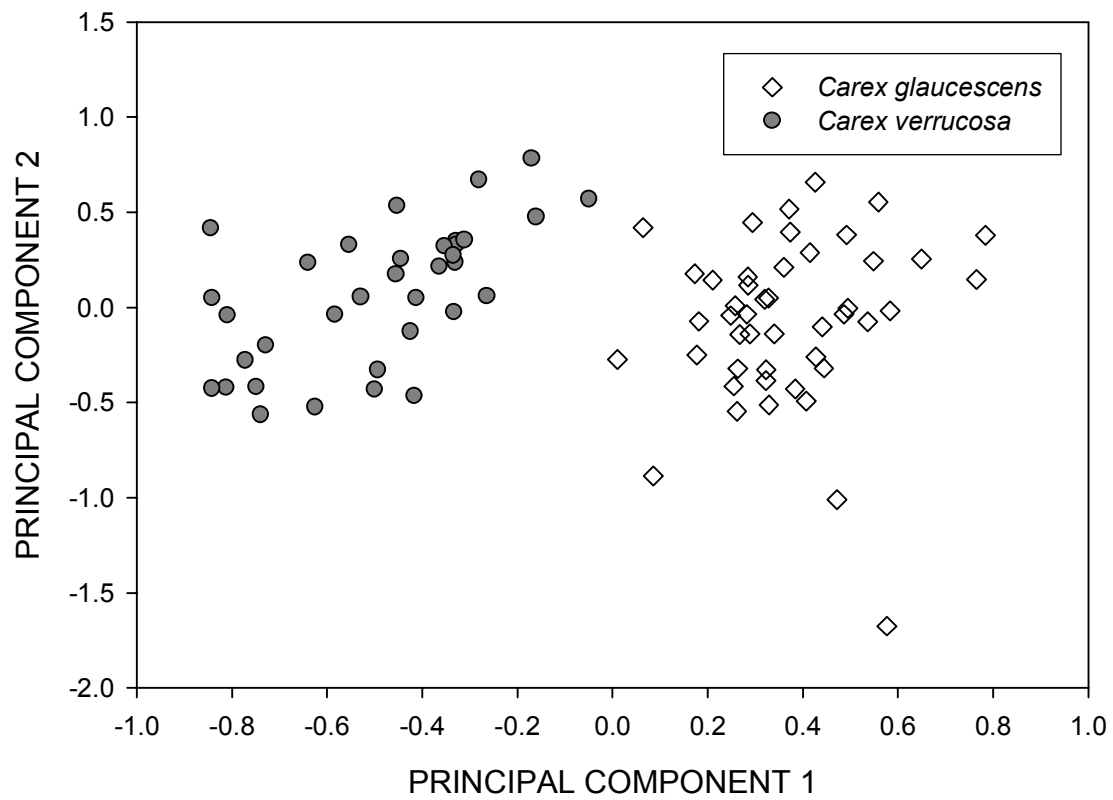


FIG. 5. PCA on *Carex glaucescens* and *C. verrucosa* in a taxon pairing with 26 characters. The first and second principal components are plotted.

and length of lowermost lateral spike peduncle (LLP) (0.678) (Table 13). The first three principal components explain 50.339 percent of the total variation among the OTU's. *Carex jorii* and *C. verrucosa* are separated by the first two principal components (Fig. 6). Characters selected from the taxon pairings were used in a PCA of all three taxa. The taxon pairing PCA's have low percent variation explained among the OTU's (Table 14). Each pairing only marginally increased percent variation explained because the taxon excluded from each pairing is closely related to the two remaining taxa. The large number of taxonomically non-important characters is also responsible for decreased percent variation.

Finally, a PCA was performed on the data matrix of 117 OTU's using 10 characters selected from taxon pairing PCA's (Table 14). The ten characters selected from the three pairings are LLS, IL, SLS, AW, GNN, GLB, GWB, GLBK, PLLP, LLP. The first three principal components explained 76.77 percent of the variation among OTU's. Reducing the data matrix to 10 characters greatly increased the percent variation explained among OTU's. A three dimensional representation of the PCA shows three groups of taxa (Fig. 7). The groups correspond with the a priori labeling of OTU's with *C. glaucescens*, *C. jorii* and *C. verrucosa*. OTU's of the same taxon are grouped more closely to each other and further from other taxa than when 26 characters were used. *Carex glaucescens* and *C. jorii* are separated by principal components 1 and 3 (Fig. 8). *Carex glaucescens* and *C. verrucosa* are separated by principal components 1 and 2 (Fig. 9). *Carex jorii* and *C. verrucosa* are separated by principal components 1 and 3 (Fig. 10). The OTU's from the PCA of 10 characters were labeled by state abbreviation (e.g. TX and LA). Scatter plots of principal components 1

TABLE 13. Eigenvector coefficients from the taxon pairing PCA of 26 characters from *Carex jorii* and *C. verrucosa*. Percent trace and cumulative variation explained for the first three principal components are presented.

CHARACTER <sup>a</sup>	PC 1	PC 2	PC 3
FD	0.363	0.275	0.570
IL	0.239	-0.661	-0.337
HP	-0.224	-0.728	0.324
SLS	0.498	-0.614	-0.060
SLP	-0.118	-0.403	-0.014
SLSC	-0.032	-0.457	0.315
SWSC	-0.039	-0.233	0.316
SWSP	-0.505	-0.241	0.234
PLLP*	0.802	-0.181	-0.159
PLUP	0.595	-0.182	0.013
PLLS	0.469	-0.679	-0.140
PLUS	0.481	-0.569	0.106
PLC	-0.428	-0.486	0.439
PWC	-0.144	0.088	0.170
PLCW	-0.360	-0.288	0.337
LLS	0.569	-0.617	-0.121
LLP*	0.678	-0.323	-0.286
GL	0.377	0.011	0.593
GLBK*	0.841	0.273	0.137
GW	0.344	-0.043	0.260
GWB	-0.676	-0.433	-0.188
GLB*	-0.690	-0.443	-0.075
LWP	-0.326	-0.633	-0.040
ALWOS	0.232	0.112	0.737
AW	-0.161	-0.184	0.632
GNN	0.655	0.068	0.238
% TRACE	22.4625	17.2264	10.6498
COMBINED % TRACE			50.339

a Character codes are in Table 2.

\* Characters selected for final PCA.

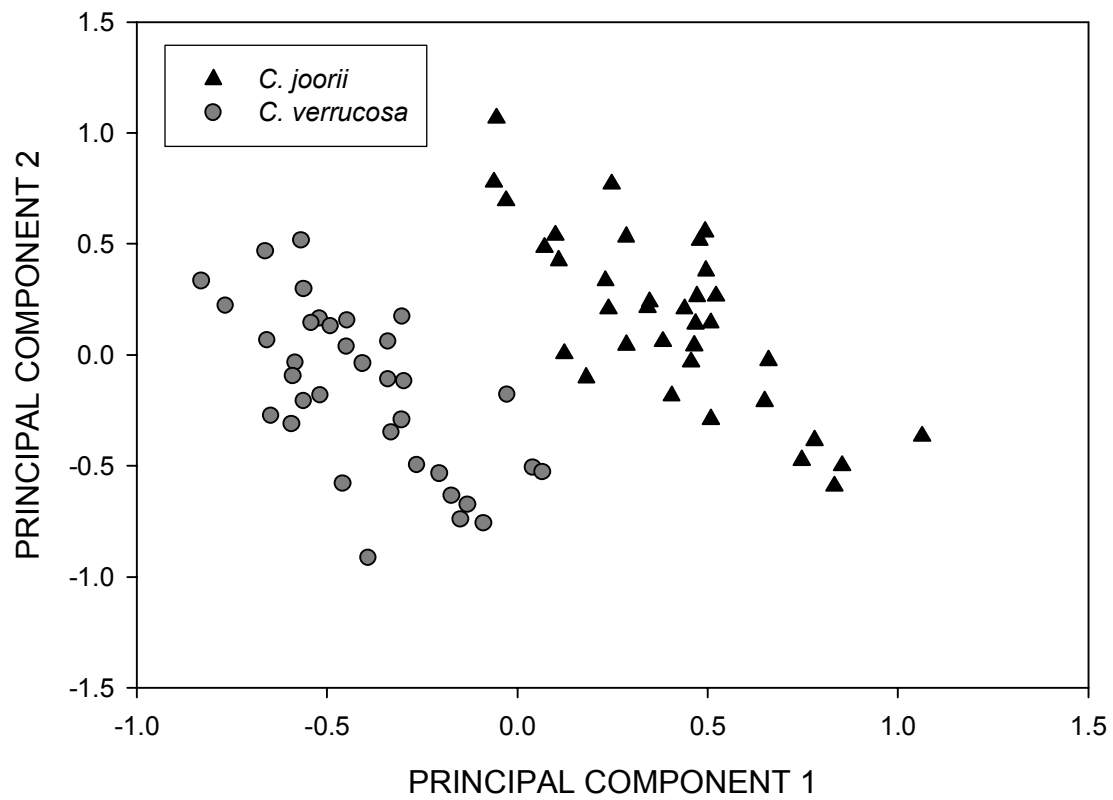


FIG. 6. PCA on *Carex jorii* and *C. verrucosa* in a taxon pairing with 26 characters. The first and second principal components are plotted.

TABLE 14. Eigenvector coefficients from the PCA of 10 characters. Percent trace and cumulative variation explained for the first three principal components are presented.

CHARACTER <sup>a</sup>	PC 1	PC 2	PC 3
IL	0.715	0.422	0.312
SLS	0.694	-0.069	0.327
PLLP	0.695	-0.500	0.275
LLS	0.763	0.128	0.118
LLP	0.824	-0.166	0.253
GLBK	0.589	0.005	-0.701
GWB	-0.198	0.767	0.437
GLB	-0.185	0.777	0.434
AW	0.257	0.742	-0.368
GNN	0.427	0.551	-0.620
% TRACE	34.057	25.221	17.491
COMBINED % TRACE			76.770

a Character codes are in Table 2.

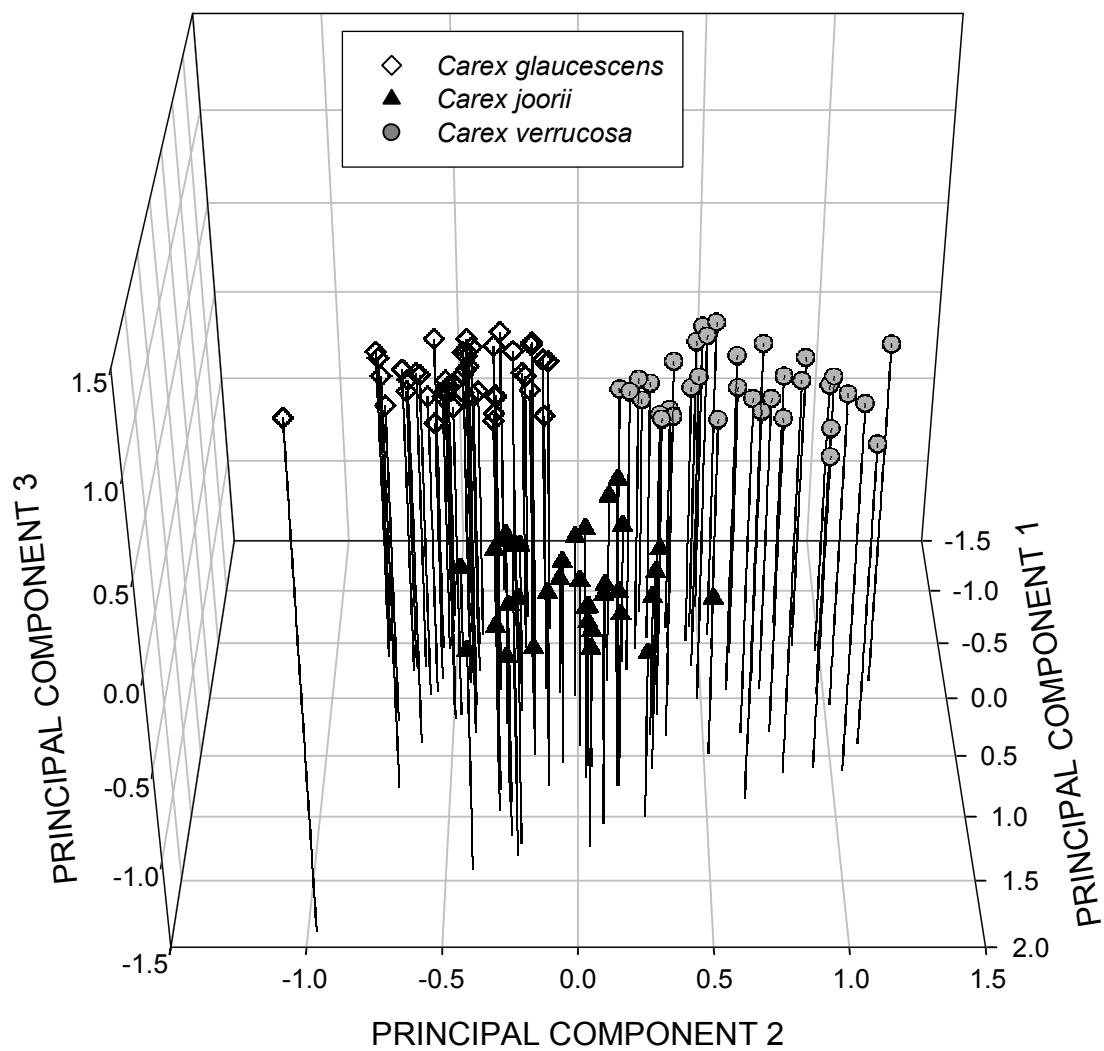


FIG. 7. PCA for *Carex glaucescens*, *C. jorii* and *C. verrucosa* with a reduced data matrix of 10 characters. The first three principal components are plotted.



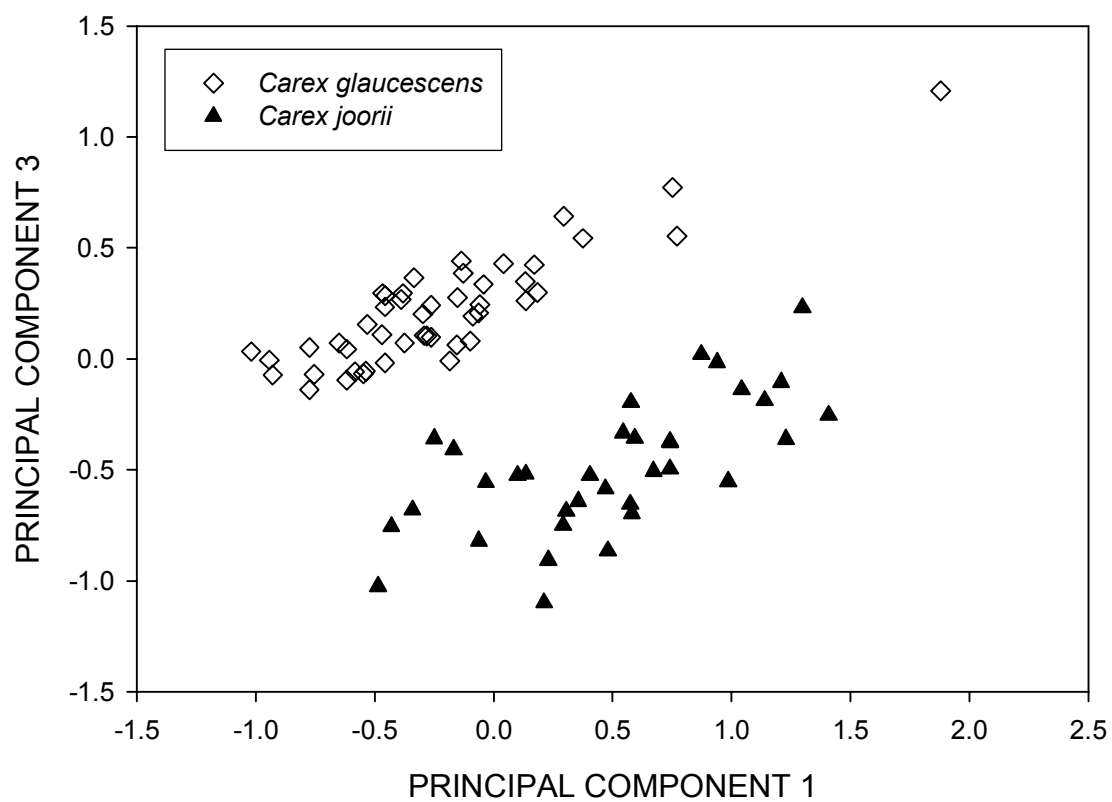


FIG. 8. The first and third principal components are plotted for *Carex glaucescens* and *C. jorii* from a PCA of *C. glaucescens*, *C. jorii* and *C. verrucosa* on a reduced data matrix of 10 characters.

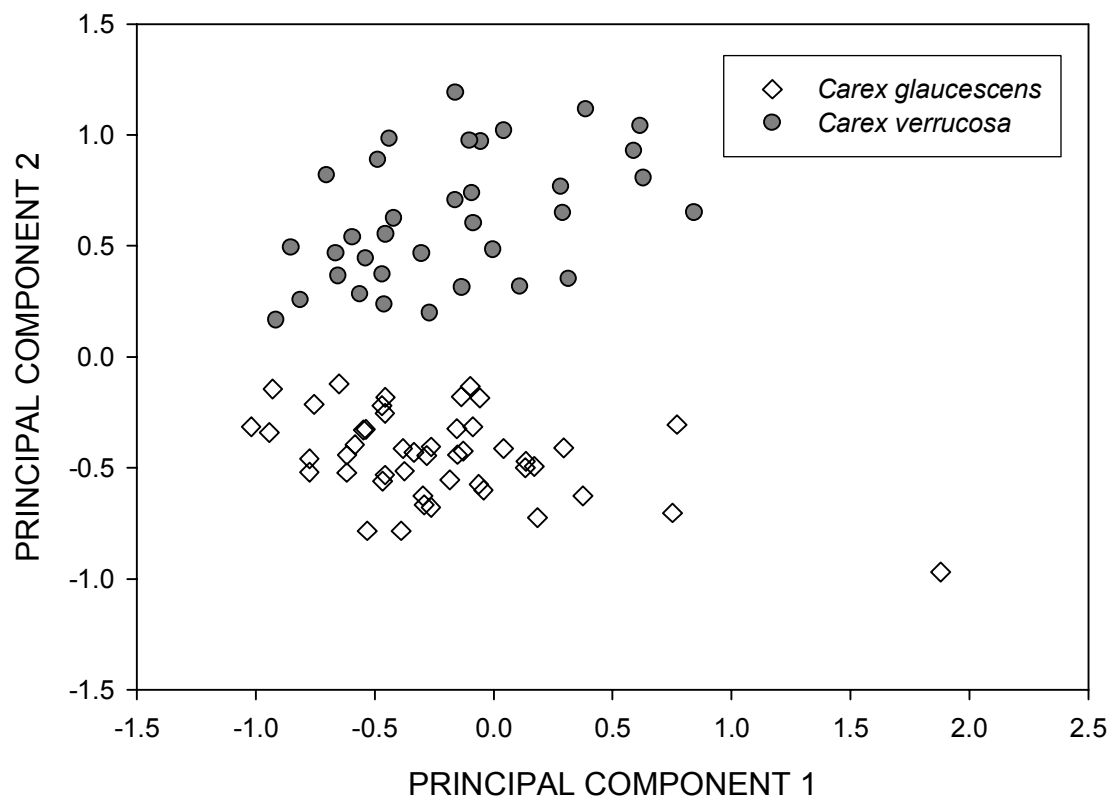


FIG. 9. The first and second principal components are plotted for *Carex glaucescens* and *C. verrucosa* from a PCA of *C. glaucescens*, *C. jorii* and *C. verrucosa* on a reduced data matrix of 10 characters.

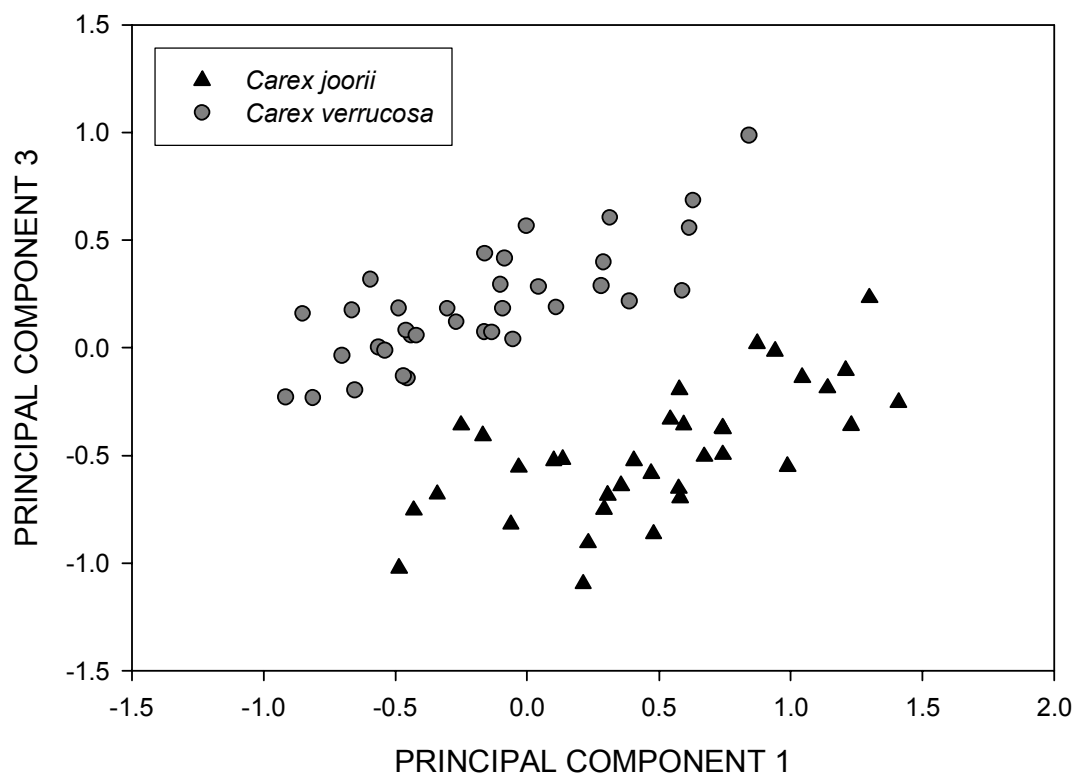


FIG. 10. The first and third principal components are plotted for *Carex jorii* and *C. verrucosa* from a PCA for *C. glaucescens*, *C. jorii* and *C. verrucosa* on a reduced data matrix of 10 characters.

and 2 were graphed separately for each taxon (Figs. 11-13). The scatter plots were evaluated for clustering of OTU's by single and multiple state groupings. There was no distinct clustering by states for *C. glaucescens*, *C. jorii* or *C. verrucosa*, suggesting that there are not distinct morphological differences based on longitude, latitude or elevation within each taxon.

#### *Multivariate Analysis of Collected Populations Including Suspected Hybrids*

The same three separate clusters were found in the combined PCA of borrowed specimens and personal collections. Eigenvector coefficients greater than 0.7 from principal component 1 are LLP (0.729) and LLS (0.719) (Table 15). Characters with eigenvector coefficients greater than 0.7 from the second principal component are GLB (0.756), GWB (0.749) and AW (0.743). The first three principal components explain about 74 percent of the total variation among the OTU's. The percent variation is reduced by about 2.6 percent, which is likely explained because the suspected hybrid taxa were included. These specimens were infertile and therefore achene width was not measured. *Carex glaucescens*, *C. jorii* and *C. verrucosa* are depicted as separate clusters by the first three principal components (Fig. 14). Personal collection specimens align with borrowed specimens of the same taxon. This reaffirms the lack of phenotypic variation based on geographic locality.

The suspected hybrid specimens are positioned at the margin of *C. jorii* and *C. verrucosa* PCA graphical data display (Fig. 14). The three suspected hybrid specimens are infertile and did not produce achenes in the year of study. Morphologically, the specimens mostly resemble *C. jorii* with squarrose, inflated and prominently veined perigynia and tapered pistillate scales. They resemble *C. verrucosa* by flowering in

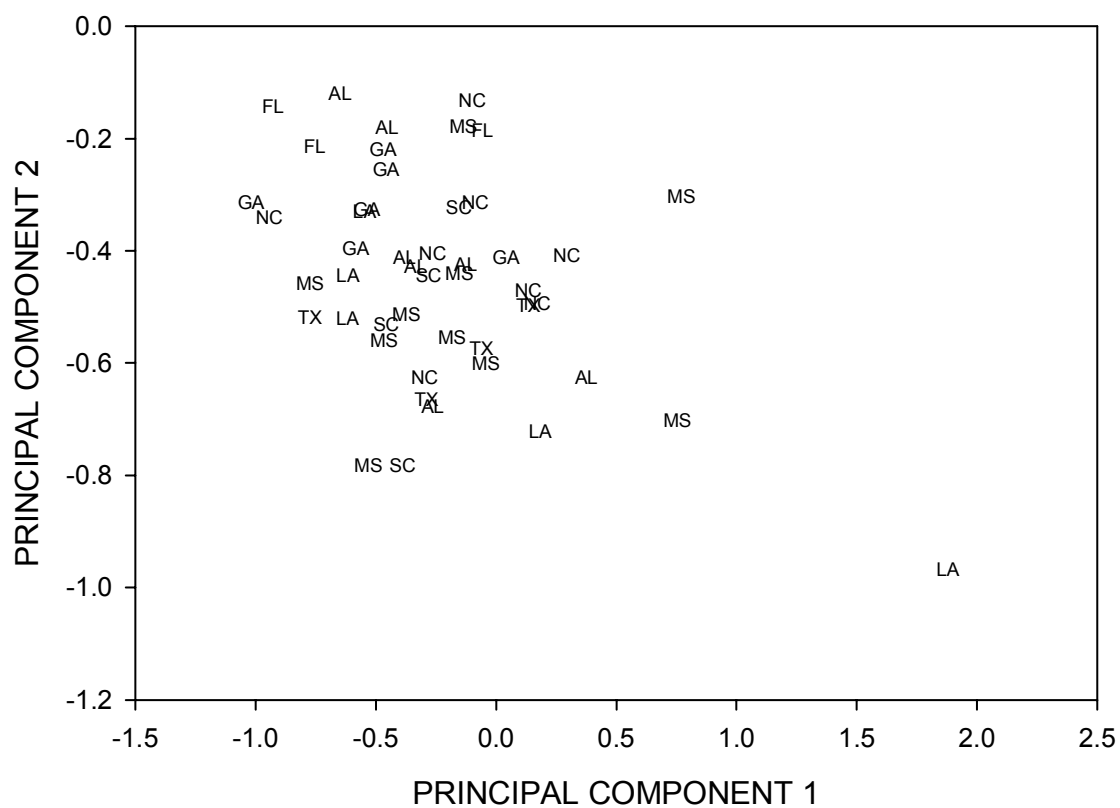


FIG. 11. The first and second principal components are plotted for *Carex glaucescens*. OTU's are labeled by state abbreviation, to aid in the examination of geographic phenotypic variation.

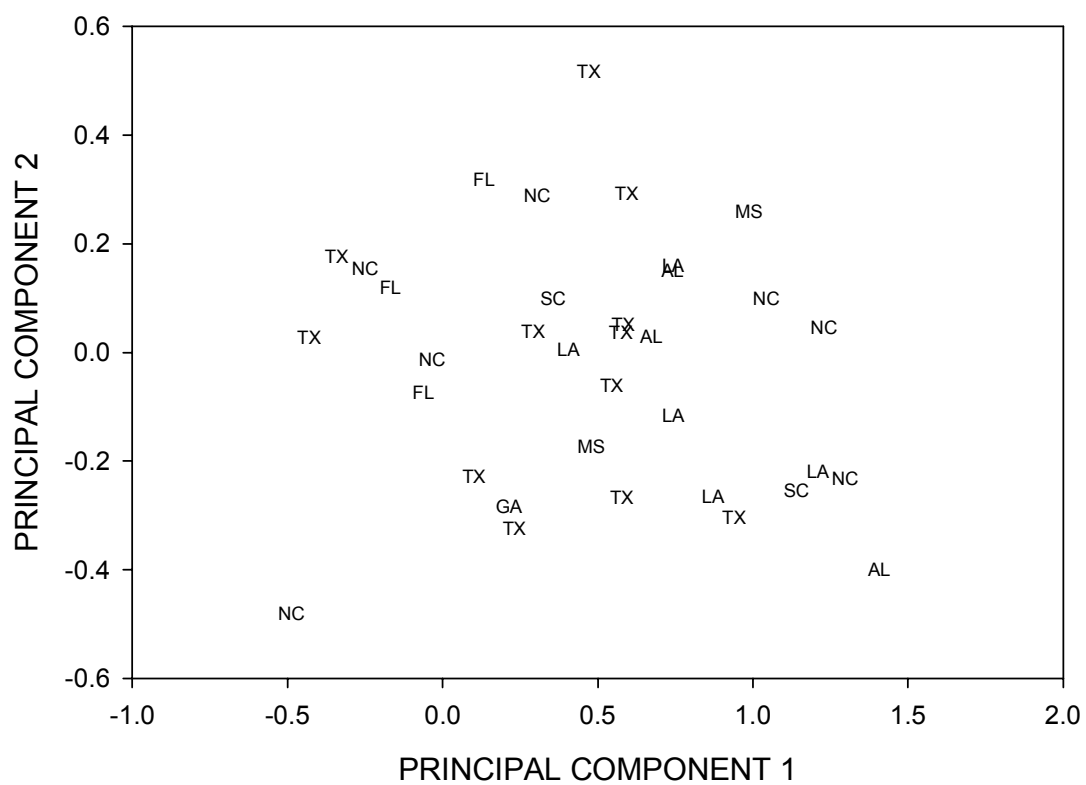


FIG. 12. The first and second principal components are plotted for *Carex joorii*. OTU's are labeled by state abbreviation to aid in the examination of geographic phenotypic variation.

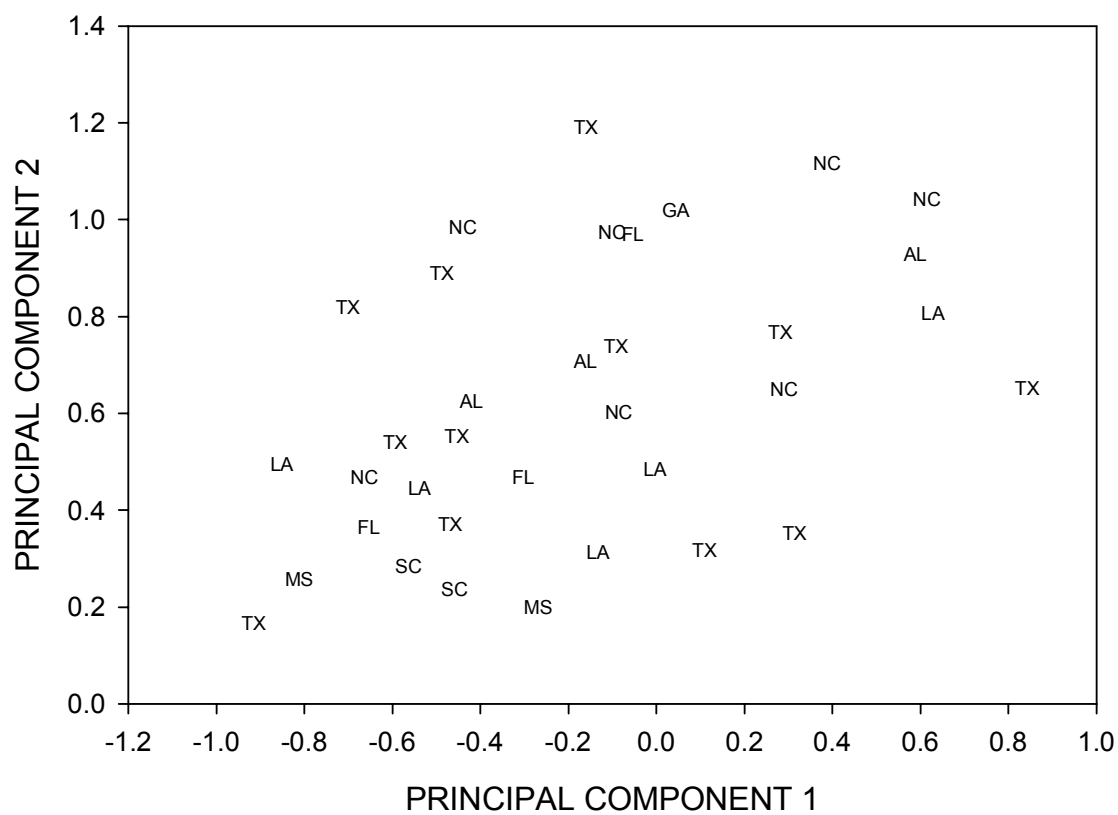


FIG. 13. The first and second principal components are plotted for *Carex verrucosa*. OTU's are labeled by state abbreviation to aid in the examination of geographic phenotypic variation.

TABLE 15. Eigenvector coefficients from the PCA of 10 characters from borrowed herbarium specimens and personal collections. Percent trace and cumulative variation explained for the first three principal components are presented.

CHARACTER <sup>a</sup>	PC 1	PC 2	PC 3
IL	0.674	0.393	0.369
SLS	0.653	0.064	0.372
PLLP	0.575	-0.629	0.306
LLS	0.719	0.167	0.118
LLP	0.729	-0.370	0.271
GLBK	0.625	-0.019	-0.667
GWB	-0.180	0.749	0.416
GLB	-0.165	0.756	0.416
AW	0.330	0.743	-0.301
GNN	0.465	0.497	-0.623
% TRACE	30.338	26.503	17.241
COMBINED % TRACE			74.082

a Character codes are in Table 2.



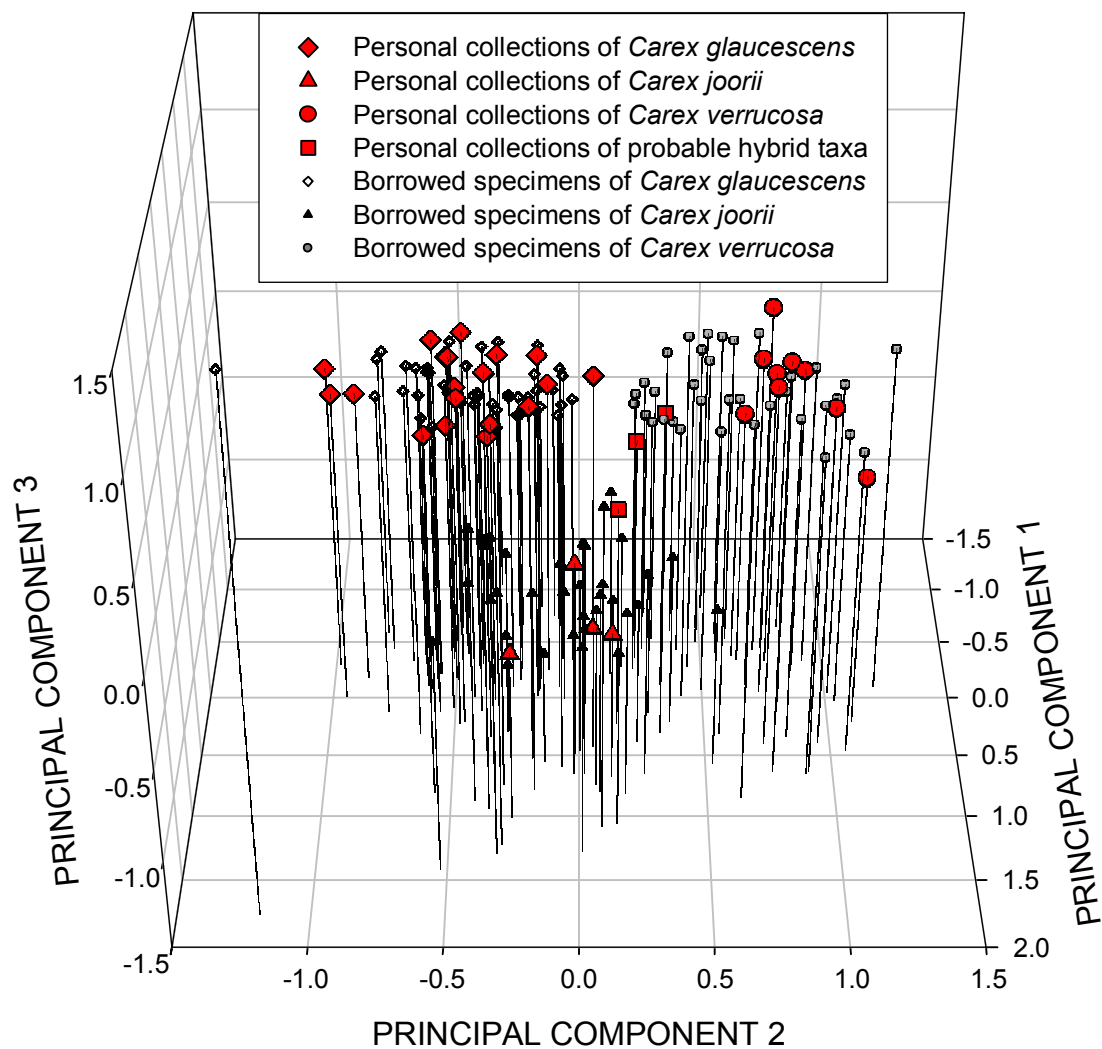


FIG. 14. The first three principal components are plotted for a PCA of borrowed herbarium specimens and personal collections, including possible hybrid taxa.

early spring, having a terminal spike of mostly pistillate spikelets and reduced peduncles.

Another possible hybrid is from *C. glaucescens* X *C. jorii*. They are suspected to produce a fertile hybrid, resembling *C. jorii*, with clavate papillose perigynia (Standley 2002). The clavate papillose specimens do not cluster separately from *C. jorii* specimens with rounded and horizontally sunken papillae. Hybridization may be a possible explanation for the clavate papillae on *C. jorii*; however this is a not consistent explanation. Many *C. jorii* specimens have clavate papillae on the beak and rounded and horizontally sunken papillae on the perigynia body. From the total borrowed herbarium specimens, there are five specimens from Arkansas with clavate papillae on the perigynia body (Table 16). It is difficult to accept these *C. jorii* specimens as hybrids because both *C. glaucescens* and *C. verrucosa* do not occur in Arkansas.

#### *Multivariate Analysis with Carex jorii* Holotype Specimen

The first three principal components for this PCA included the *C. jorii* holotype specimen explained 75.37 percent of the total variation among OTU's (Table 17). Percent total variation explained was marginally increased by adding the type specimen. The *C. jorii* type specimen corresponded to specimens labeled with the respective taxon (Fig. 18). No type specimen for *C. glaucescens* or *C. verrucosa* were located for measurement and analysis.

#### Taxa Distributions

Taxa in the section *Glaucescentes* have overlapping geographical distributions. *Carex glaucescens* is distributed in AL, FL, GA, LA, MD, MS, NC, SC, TN, TX and VA (Fig. 16). *Carex jorii* is distributed in AR, FL, GA, KY, LA, MD, MO, MS, NC, SC, TN, TX and VA (Fig. 17). *Carex verrucosa* is distributed in AL, FL, GA, LA, MS, NC, SC and

TABLE 16. Borrowed specimens of *Carex joorii* from Arkansas with clavate papillae.

HERBARIUM AND ACCESSION NUMBER		COLLECTOR	COLLECTOR NUMBER	STATE	COUNTY	COLLECTION DATE
MICH	—	<u>Steve L. Orzell</u>	<u>2646</u>	AR	BRADLEY	8 Aug 1985
MICH	—	<u>Steve L. Orzell</u>	<u>3178</u>	AR	CALHOUN	6 Oct 1985
NLU	92010	<u>David Lawson</u>	<u>2071</u>	AR	SEVIER	23 Jul 1974
NLU	287925	<u>Sundell &amp; Ethridge</u>	<u>7995</u>	AR	BRADLEY	8 Oct 1987
SFRP	5327	<u>A. Martin</u>	<u>344</u>	AR	DREW	6 Sep 1978

TABLE 17. Eigenvector coefficients from the PCA of 10 characters from borrowed herbarium specimens including type specimens. Percent trace and cumulative variation explained for the first three principal components are presented.

CHARACTER <sup>a</sup>	PC 1	PC 2	PC 3
IL	0.729	0.418	0.281
SLS	0.696	-0.072	0.336
PLLP	0.706	-0.505	0.231
LLS	0.769	0.121	0.098
LLP	0.832	-0.175	0.202
GLBK	0.512	0.019	-0.709
GWB	-0.176	0.767	0.442
GLB	-0.155	0.780	0.416
AW	0.242	0.735	-0.358
GNN	0.406	0.549	-0.620
% TRACE	33.395	25.181	16.792
COMBINED % TRACE			75.368

a Character codes are in Table 2.

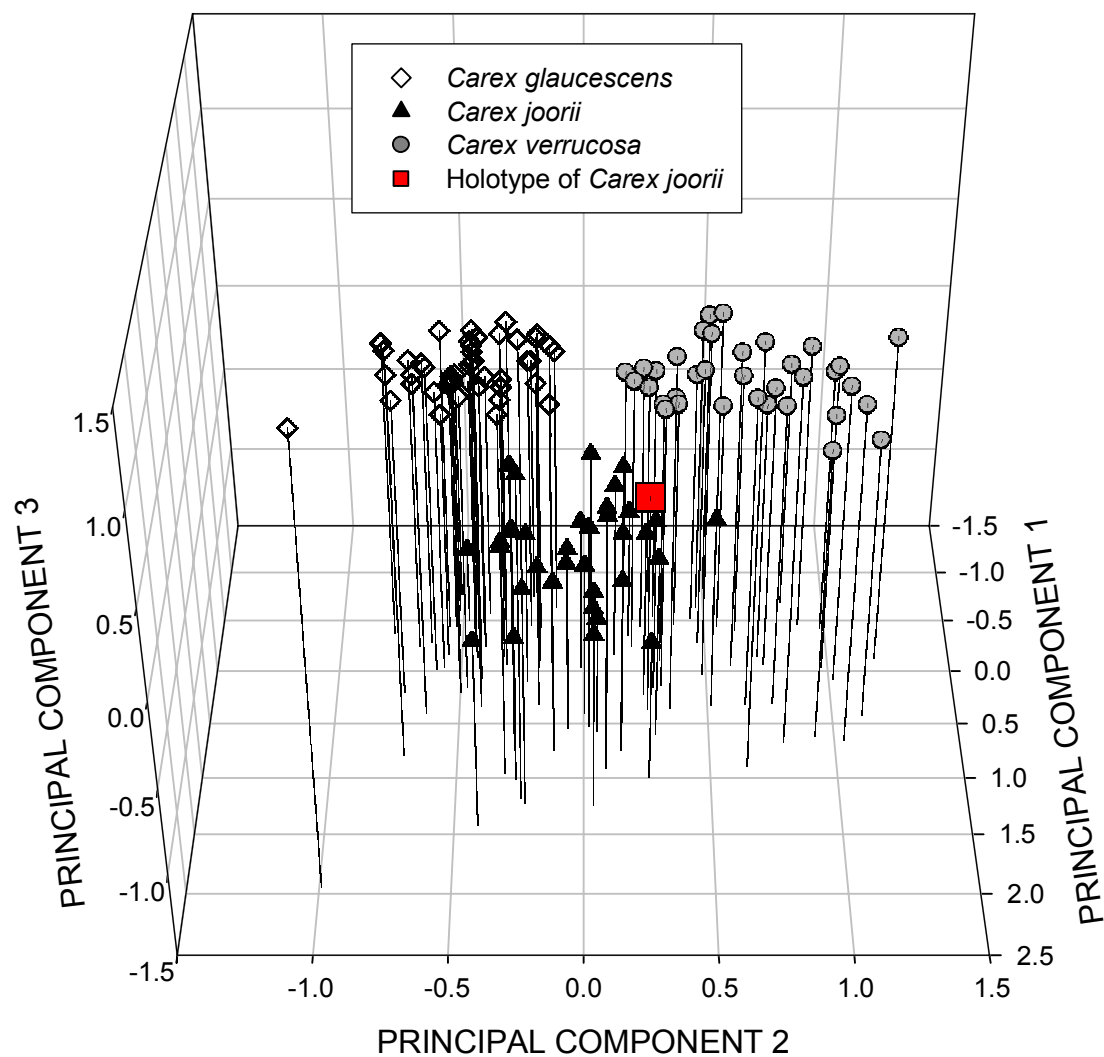


FIG. 15. The first three principal components are plotted for a PCA with 10 characters from *Carex glaucescens*, *C. jorii* and *C. verrucosa*, including the holotype of *C. jorii*.

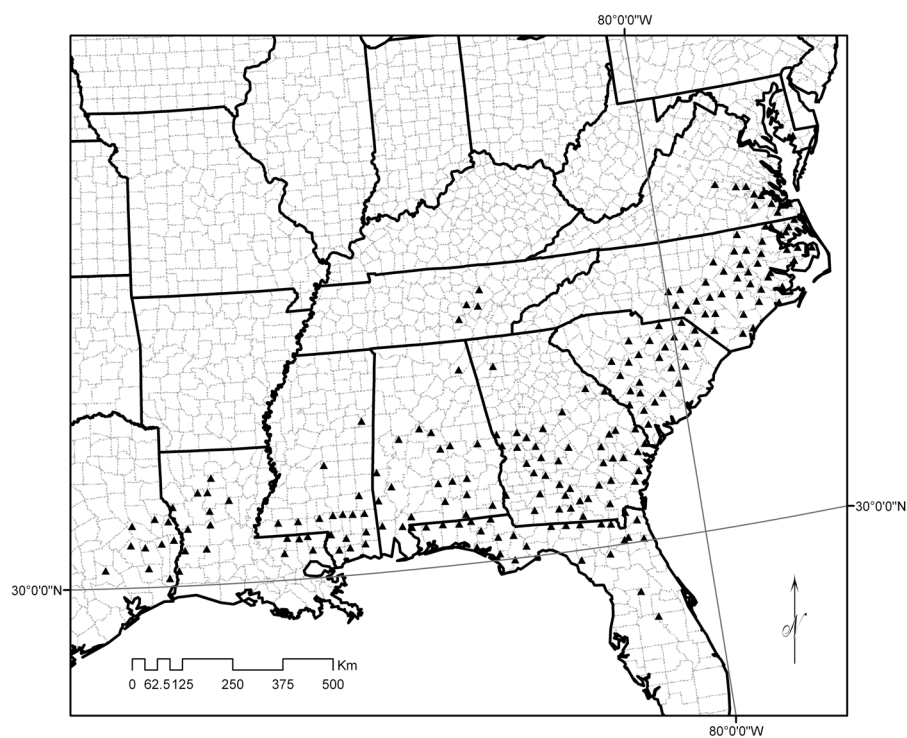


FIG. 16. County and state distribution map of *Carex glaucescens*, based on annotated herbarium specimens.

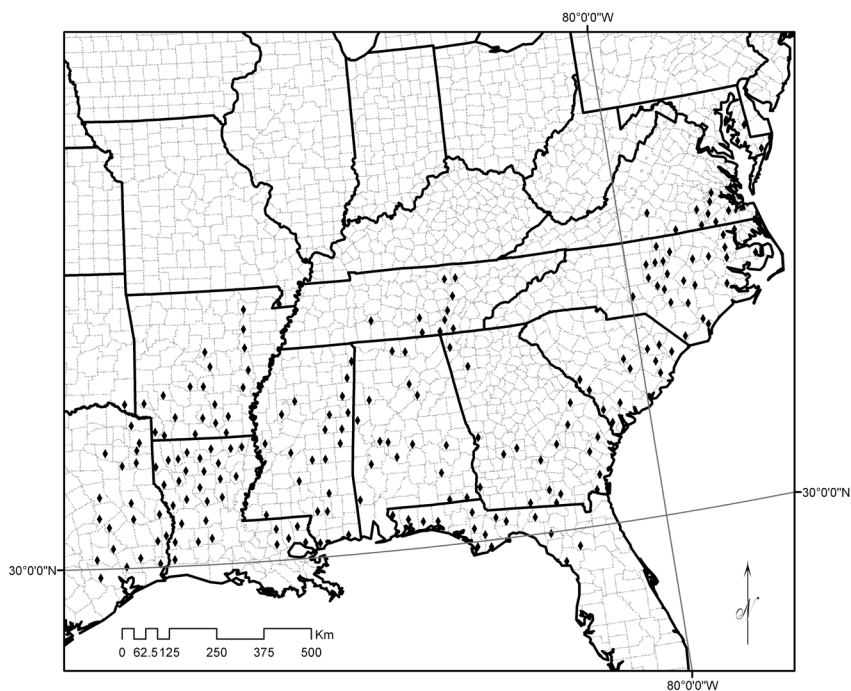


FIG. 17. County and state distribution map of *Carex jorii*, based on annotated herbarium specimens.

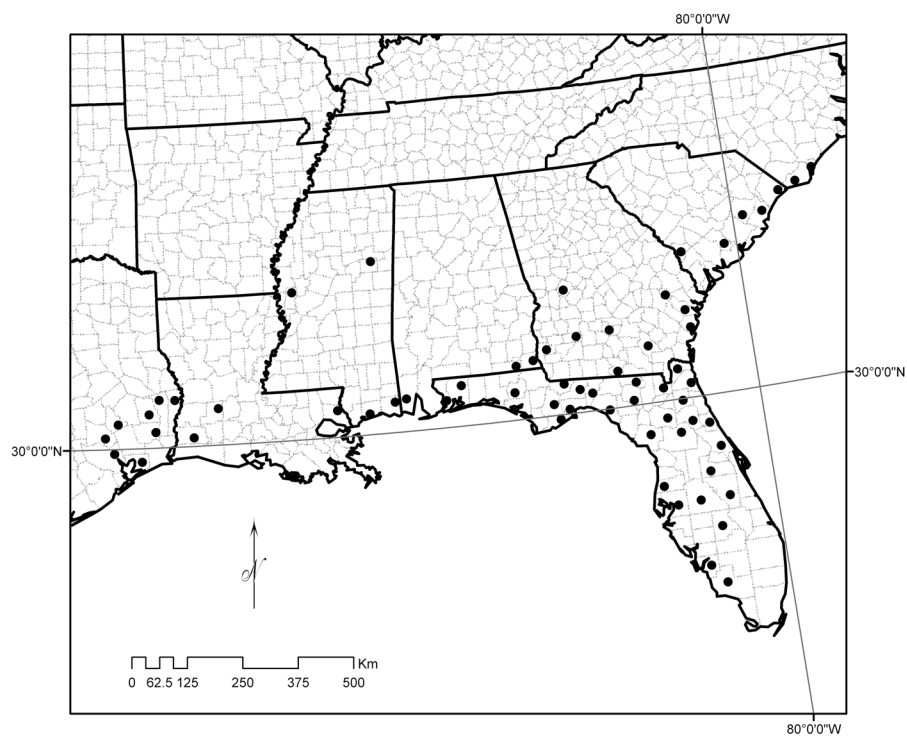


FIG. 18. County and state distribution map of *Carex verrucosa*, based on annotated herbarium specimens.



TX (Fig. 18). *Carex verrucosa* is limited to the southeastern coastal plain. *Carex glaucescens* extends north into the Piedmont and Appalachian plateaus. Distribution of *C. jorii* includes the southeastern coastal plain, Piedmont and Appalachian plateaus as well as the valley and ridge and the interior low plateau.

Results of data for allopatry and sympatry were taken from 16 populations of section *Glaucescetes* collected in Texas (Table 4). Even numbers of all taxa were not located, which may affect conclusions. *Carex glaucescens* and *C. jorii* were found to occur together without *C. verrucosa* twice and once (possibly twice) with *C. verrucosa*. Specimens found separately were *C. glaucescens* nine times, *C. jorii* once, and *C. verrucosa* three times (possibly twice). *Carex glaucescens* and *C. jorii* were the only two taxa to be found together without a third taxon present.

#### Phenology

Histograms for flowering dates by month are summarized by species in Figure 19. *Carex glaucescens* flowers from January to November, with most specimens collected during July, August and September and peaking in August. *Carex jorii* flowers from April to November, with most specimens collected during July to October and peaking during August. *Carex verrucosa* flowers from February to December with most specimens collected during March, April, May and June and peaking during April. Peak flowering times for *C. glaucescens* and *C. jorii* overlap in the summer, however the peak flowering time for *C. verrucosa* is in the spring. Most specimens of *C. verrucosa* are reproductively isolated from *C. glaucescens* and *C. jorii* by flowering time.

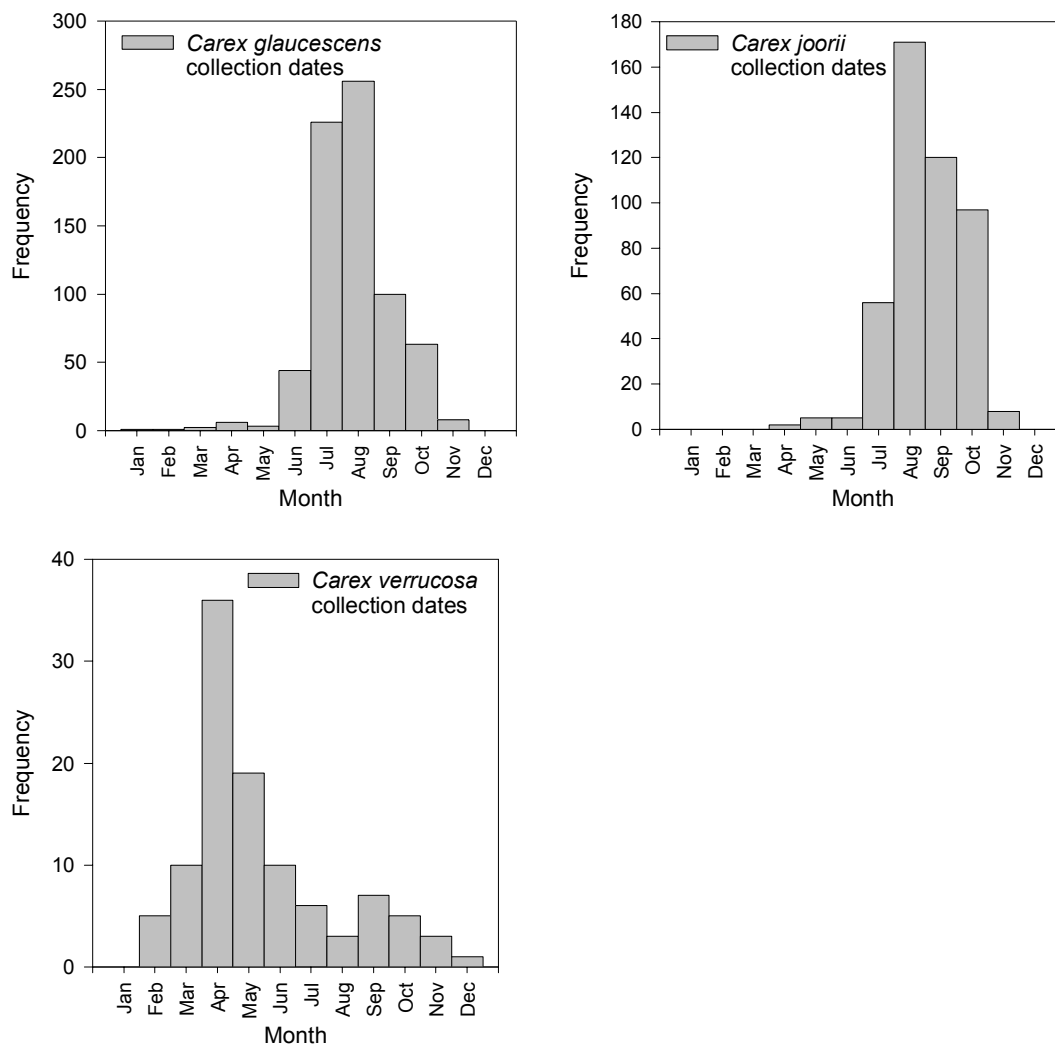


FIG. 19. Histogram of *Carex glaucescens*, *C. jorii* and *C. verrucosa* collection dates by month.

## Taxonomy

### *Type Specimens*

The *C. jorii* holotype is deposited at L. H. Bailey Hortorium Herbarium (BH). Type specimen images are in Appendix A. Holotype specimens for *C. glaucescens* and *C. verrucosa* were not located in herbaria or literature.

During the early nineteenth century it was typical not to designate a type, but to have one or two specimens on which to base descriptions. Two possibilities for typification of *C. verrucosa* were located in the literature and herbaria. First, Mackenzie (1935) noted a scrap of *C. verrucosa* as received from Muhlenberg and preserved in the Torrey Herbarium. This specimen, deposited at NY, consists merely of immature perigynia in a fragment packet. Comparisons of writing on the fragment packet label, by the author, with samples of Muhlenberg's handwriting do not appear to match. Regardless of handwriting on the packet, the specimen may still be Muhlenberg material labeled by another individual. Second, Smith (1962), while cataloging and researching Muhlenberg specimens at PH, located an Elliott specimen (an inflorescence) labeled in Muhlenberg's handwriting "215 *Carex verrucosa* ns". It was common for an author of a new species to write on labels the scientific name followed by species novum (ns, n.s., n. sp., sp. nov.). Considering these two specimens for type designation, with the conclusion that they were both examined by Muhlenberg and the determination that they are *C. verrucosa* material, it would be more prudent to accept Elliott 215 (PH) as the type specimen because of "ns" on the label and it is a more complete specimen. Further examination of correspondence between Muhlenberg and Elliott along with recommendations in the ICBN, will exclude these 2 specimens from typification.

Examination of correspondence housed at the Cambridge Harvard University Herbarium (GH) from Muhlenberg to Elliott added more information to Elliott 215 (PH). On 2 July 1810, Muhlenberg wrote a letter replying to a letter from Elliott dated 15 May 1810. Elliott had sent over 200 specimens with his letter, to Muhlenberg for exchange and identification. Muhlenberg replied to Elliott with partial and complete identifications for specimens 32 through 80 and 201 through 238. Specimen number 215 was labeled “*Carex verrucosa* Schkuhr, *flacca* affin “. This taxon described by Schkuhr was not located in the literature. It is unclear why Muhlenberg used this authority; however this name is not validly published and is therefore not an accepted authority for the taxon. Three years later, Muhlenberg (1813) published a vascular plant checklist for North America, in which he listed *C. verrucosa* without an authority. Muhlenberg’s (1817) description of *C. verrucosa* validly published the taxon with him as the authority. Elliott’s (1824) description of *C. verrucosa*, stated that he collected a specimen many years prior in southern South Carolina between Stono and the Combahee Ferry (inferring collection either in Charleston, Dorchester, Colleton or Hampton counties). The specimen mentioned is most likely the specimen mentioned in correspondence from Muhlenberg to Elliott (Elliott 215 (PH)).

It is possible that Elliott 215 (PH) is a specimen on which Muhlenberg based the description of *C. verrucosa*. The type description by Muhlenberg (1817) describes *C. verrucosa* as having five lateral spikes, however the Elliott 215 (PH) specimen has six lateral spikes. This discrepancy makes it problematic to conclude that Muhlenberg based the *C. verrucosa* description on Elliott 215 (PH). It would not be a stretch to lectotypify Elliott 215 (PH), if Muhlenberg had described *C. verrucosa* with a range for spike number.

Elliott 215 (PH) is a valuable specimen because it is labeled as *C. verrucosa* by Muhlenberg. The *C. verrucosa* type description does not distinctly separate it from *C. glaucescens*. The specimen is useful in that it indicates what Muhlenberg described as *C. verrucosa*. The Elliott 215 (PH) specimen will not be lectotypified for *C. verrucosa* within this manuscript.

*Artificial Dichotomous Key to Carex Section Glaucescences in North America*

- 1a. Pistillate scale tapered into an awn, sometimes minutely emarginate; perigynia inflated and abruptly narrowing into a beak 0.7 to 1.4 mm long, veins prominent with (12-) 14-18 (-20). (Fig. 20) \_\_\_\_\_ ***C. joorii***
- 1b. Pistillate scale emarginate to retuse at awn base; perigynia not inflated and nearly beakless at 0.2 to 0.5 mm long (some *C. glaucescens* may be consider longer), veins prominent to somewhat obscure with 2-12.
  - 2a. Perigynia rhombic, pedicel surrounded with perigynia tissue, 0.2-0.5 mm long, 0.2-0.5 mm wide; lowermost pistillate spike peduncles are generally erect. (Fig. 20) \_\_\_\_\_ ***C. verrucosa***
  - 2b. Perigynia elliptic to ovate, pedicel not surrounded with perigynia tissue, 0.1-0.2 mm long, 0.1-0.2 mm wide; lowermost pistillate spike peduncles are generally drooping. (Fig. 20) \_\_\_\_\_ ***C. glaucescens***

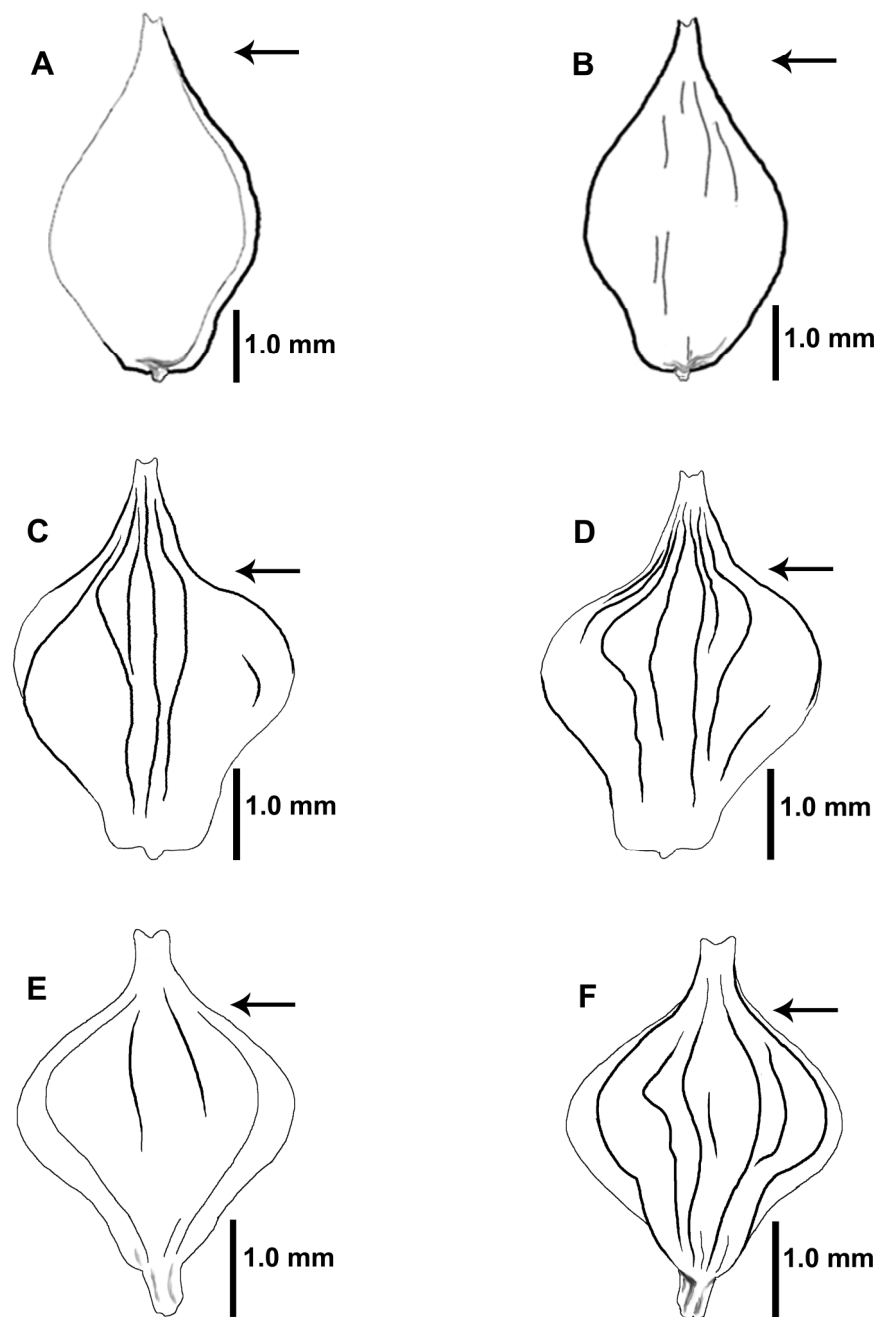


FIG. 20. Line drawings of perigynia from North American taxa of *Carex* section *Glaucescentes*. Arrows indicate beginning of beak. Dark longitudinal lines represent prominent veins. *C. glaucescens*: A. Abaxial view, B. Adaxial view, *C. jorii*: C. Abaxial view, D. Adaxial view, *C. verrucosa*: E. Abaxial view, F. Adaxial view.

### *Species Descriptions*

***Carex glaucescens*** Elliott, Sketch Bot. S. Carolina. 2: 553. 1824. TYPE not designated.

*Carex glaucescens* Elliott var *polystachys* M.A. Curtis, Amer. J. Sci. Arts 57: 410. 1849.

*Carex inundata* Willd. ex Schlttdl., Linnaea 10: 267. 1835.

*Carex rufidula* Steud., Syn. Pl. Glumac. 2: 220. 1885.

*Carex sempervirens* Schwein., Ann. Lyceum Nat. Hist. New York 1: 70. 1824, non Villars 1787.

*Carex verrucosa* Muhl. var. *glaucescens* A.W. Wood, Amer. Bot. Fl., ed. 2. 379. 1871.

*Edritria glaucescens* (Elliott) Raf., Good Book 26. 1840. Publication not viewed by author.

Robust perennials forming cespitose clumps. **Rhizomes** short, scaly and brown to black. **Culms** (58-) 79-114 (-150) cm tall, erect. **Leaves:** Abaxial sheath surfaces many veined; Adaxial sheath surfaces membranous, red to brown dotted, apically acute to rounded and sometimes fibrous splitting centrally; Blades extending beyond terminal spike, (2.4-) 3.8-6.0 (-7.1) mm wide; Abaxial blade surface papillose glaucous with one prominent central vein; Adaxial blade surface scabrous with many veins and sometimes 2 distinct veins forming a **M**-cross section; Blade margins revolute and antrorsely serrulate. **Ligules** minute, membranous, apically v-shaped and turning brown. **Inflorescences** (7-) 10-18 (-29) cm long; **Bracts** variable in length, but progressively

shorter proximally to distally, generally the lowermost spike bract extends beyond the terminal spike, bracts with minute to erect rounded or acute appendages proximally.

**Terminal spikes** assurgent to the rachis, 1 (-3) including reduced spikes, frequently staminate, rarely with pistillate spikelets, (1.3-) 2.8-5.4 (-7.7) cm long, (4.1-) 4.7-7.1 (-8.9) mm wide; peduncle 0.3-11.9 (-30+) cm long; staminate scales (1.3-) 5.0-7.6 (-8.5) mm long, 1.0-2.2 (-5.0) mm wide, apically emarginate to retuse with rounded to acute lobes, oblong, 3-veined with central vein extending into an antrorsely barbed awn.

**Lateral spikes** pendulous to the rachis, 3-5 (-7) including lowermost pistillate spike, androgynecandrous or gynecandrous (0-numerous sterile or staminate distal spikelets, many central pistillate spikelets and (0-) 1-3 staminate proximal spikelets); peduncles drooping. *Uppermost pistillate spikes* (0.4-) 1.9-3.6 (-4.2) cm long, peduncles (0-) 0.4-2.7 (-6.2) cm long. *Second lowermost pistillate spikes* (2.1-) 2.8-4.6 (-6.3) cm long, peduncles (0.0-) 0.7-3.9 (-9.8) cm long. *Lowermost pistillate spikes* (2.2-) 2.7-4.4 (-5.6) cm long, peduncles (0.5-) 1.3-5.2 (-12.2) cm long, (0.2-) 0.3-0.5 (-0.6) mm wide.

**Pistillate scales** apically emarginate to retuse with rounded to acute lobes, oblong, 3-veined with central vein extending into an (0.5-) 0.9-2.0 (-3.3) mm long antrorsely barbed awn, scale body shorter than perigynia, scale (2.5-) 3.5-4.8 (-6.1) mm long, (1.0-) 1.1-1.7 (-2.0) mm wide; **Perigynia** ascending to rarely divergent, elliptic to ovate, (3.0-) 3.4-4.1 (-4.7) mm long, (1.8-) 2.0-2.4 (-2.5) mm wide; Beak (0.2-) 0.3-0.5 (-0.7) mm long; Surfaces with clavate to terete and sometimes longitudinally troughed papillae; Faces and angles with 2-6 (-10) mostly indistinct veins including 2 distinct lateral veins (suture lines) extending into a bidentate beak; pedicel 0.1-0.2 mm long, 0.1-0.2 mm wide. **Achenes** triquetrous, obtrullate, (1.9-) 2.3-2.8 (-3.0) mm long, (1.7-)



1.8-2.1 (-2.3) mm wide; Surface of nonisodymetric epidermal cells with a central silica body; Style deciduous.

**Phenology:** (Jan-) Jul-Sep (-Nov).

**Distribution:** AL, FL, GA, LA, MD, MS, NC, SC, TN, TX and VA (Fig. 16).

Distribution of *Carex glaucescens* is along the southeastern Gulf coastal plains, north and east into the Piedmont province and Appalachian plateau and northeastward into the Atlantic coast plain.

**Habitat:** margins of streams, shallow ponds, logged pine plantations, roadside ditches, seasonally wet flatwood ponds and seasonally wet meadows; sandy, loamy and clay soils.

***Carex jorii*** Bailey, Proc. Amer. Acad. Arts. 22(1): 72, 1886. TYPE: U.S.A., LA: Comite Swamp, near Baton Rouge, J.F. Joor s.n., 5 Aug 1885: (HOLOTYPE: BH!).

Robust perennials forming cespitose clumps. **Rhizomes** short, sometimes curved, scaly and brown to black. **Culms** (23-) (62-99) (-119) cm tall, erect. **Leaves:** Abaxial sheath surfaces many veined; Adaxial sheath surfaces membranous, red to brown dotted, apically acute to rounded and sometimes fibrous; Blades extending beyond terminal spike, (2.9-) 3.9-5.3 (-5.5) mm wide, Abaxial blade surface papillose glaucous to somewhat glabrate with one prominent vein, Adaxial blade surface scabrous to smooth with many veins and sometimes 2 distinct veins forming a **M**-cross section; Blade margins revolute and antrorsely serrulate. **Ligules** minute, membranous, apically v-shaped and turning brown. **Inflorescences** (7-) 13-24 (-29) cm long, **Bracts** variable in length, but progressively shorter proximally to distally, generally

the lowermost spike bract extends beyond the terminal spike, bracts with minute to erect rounded or acute appendages. **Terminal spikes** assurgent to the rachis, 1 (-3) including reduced spikes, staminate, rarely with pistillate spikelets, (2.0-) 3.1-5.4 (-6.0) cm long, (2.5-) 3.4-5.9 (-9.5) mm wide; peduncle (3-) 1.5-4.6 (-6.2) cm long; staminate scales (4.1-) 5.1-7.5 (-9.1) mm long, (1.1-) 1.3-1.6 (-1.8) mm wide apically acute, ovate to oblong, 3-veined with central vein extending into an antrorsely barbed awn. **Lateral spikes** pendulous to the rachis, 4-7 spikes including lowermost pistillate, androgynecandrous or rarely gynecandrous (0-numerous sterile or staminate distal spikelets, many central pistillate spikelets and (0-) 1-3 staminate proximal spikelets); Peduncles drooping. *Uppermost pistillate spikes* (0.8-) 1.6-3.4 (-5.5) cm long; peduncles 0.0-1.0 (-2.7) cm long. *Second lowermost pistillate spikes* (2.3-) 2.9-4.9 (-6.7) cm long; peduncles (0.6-) 1.2-3.6 (-5.3) cm long. *Lowermost pistillate spikes* (2.6-) 3.2-5.3 (-6.5) cm long; staminate distal (0-) 0.8-7.3 mm long; peduncles (2.0-) 2.6-6.2 (-7.8) cm long, 0.3-0.5 (-0.6) mm wide. **Pistillate scales** apically acute, ovate, 3-veined with central vein extending into an antrorsely barbed awn (0.3-) 0.7-1.5 (-1.6) mm long, scale body shorter than perigynia, scale (2.5-) 3.3-4.5 (-5.3) mm long, (1.2-) 1.3-1.7 (-2.1) mm wide. **Perigynia** squarrose, rhombic, (3.4-) 3.6-4.3 (-4.9) mm long, (2.1-) 2.5-2.9 (-3.1) mm wide; Beak (0.7-) 0.8-1.1 (-1.4) mm long; Surfaces with concave reticulate with absent, clavate or rounded papillae, generally with clavate papillae apically and rounded to absent papillae below beak; Faces and angles with 13-16 distinct veins, including 2 suture lines which sometimes extend to form a bidentate beak; Pedicel (0.0-) 0.1-0.1 (-0.3) mm wide, (0.0) 0.1-0.1 (-0.2) mm long. **Achenes** triquetrous, obtrullate, (2.3-) 2.5-2.8 (-2.9) mm long, (1.9-) 2.2-2.6 (-2.8) mm wide; Surface of nonisodymetric epidermal cells with a central silica body; Style deciduous.

**Phenology:** (Apr-) Jul-Oct (-Nov).

**Distribution:** AR, FL, GA, KY, LA, MD, MO, MS, NC, SC, TN, TX and VA (Fig. 17).

Distribution of *C. jorii* is along the southeastern coastal plains, north and east into the unglaciated plains, the Piedmont province, the Appalachian plateau, the valley and ridge province, the interior low plateau and eastward into the Atlantic coast plain.

**Habitat:** margins of streams, shallow ponds, logged pine plantations, roadside ditches, seasonally wet flatwood ponds and seasonally wet meadows; sandy, loamy and clay soils.

***Carex verrucosa*** Muhl., Descr. Gram. 261. 1817. TYPE: not designated. Potential specimen for lectotypification by author: Elliott 215 (BH).

*Carex glaucescens* Elliott var. *androgyna* M.A. Curtis, Amer. J. Sci. Arts 44:84. 1843.

*Carex verrucosa* Muhl. var. *androgyna* (M.A. Curtis) Dewey, Amer. J. Sci. Arts 48: 140. 1845.

*Carex macrokolea* Steud., Syn. Pl. Glumac. 2: 223. 1855.

*Carex brasiliensis* A. St.-Hil. var. *gracilis* Boeck., Linnaea 41: 292. 1877.

*Carex glaucescens* Ell. forma *macrokolea* (Steud.), Kükenthal in Engler, Pflanzenr. 4 (38): 733. 1909.

Robust perennials forming cespitose clumps. **Rhizomes** short, scaly, brown to black. Culms (63-) (80-119 (-157) cm tall, erect. **Leaves** basal, extending beyond the terminal spike, (3-) (4.2-6.8) (-8) mm wide, abaxial surface papillose glaucous with one

prominent central vein, adaxial surface scabrous with many veins, margins revolute and barbed. **Abaxial sheath surfaces** many veined. **Adaxial sheath surfaces** membranous, red to brown dotted, apex rarely intact, fibrous. **Ligules** minute, membranous, apically acute apically v-shaped and turning brown. **Inflorescences** (8-) 12-28 (-34) cm long, **Bracts** variable in length, progressively shorter from base to apex of inflorescence, generally the lowermost spike bract extends beyond the terminal spike. **Auricles** erect and emarginate to rarely tapered. **Terminal spikes**, staminate or gynecandrous and rarely androgynecandrous or solely pistillate, 2.7-5.0 cm long, 4.2-10.7 mm wide; peduncle 0.31-14.4 (-30.8) cm long; staminate scales 5.3-8.0 mm long, 1.1-1.9 mm wide. **Lateral spikes** androgynecandrous or sometimes gynecandrous (0-numerous distal staminate spikelets, many central pistillate spikelets and (0-) 1-3 proximal staminate spikelets), 3-7 spikes including lowermost pistillate, peduncles erect. *Uppermost pistillate spikes* (0.8-) 1.4-2.9 (-3.7) cm long, peduncles 0-0.5 (-1.2) cm long. *Second lowermost pistillate spikes* (1.5-) 2.6-4.5 (-5.2) cm long, peduncles (0.2-) 0.6-1.7 (-2.8) cm long. *Lowermost pistillate spikes* (2.0-) 2.8-4.7 (-6.1) cm long, peduncles (0.7-) 1.1-4.7 (-8.4) cm long, (0.3-) 0.4-0.6 (-0.7) mm wide, staminate distal 0-1.5 (-5.9) mm long. **Pistillate scales** emarginate to retuse with rounded to acute lobes, 3-veined with central vein extending into an awn, scale body shorter than perigynia, scale (3.6-) 3.9-5.7 (-7.0) mm long, 1.1-2.0 (-4.1) mm wide; awn antrorsely barbed, (4.0-) 1.1-2.0 (-3.7) mm long. **Perigynia** rhombic, papillose, (3.2-) 3.4-4.2 (-4.6) mm long, (2.2-) 2.4-2.7 (-3.1) mm wide, beak length (0.1-) 0.2-0.6 (-0.9) mm long, veins faint, (4-) 8-14 (-16); pedicel 0.2-0.4 (-0.5) mm long, 0.2-0.4 (-0.5) mm wide. **Achenes** triquetrous, obtrullate, (2.2-) 2.8-2.8 (-3.2) mm long, (2.1-) 2.3-2.6 (-3.0) mm; Surface of nonisodymetric epidermal cells with a central silica body; Style deciduous.

**Phenology:** (Feb-) Mar-May (-Dec).

**Distribution:** AL, FL, GA, LA, MS, NC, SC and TX (Fig. 18). Distribution of *Carex verrucosa* is along the Gulf coast plains eastward into the Atlantic coast plain.

**Habitat:** shallow ponds, roadside ditches and flatwood ponds; sandy, loamy and clay soils.

## CONCLUSIONS

Currently accepted North America taxa in *Carex* section *Glaucescentes* should retain species rank. The taxa are separated to species by morphology, geographical distribution, phenology, statistical analysis and hybridization characteristics. Qualitative and quantitative morphological characters that delimit the three taxa are presented in an artificial dichotomous key and species descriptions. Some previously used characters in current keys (Godfrey & Wooten 1979; Standley 2002), were found to separate the taxa when wording or ranges were changed. Other characters were found to inconsistently separate the taxa. The species descriptions and artificial dichotomous key produced from this manuscript should increase the accuracy of identification for these taxa.

Specifically, *C. jorii* is separated from the other taxa by having inflated, squarrose perigynia, 12 or more prominent veins on the perigynia and perigynia beak length, wider leaves and a tapered scale. *Carex verrucosa* has erect peduncles (rarely the lowermost spike drooping), rhombic, but not inflated perigynia and a pedicel surrounded with perigynia tissue that is generally wider and longer than that of *C. jorii* and *C. glaucescens*. *Carex glaucescens* has elliptic to ovate and longer than wide perigynia.

Geographical distributions overlap within the coastal plain region for all three taxa, with *C. verrucosa* extending furthest south. *Carex jorii* has the widest distribution, followed by *C. glaucescens* and finally *C. verrucosa* with the narrowest distribution. *Carex jorii* and *C. glaucescens* are both late summer to early fall flowering species, which differentiates them from the spring flowering species *C. verrucosa*.

PCA separated the taxa into 3 clusters, which align with the accepted taxa, *C. glaucescens*, *C. jorii* and *C. verrucosa* based on type descriptions, *Carex jorii* and *C. verrucosa* should be considered separate species because they hybridize to produce infertile offspring.

The holotype specimen for *C. jorii* was located. Muhlenberg and Elliott did not designate type specimens for *C. verrucosa* or *C. glaucescens*, respectively, and one was not found in herbaria.

Further research for the section should include thorough sampling from sympatric populations containing all three taxa, along with evaluation of reproductive methods. Morphological characters that should be further evaluated include length of the leaf sheath and rhizome inner node length. These characters may be of little taxonomic value, however I suspect that *C. verrucosa* has longer sheaths and that *C. jorii* has longer inner-nodes.

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## APPENDIX A

TYPE IMAGES OF *CAREX JOORII*

## NORTH AMERICAN CARICES.

L. H. BAILEY, Jr.

*C. Joori* n. sp.*Carnite Swamp, near  
Baton Rouge, La.**J. F. Joor. Aug. 5, 1883-*







## APPENDIX B

ASSOCIATED SPECIES OF *CAREX* SECTION *GLAUCESCENTES*

SPECIES	AUTHORITY	SOURCE/s
1 <i>Acer rubrum</i> var. <i>drummondii</i>	L., (Hook. & Arn. ex Nutt.) Sarg.	Weakley et al. 1998
2 <i>Acer rubrum</i> var. <i>rubrum</i>	L.	Weakley et al. 1998
3 <i>Alnus serrulata</i>	(Dryand ex Ait.) Willd.	Weakley et al. 1998
4 <i>Amsonia glaberrima</i>	Woods.	Weakley et al. 1998
5 <i>Amsonia tabernaemontana</i>	Walt.	D.A. Kruse 812
6 <i>Andropogon capillipes</i>	Nash	Weakley et al. 1998
7 <i>Andropogon glomeratus</i>	(Walt.) B.S.P.	Weakley et al. 1998
8 <i>Andropogon virginicus</i>	L.	Weakley et al. 1998
9 <i>Aristida palustris</i>	(Chapm.) Vasey	Weakley et al. 1998
10 <i>Aronia arbutifolia</i>	(L.) Ell.	Weakley et al. 1998
11 <i>Arundinaria gigantea</i>	(Walt.) Muhl.	Weakley et al. 1998
12 <i>Asclepias perennis</i>	Walt.	Weakley et al. 1998
13 <i>Bacopa caroliniana</i>	(Walt.) Robins.	Weakley et al. 1998
14 <i>Bartonia virginiana</i>	(L.) B.S.P.	Weakley et al. 1998
15 <i>Berchemia scandens</i>	(Hill) K. Koch	Weakley et al. 1998
16 <i>Bidens discoidea</i>	(T. & G.) Britt.	Weakley et al. 1998
17 <i>Bignonia capreolata</i>	L.	Weakley et al. 1998
18 <i>Boehmeria cylindrica</i>	(L.) Sw.	Weakley et al. 1998
19 <i>Botrychium dissectum</i>	Spreng.	Weakley et al. 1998
20 <i>Brasenia schreberi</i>	J. F. Gmel.	Weakley et al. 1998
21 <i>Brunnichia ovata</i>	(Walt.) Shinnars	Weakley et al. 1998
22 <i>Cabomba caroliniana</i>	Gray	Weakley et al. 1998
23 <i>Calamagrostis coarctata</i>	(Torr.) Eat.	Weakley et al. 1998
24 <i>Cardiospermum halicacabum</i>	L.	Weakley et al. 1998
25 <i>Carex caroliniana</i>	Schwein.	D.C. McLaughlin



26	<i>Carex crinita</i>	Lam.	Weakley et al. 1998
27	<i>Carex festucacea</i>	Willd.	D.C. McLaughlin
28	<i>Carex flaccosperma</i>	L. H. Dewey	D.C. McLaughlin
29	<i>Carex frankii</i>	Kunth.	D.C. McLaughlin 506
30	<i>Carex gigantea</i>	Rudge	Weakley et al. 1998
31	<i>Carex intumescens</i>	Rudge	Weakley et al. 1998
32	<i>Carex lonchocarpa</i>	Willd.	Weakley et al. 1998
33	<i>Carex striata</i>	Michx.	Weakley et al. 1998
34	<i>Carex triangularis</i>	Boeck.	D.C. McLaughlin
35	<i>Carex turgescens</i>	Torr.	Weakley et al. 1998
36	<i>Carex venusta</i> var. <i>minor</i>	Dewey, Boeckl.	Weakley et al. 1998
37	<i>Carpinus caroliniana</i>	Walt.	Weakley et al. 1998
38	<i>Carya alba</i>	(Miller) K. Koch	Weakley et al. 1998
39	<i>Carya aquatica</i>	(Michx. f.) Nutt.	Weakley et al. 1998
40	<i>Carya glabra</i>	(Mill.) Sweet	Weakley et al. 1998
41	<i>Carya ovata</i>	(Miller) K. Koch	Weakley et al. 1998
42	<i>Cephalanthus occidentalis</i>	L.	D.C. McLaughlin, Weakley et al. 1998
43	<i>Chasmanthium latifolium</i>	(Michx.) Yates	Weakley et al. 1998
44	<i>Chasmanthium ornithorhynchum</i>	(Steud.) Yates	Weakley et al. 1998
45	<i>Chasmanthium sessiliflorum</i>	(Poir.) Yates	Weakley et al. 1998
46	<i>Chionanthus virginicus</i>	L.	Weakley et al. 1998
47	<i>Cladium jamaicense</i>	Crantz.	D.A. Kruse 489
48	<i>Clethra alnifolia</i>	L.	Weakley et al. 1998
49	<i>Cliftonia monophylla</i>	(Lam.) Sarg.	Weakley et al. 1998
50	<i>Coreopsis linifolia</i>	Nutt.	Weakley et al. 1998
51	<i>Coreopsis nudata</i>	Nutt.	Weakley et al. 1998

52	<i>Cornus florida</i>	L.	Weakley et al. 1998
53	<i>Cornus foemina</i>	Mill.	Weakley et al. 1998
54	<i>Crataegus marshallii</i>	Eggston	Weakley et al. 1998
55	<i>Crataegus viridis</i>	L.	Weakley et al. 1998
56	<i>Cyperus erythrorhizos</i>	Muhl.	D.C. McLaughlin 483, Weakley et al. 1998
57	<i>Cyperus haspan</i>	L.	D.C. McLaughlin 485
58	<i>Cyrilla parvifolia</i>	Raf.	Weakley et al. 1998
59	<i>Cyrilla racemiflora</i>	L.	Weakley et al. 1998
60	<i>Dichantherium boscii</i>	(Poir.) Gould & C.A. Clark	Weakley et al. 1998
61	<i>Dichantherium longiligulatum</i>	(Nash) Freckmann	D.C. McLaughlin
62	<i>Dichantherium scabriusculum</i>	(Ell.) Gould & C.A. Clark	Weakley et al. 1998
63	<i>Diodia virginiana</i>	L.	D.C. McLaughlin 438
64	<i>Dulichium arundinaceum</i>	(L.) Britt.	Weakley et al. 1998
65	<i>Echinodorus cordifolius</i>	(L.) Griseb.	Weakley et al. 1998
66	<i>Eleocharis equisetoides</i>	(Ell.) Torr.	Weakley et al. 1998
67	<i>Eleocharis melanocarpa</i>	Torr.	Weakley et al. 1998
68	<i>Eleocharis microcarpa</i>	Torr.	D.C. McLaughlin 402B, D.A. Kruse 803, Weakley et al. 1998
69	<i>Eleocharis montana</i>	(Kunth in H.B.K.) Roem. & Schult.	D.A. Kruse 597
70	<i>Eleocharis montevidensis</i>	Kunth.	D.A. Kruse 494 , 823
71	<i>Eleocharis quadrangulata</i>	(Michx.) Roem. & Schult.	D.C. McLaughlin 753, Weakley et al. 1998
72	<i>Eleocharis tuberculosa</i>	(Michx.) R.&S.	D.C. McLaughlin 414, D.A. Kruse 818, 500, Weakley et al. 1998
73	<i>Eleocharis vivipara</i>	Link.	D.A. Kruse 596
74	<i>Elephantopus carolinianus</i>	Raeusch.	Weakley et al. 1998
75	<i>Erigeron vernus</i>	(L.) Torr. & Gray	Weakley et al. 1998
76	<i>Eriocaulon compressum</i>	Lam.	Weakley et al. 1998

77	<i>Eriocaulon decangulare</i>	L.	Weakley et al. 1998
78	<i>Eupatorium leptophyllum</i>	DC.	Weakley et al. 1998
79	<i>Eupatorium mohrii</i>	Greene	Weakley et al. 1998
80	<i>Eupatorium semiserratum</i>	DC.	Weakley et al. 1998
81	<i>Euthamia leptcephala</i>	(T. & G.) Greene	Weakley et al. 1998
82	<i>Euthamia tenuifolia</i>	(Pursh) Nutt.	Weakley et al. 1998
83	<i>Fagus grandifolia</i>	Ehrh.	Weakley et al. 1998
84	<i>Fimbristylis autumnalis</i>	(L.) R.&S.	D.C. McLaughlin 496
85	<i>Fraxinus caroliniana</i>	Mill.	Weakley et al. 1998
86	<i>Fraxinus profunda</i>	(Bush) Bush	Weakley et al. 1998
87	<i>Fuirena bushii</i>	Kral	Weakley et al. 1998
88	<i>Gaylussacia mosieri</i>	Small	Weakley et al. 1998
89	<i>Gelsemium rankinii</i>	(L.) Ait. f.	Weakley et al. 1998
90	<i>Gratiola brevifolia</i>	Raf.	D.C. McLaughlin, 403, 432, Weakley et al. 1998
91	<i>Hamamelis virginiana</i>	L.	Weakley et al. 1998
92	<i>Helenium drummondii</i>	H. Rock	Weakley et al. 1998
93	<i>Heliotropium indicum</i>	L.	Weakley et al. 1998
94	<i>Hibiscus moscheutos</i>	L.	Weakley et al. 1998
95	<i>Hydrocotyle spp.</i>	L.	Weakley et al. 1998
96	<i>Hydrolea ovata</i>	Choisy	D.C. McLaughlin 488, Weakley et al. 1998
97	<i>Hypericum brachyphyllum</i>	(Spach) Steud.	Weakley et al. 1998
98	<i>Hypericum chapmanii</i>	P. Adams	Weakley et al. 1998
99	<i>Hypericum galioides</i>	Lam.	Weakley et al. 1998
100	<i>Hypericum hypericoides</i>	L.	Weakley et al. 1998
101	<i>Hyptis alata</i>	(Raf.) Shinnars	Weakley et al. 1998
102	<i>Ilex coriacea</i>	(Pursh.) Chapm.	Weakley et al. 1998

103	<i>Ilex decidua</i>	Walt.	Weakley et al. 1998
104	<i>Ilex glabra</i>	(L.) Gray	Weakley et al. 1998
105	<i>Ilex myrtifolia</i>	Walt.	Weakley et al. 1998
106	<i>Ilex verticillata</i>	(L.) Gray	Weakley et al. 1998
107	<i>Ilex vomitoria</i>	Soland. In Ait.	Weakley et al. 1998
108	<i>Isoetes melanopoda</i>	J. Gay & Durieu ex Durieu	D.C. McLaughlin, D.A. Kruse 826
109	<i>Itea virginica</i>	L.	Weakley et al. 1998
110	<i>Juncus brachycarpus</i>	Engelm. In A. Gray	D.A. Kruse 822
111	<i>Juncus difusissimus</i>	Buckl.	D.C. McLaughlin 413, D.A. Kruse 811
112	<i>Juncus effusus</i>	L.	D.A. Kruse 808, Weakley et al. 1998
113	<i>Juncus elliotii</i>	Coville	D.A. Kruse 821
114	<i>Juncus marginatus</i>	Rostk.	D.C. McLaughlin 435, Weakley et al. 1998
115	<i>Juncus nodatus</i>	Cov.	D.C. McLaughlin 402A
116	<i>Juncus polycephalus</i>	Michx.	D.C. McLaughlin 433, Weakley et al. 1998
117	<i>Juncus repens</i>	Michx.	D.C. McLaughlin 408, Weakley et al. 1998
118	<i>Juncus tenuis</i>	Willd.	D.A. Kruse 824, 810
119	<i>Juncus validus</i>	Cov.	Weakley et al. 1998
120	<i>Justicia ovata</i>	(Walt.) Lindau	D.C. McLaughlin 406, Weakley et al. 1998
121	<i>Lachnocaulon beyrichianum</i>	Sporleder ex Koern.	Weakley et al. 1998
122	<i>Lauchnocaulon anceps</i>	(Walt.) Morong.	D.A. Kruse 492
123	<i>Leersia hexandra</i>	Sw.	D.C. McLaughlin 486, D.A. Kruse 596, Weakley et al. 1998
124	<i>Leersia lenticularis</i>	Michx.	Weakley et al. 1998
125	<i>Leersia oryzoides</i>	(L.) Sw.	Weakley et al. 1998
126	<i>Lemna minor</i>	L.	Weakley et al. 1998
127	<i>Leucothoe racemosa</i>	(L.) DC.	Weakley et al. 1998

128	<i>Limnobiium spongia</i>	(Bosc) L. C. Rich. ex Steud.	Weakley et al. 1998
129	<i>Liquidambar styraciflua</i>	L.	D.C. McLaughlin, Weakley et al. 1998
130	<i>Ludwigia microcarpa</i>	Michx.	Weakley et al. 1998
131	<i>Ludwigia pilosa</i>	Walt.	Weakley et al. 1998
132	<i>Ludwigia sphaerocarpa</i>	Ell.	Weakley et al. 1998
133	<i>Ludwigia suffruticosa</i>	Walt.	Weakley et al. 1998
134	<i>Lycopodiella alopecuroides</i>	(L.)	Weakley et al. 1998
135	<i>Lycopodiella appressa</i>	(Chapman) Cranfill	Weakley et al. 1998
136	<i>Lycopodiella caroliniana</i>	(L.) Pichi Sermolli	Weakley et al. 1998
137	<i>Lycopus rubellus</i>	Moench.	Weakley et al. 1998
138	<i>Lyonia ligustrina</i>	(L.) DC.	Weakley et al. 1998
139	<i>Lyonia lucida</i>	(Lam.) K. Koch	Weakley et al. 1998
140	<i>Magnolia grandiflora</i>	L.	Weakley et al. 1998
141	<i>Magnolia virginiana</i>	L.	Weakley et al. 1998
142	<i>Mecardonia acuminata</i>	(Walt.) Small	Weakley et al. 1998
143	<i>Mitchella repens</i>	L.	Weakley et al. 1998
144	<i>Mitreola petiolata</i>	(J.F. Gmel.) Torr. & A. Gray	D.C. McLaughlin 481
145	<i>Mnesithia rugosum</i>	(Michx.) Koning & Sosef	D.C. McLaughlin 480, 482
146	<i>Myrica cerifera</i>	L.	D.A. Kruse 636, Weakley et al. 1998
147	<i>Myrica heterophylla</i>	Raf.	D.A. Kruse 417, Weakley et al. 1998
148	<i>Nelumbo lutea</i>	(Willd.) Pers.	Weakley et al. 1998
149	<i>Nymphoides aquatica</i>	(Gmel.) O. Ktz.	D.C. McLaughlin 490, D.A. Kruse 634
150	<i>Nyssa aquatica</i>	L.	Weakley et al. 1998
151	<i>Nyssa biflora</i>	Walt.	Weakley et al. 1998
152	<i>Nyssa ogeche</i>	L.	Weakley et al. 1998
153	<i>Osmunda cinnamomea</i>	L.	Weakley et al. 1998

154	<i>Oxypolis filiformis</i>	(Walt.) Britt.	Weakley et al. 1998
155	<i>Panicum gymnocarpon</i>	Ell.	Weakley et al. 1998
156	<i>Panicum hemitomom</i>	Schult.	Weakley et al. 1998
157	<i>Panicum hians</i>	Ell.	D.A. Kruse 825
158	<i>Panicum tenerum</i>	Beyr. ex Trin.	Weakley et al. 1998
159	<i>Panicum verrucosum</i>	Muhl.	Weakley et al. 1998
160	<i>Panicum virgatum</i>	L.	Weakley et al. 1998
161	<i>Pedicularis canadensis</i>	L.	Weakley et al. 1998
162	<i>Persea palustris</i>	(Raf.) Sarg.	Weakley et al. 1998
163	<i>Physostegia purpurea</i>	Engelm.	D.C. McLaughlin 431
164	<i>Pinus elliottii</i> var. <i>elliottii</i>	Engelm.	Weakley et al. 1998
165	<i>Pinus glabra</i>	Mill.	Weakley et al. 1998
166	<i>Pinus glabra</i>	Walt.	Weakley et al. 1998
167	<i>Pinus palustris</i>	L.	Weakley et al. 1998
168	<i>Pinus taeda</i>	L.	Weakley et al. 1998
169	<i>Planera aquatica</i>	(Walt.) J.F. Gmel.	Weakley et al. 1998
170	<i>Pluchea camphorata</i>	(L.) DC.	Weakley et al. 1998
171	<i>Polygonum hydropiperoides</i>	Michx.	D.C. McLaughlin 405, 410, Weakley et al. 1998
172	<i>Pontederia cordata</i>	L.	Weakley et al. 1998
173	<i>Populus heterophylla</i>	L.	Weakley et al. 1998
174	<i>Potamogeton</i> spp.	L.	Weakley et al. 1998
175	<i>Proserpinaca palustris</i>	L.	D.C. McLaughlin 411, Weakley et al. 1998
176	<i>Proserpinaca pectinata</i>	Lam.	Weakley et al. 1998
177	<i>Quercus alba</i>	L.	Weakley et al. 1998
178	<i>Quercus laurifolia</i>	Michx.	Weakley et al. 1998
179	<i>Quercus lyrata</i>	Walt.	Weakley et al. 1998

180	<i>Quercus michauxii</i>	Nutt.	Weakley et al. 1998
181	<i>Quercus nigra</i>	L.	Weakley et al. 1998
182	<i>Quercus pagoda</i>	Raf.	Weakley et al. 1998
183	<i>Quercus palustris</i>	Muench.	Weakley et al. 1998
184	<i>Quercus phellos</i>	L.	Weakley et al. 1998
185	<i>Quercus similis</i>	Ashe	Weakley et al. 1998
186	<i>Rhexia mariana</i>	L.	Weakley et al. 1998
187	<i>Rhexia virginica</i>	L.	Weakley et al. 1998
188	<i>Rhododendron canescens</i>	(Michx.) Sweet	Weakley et al. 1998
189	<i>Rhododendron viscosum</i>	(L.) Torr.	Weakley et al. 1998
190	<i>Rhynchospora caduca</i>	Ell.	D.A. Kruse 502, 809, Weakley et al. 1998
191	<i>Rhynchospora cephalantha</i>	Gray	Weakley et al. 1998
192	<i>Rhynchospora colorata</i>	(L.) Pfeifer	D.A. Kruse 817
193	<i>Rhynchospora corniculata</i>	(Lam.) Gray.	D.C. McLaughlin, 404, 412, 437, Weakley et al. 1998
194	<i>Rhynchospora elliotii</i>	A. Dietr.	D.C. McLaughlin 417, Weakley et al. 1998
195	<i>Rhynchospora fascicularis</i> var. <i>fascicularis</i>	(Michx.) Vahl	Weakley et al. 1998
196	<i>Rhynchospora globularis</i>	(Chapm.) Small	D.A. Kruse 816
197	<i>Rhynchospora glomerata</i>	(L.) Vahl.	D.C. McLaughlin 487, 497
198	<i>Rhynchospora inundata</i>	(Oakes) Fern.	Weakley et al. 1998
199	<i>Rhynchospora macrostachya</i>	Torr. Ex Gray	Weakley et al. 1998
200	<i>Rhynchospora microcarpa</i>	Gray	Weakley et al. 1998
201	<i>Rhynchospora perplexa</i>	Britt. Ex Small.	D.A. Kruse 490
202	<i>Rhynchospora rariflora</i>	(Michx.) Ell.	D.A. Kruse 819
203	<i>Rosa palustris</i>	Marsh.	Weakley et al. 1998
204	<i>Rudbeckia nitida</i> var. <i>texana</i>	Perdue	Weakley et al. 1998

205	<i>Sabal minor</i>	(Jacq.) Pers.	D.C. McLaughlin, Weakley et al. 1998
206	<i>Sabatia bartramii</i>	Wilbur	Weakley et al. 1998
207	<i>Sabatia campanulata</i>	(L.) Torr.	Weakley et al. 1998
208	<i>Saccharum baldwinii</i>	Spreng.	Weakley et al. 1998
209	<i>Sagittaria graminea</i>	Michx.	Weakley et al. 1998
210	<i>Sagittaria lancifolia</i>	L.	D.C. McLaughlin 439, D.A. Kruse 632
211	<i>Salix nigra</i>	Marsh.	Weakley et al. 1998
212	<i>Sambucus canadensis</i>	L.	Weakley et al. 1998
213	<i>Saururus cernuus</i>	L.	D.C. McLaughlin 409, D.A. Kruse 807, Weakley
214	<i>Scirpus cyperinus</i>	(L.) Kunth	Weakley et al. 1998
215	<i>Scleria baldwinii</i>	(Torr.) Steud.	Weakley et al. 1998
216	<i>Scleria georgiana</i>	Core	Weakley et al. 1998
217	<i>Scleria triglomerata</i>	Michx.	D.A. Kruse 491
218	<i>Scutellaria integrifolia</i>	L.	Weakley et al. 1998
219	<i>Sebastiania fruticosa</i>	(Batr.) Fern.	Weakley et al. 1998
220	<i>Smilax laurifolia</i>	L.	Weakley et al. 1998
221	<i>Smilax rotundifolia</i>	L.	Weakley et al. 1998
222	<i>Smilax walteri</i>	Pursh	Weakley et al. 1998
223	<i>Sparganium americanum</i>	Nutt.	D.A. Kruse, D.C. McLaughlin
224	<i>Sphagnum lescurii</i>	Sull. in Gray	Weakley et al. 1998
225	<i>Sphagnum macrophyllum</i>	Brid.	Weakley et al. 1998
226	<i>Spiranthes laciniata</i>	(Small) Ames	Weakley et al. 1998
227	<i>Stylisma aquatica</i>	(Walt.) Chapm.	Weakley et al. 1998
228	<i>Styrax americana</i>	Lam.	D.C. McLaughlin 509, Weakley et al. 1998
229	<i>Taxodium ascendens</i>	Brongn.	Weakley et al. 1998
230	<i>Taxodium distichum</i>	(L.) L. Rich.	Weakley et al. 1998



## APPENDIX C

SPECIES ALLIANCES AND ASSOCIATIONS FROM THE INTERNATIONAL  
CLASSIFICATION OF ECOLOGICAL COMMUNITIES

ALLIANCE	ASSOCIATION	CAREX GLAUCECENS	CAREX JOORII	CAREX VERRUCOSA
<i>Fagus grandifolia</i> Temporarily Flooded Forest Alliance		X	X	
	<i>Fagus grandifolia</i> - ( <i>Magnolia grandiflora</i> ) / <i>Hamamelis virginiana</i> / <i>Arundinaria gigantea</i> Forest G3?	X	X	
<i>Quercus (phellos, nigra, laurifolia)</i> Temporarily Flooded Forest Alliance		X	X	
	<i>Quercus phellos</i> - <i>Quercus nigra</i> - <i>Quercus alba</i> / <i>Chasmanthium sessiliflorum</i> Forest G3, G4	X	X	
<i>Nyssa (aquatica, biflora, ogeche)</i> Pond Seasonally Flooded Forest Alliance		X		X
	<i>Nyssa biflora</i> / <i>Ilex myrtifolia</i> / <i>Carex glaucescens</i> - <i>Eriocaulon compressum</i> Forest G2, G3	X		
	<i>Nyssa ogeche</i> / <i>Ilex myrtifolia</i> / <i>Carex turgescens</i> - <i>Carex striata</i> Forest G2?	X		X
<i>Taxodium ascendens</i> Seasonally Flooded Forest Alliance		X		
	<i>Taxodium ascendens</i> / <i>Ilex myrtifolia</i> Depression Forest G3?	X		
<i>Nyssa aquatica</i> - ( <i>Taxodium distichum</i> ) Semipermanently Flooded Forest Alliance		X		

<b>Taxodium distichum</b> <b>Semipermanently Flooded</b> <b>Forest Alliance</b>				
	<i>Taxodium distichum</i> / <i>Lemna minor</i> Forest G5	X		
<i>Magnolia virginiana</i> - <i>Nyssa biflora</i> - ( <i>Quercus laurifolia</i> ) <b>Saturated Forest Alliance</b>				
	<i>Quercus laurifolia</i> - <i>Magnolia virginiana</i> - <i>Nyssa biflora</i> / <i>Chasmanthium ornithorhynchum</i> Forest G2?	X		
<i>Pinus elliotii</i> - <i>Magnolia virginiana</i> - <i>Nyssa biflora</i> - ( <i>Taxodium ascendens</i> ) <b>Saturated Forest Alliance</b>		X		
	<i>Pinus elliotii</i> var. <i>elliotii</i> - <i>Magnolia virginiana</i> - <i>Taxodium ascendens</i> - <i>Nyssa biflora</i> / <i>Polygala cymosa</i> - <i>Carex glaucescens</i> Forest G2, G3	X		
<i>Pinus elliotii</i> - <i>Taxodium ascendens</i> <b>Saturated Woodland Alliance</b>		X		
	<i>Pinus elliotii</i> var. <i>elliotii</i> - <i>Taxodium ascendens</i> / <i>Polygala cymosa</i> - <i>Rhynchospora</i> spp. Woodland G2?	X		
<i>Vaccinium formosum</i> - <i>Vaccinium fuscum</i> <b>Seasonally Flooded Shrubland Alliance</b>		X		
	<i>Leucothoe racemosa</i> - <i>Vaccinium fuscum</i> - <i>Smilax walteri</i> Shrubland G1?	X		

<i>Aristida palustris</i> - <i>Andropogon</i> ( <i>capillipes</i> , <i>glaucopsis</i> ) - <i>Rhynchospora</i> spp. Seasonally Flooded Herbaceous Alliance		X		X
	<i>Aristida palustris</i> - <i>Panicum</i> <i>virgatum</i> - <i>Eriocaulon</i> <i>compressum</i> - <i>Eleocharis</i> <i>equisetoides</i> Herbaceous Vegetation G2, G3	X		X
<i>Panicum hemitomon</i> Seasonally Flooded Temperate Herbaceous Alliance	<i>Panicum hemitomon</i> - <i>Gratiola</i> <i>brevifolia</i> Herbaceous Vegetation G3?	X		
<i>Panicum verrucosum</i> - <i>Panicum rigidulum</i> - <i>Saccharum baldwinii</i> Seasonally Flooded Herbaceous Alliance		X		

## APPENDIX D

REPRESENTATIVE ANNOTATED HERBARIUM SPECIMENS USED TO CREATE  
*CAREX GLAUCESCENS* DISTRIBUTION MAPS

HERBARIUM	ACCESION NUMBER	COLLECTOR	COLLECTION NUMBER	COLLECTION YEAR	JULIAN COLLECTION DATE	STATE	COUNTY/ PARISH
SWSL	—	<u>R. Kral</u>	<u>56991</u>	1975	223	AL	AUTAUGA
UNA	848	<u>R.R. Haynes</u>	<u>7524</u>	1979	219	AL	BALDWIN
SWSL	—	<u>R. Kral</u>	<u>47903</u>	1972	210	AL	BIBB
UNA	977	<u>J.H.</u> <u>Wiersema</u>	<u>1299</u>	1979	258	AL	CHILTON
UNA	863	<u>R.R. Haynes</u>	<u>7456</u>	1979	191	AL	CHOCTAW
SWSL	—	<u>R. Kral</u>	<u>41129</u>	1970	248	AL	CLARKE
SWSL	—	<u>R. Kral</u>	<u>51266</u>	1973	225	AL	COFFEE
UNA	878	<u>J.S. Williams</u>	<u>51</u>	1979	272	AL	COVINGTON
UNA	987	<u>J.H.</u> <u>Wiersema</u>	<u>2056</u>	1980	242	AL	ELMORE
UNA	23507	<u>R.R. Haynes</u>	<u>9457</u>	1994	227	AL	ESCAMBIA
SWSL	—	<u>R. Kral</u>	<u>79092</u>	1991	175	AL	GENEVA
UNA	992	<u>J.H.</u> <u>Wiersema</u>	<u>2061</u>	1980	254	AL	HALE
SWSL	—	<u>J.R.</u> <u>MacDonald</u>	<u>2864</u>	1993	201	AL	HOUSTON
SWSL	—	<u>G.P. Wills</u>	<u>—</u>	1960	201	AL	LEE
US	817420	<u>C. Mohr</u>	<u>—</u>	1878	214	AL	MOBILE
UNA	858	<u>R.R. Haynes</u>	<u>8102</u>	1980	217	AL	WASHINGTON
SWSL	—	<u>C.T. Bryson</u>	<u>2668</u>	1979	266	FL	ALACHUA
FLAS	44926	<u>L.E. Arnold</u>	<u>—</u>	1946	193	FL	BAKER
FLAS	107335	<u>J. Beckner</u>	<u>1521</u>	1966	181	FL	BAY
FLAS	131028	<u>L. Conde</u>	<u>0</u>	1977	187	FL	BRADFORD
FSU	93219	<u>S. McDaniel</u>	<u>4798</u>	1964	186	FL	CALHOUN
LL	—	<u>E.L. Bridges</u>	<u>13860</u>	1990	157	FL	CLAY
US	364797	<u>Combs</u>	<u>131</u>	1898	206	FL	COLUMBIA

FLAS	30892	—	—	1937	231	FL	DIXIE
FLAS	99866	<u>D.B. Creager</u>	<u>606</u>	1967	162	FL	DUVAL
NCU	6545	<u>P.L. Redfearn</u>	<u>2763</u>	1956	202	FL	ESCAMBIA
FSU	172153	<u>L.C. Anderson</u>	<u>8286</u>	1985	165	FL	FRANKLIN
FSU	174587	<u>L.C. Anderson</u>	<u>9422</u>	1986	135	FL	GULF
NCU	6547	<u>J.R. Bozeman</u>	<u>10134</u>	1967	170	FL	HAMILTON
NCSC	55497	<u>R. Kral</u>	<u>2860</u>	1956	201	FL	HOLMES
FLAS	186309	<u>S. McDaniel</u>	<u>6561</u>	1965	204	FL	JACKSON
NCSC	62147	<u>R.K. Godfrey</u>	<u>54893</u>	1956	175	FL	JEFFERSON
FLAS	73749	<u>R. Kral</u>	<u>7659</u>	1958	217	FL	LAKE
LL	—	<u>E.L. Bridges</u>	<u>14053</u>	1990	182	FL	LIBERTY
DUKE	137095	<u>R.K. Godfrey</u>	<u>54010</u>	1955	275	FL	MADISON
DUKE	365995	<u>R.L. Wilbur</u>	<u>68319</u>	1997	206	FL	NASSAU
FSU	186783	<u>L.C. Anderson</u>	<u>15057</u>	1994	189	FL	OKALOOSA
DUKE	26039	<u>M.L. Singeltary</u>	<u>35</u>	1938	81	FL	OSCEOLA
NCU	6543	<u>R. Brinker</u>	<u>442</u>	1941	218	FL	PENSACOLA
SWSL	—	<u>S. McDaniel</u>	<u>6605</u>	1965	205	FL	SANTA ROSA
FLAS	70015	<u>A.P. Garber</u>	—	1876	196	FL	ST. AUGUSTINE
FLAS	38655	<u>L.E. Arnold</u>	—	1941	287	FL	ST. JOHN'S
MICH	—	<u>A.A. Reznicek</u>	<u>7604</u>	1985	227	FL	WAKULLA
FLAS	195974	<u>B. Herring</u>	<u>1431</u>	1998	211	FL	WALTON
FSU	97737	<u>S. McDaniel</u>	<u>6509</u>	1965	195	FL	WASHINGTON
NCU	6558	<u>J.R. Bozeman</u>	<u>9330</u>	1967	160	GA	APPLING
NCU	6559	<u>P.R. Cowart</u>	<u>8</u>	1971	207	GA	BALDWIN

DUKE	260087	<u>W.H. Duncan</u>	<u>12876</u>	1951	222	GA	BARTOW
FLAS	6437	<u>P.C. Lemon</u>	<u>—</u>	1943	235	GA	BERRIEN
MICH	—	<u>R.L. Lane, Jr.</u>	<u>1305</u>	1967	252	GA	BIBB
LL	—	<u>G.W. Ramsey</u>	<u>169</u>	1969	180	GA	BROOKS
NCU	6562	<u>D. Hall</u>	<u>206</u>	1964	282	GA	BULLOCH
NCU	6598	<u>J.E. Benedict</u>	<u>3328</u>	1936	185	GA	CANDLER
MICH	—	<u>R. Carter</u>	<u>8860</u>	1991	190	GA	CHARLTON
FLAS	134106	<u>W.H. Duncan</u>	<u>21255</u>	1958	193	GA	CHATHAM
NLU	407330	<u>V.E. McNeilus</u>	<u>98793</u>	1998	230	GA	CLINCH
NCU	6568	<u>W.R. Faircloth</u>	<u>2999</u>	1965	264	GA	COOK
NCU	6567	<u>J.R. Bozeman</u>	<u>5366</u>	1966	187	GA	DODGE
FLAS	146155	<u>R.L. Lane, Jr.</u>	<u>879</u>	1967	175	GA	DOOLY
MICH	—	<u>E.L. Bridges</u>	<u>17949</u>	1991	232	GA	EARLY
TAES	—	<u>D.J. Rosen</u>	<u>2273</u>	2002	235	GA	ECHOLS
US	3155031	<u>D. Eyles</u>	<u>6099</u>	1939	172	GA	EFFINGHAM
NCU	6569	<u>J.R. Bozeman</u>	<u>6392</u>	1966	202	GA	GLYNN
MICH	—	<u>A. Cronquist</u>	<u>5451</u>	1948	188	GA	GRADY
NLU	416143	<u>Crook</u>	<u>466</u>	1998	197	GA	HARRIS
SWSL	—	<u>V.E. McNeilus</u>	<u>88400</u>	1988	228	GA	LANIER
NLU	384059	<u>V.E. McNeilus</u>	<u>90657</u>	1990	201	GA	LEE
SWSL	—	<u>S.B. Jones</u>	<u>24622</u>	1984	260	GA	LIBERTY
NCU	6571	<u>J.R. Bozeman</u>	<u>10286</u>	1967	174	GA	LONG
US	3283545	<u>M.T. Strong</u>	<u>1041</u>	1993	222	GA	LOWNDES
NLU	281370	<u>R.A. Norris</u>	<u>5427</u>	1987	234	GA	MACON



MICH	—	<u>S.D. Jones</u>	<u>10347</u>	1993	300	LA	TANGIPAHOA
DUKE	205124	<u>J.W. Thieret</u>	<u>27768</u>	1967	247	LA	VERNON
NLU	188708	<u>R.D. Thomas</u>	<u>77310</u>	1981	178	LA	WASHINGTON
NLU	191902	<u>R.D. Thomas</u>	<u>78408</u>	1981	256	LA	WINN
US	3262833	<u>W.D. Longbottom</u>	<u>3317</u>	1992	230	MD	WICOMICO
DUKE	365157	<u>M.H. Alford</u>	<u>1779</u>	1998	285	MS	AMITE
SWSL	—	<u>C.T. Bryson</u>	<u>11855</u>	1992	216	MS	FORREST
NCU	6586	<u>S.B. Jones</u>	<u>2158</u>	1964	199	MS	GEORGE
MICH	—	<u>C.T. Bryson</u>	<u>14943</u>	1995	206	MS	GREENE
SWSL	—	<u>C.T. Bryson</u>	<u>10839</u>	1991	238	MS	HANCOCK
NCU	6588	<u>S.M. Tracy</u>	<u>5968</u>	1899	210	MS	HARRISON
MICH	—	<u>S.R. Hill</u>	<u>22411</u>	1991	194	MS	JACKSON
SWSL	—	<u>C.T. Bryson</u>	<u>14220</u>	1994	227	MS	LAMAR
SWSL	—	<u>C.T. Bryson</u>	<u>10148</u>	1990	224	MS	LOWNDES
NLU	376564	<u>C.T. Bryson</u>	<u>14041</u>	1994	210	MS	MARION
SWSL	—	<u>S. McDaniel</u>	<u>24223</u>	1980	214	MS	PEARL RIVER
SWSL	—	<u>C.T. Bryson</u>	<u>14294</u>	1994	250	MS	PERRY
SWSL	—	<u>C.T. Bryson</u>	<u>14469</u>	1994	271	MS	PIKE
SWSL	—	<u>C.T. Bryson</u>	<u>6919</u>	1987	228	MS	SCOTT
SWSL	—	<u>C.T. Bryson</u>	<u>11006</u>	1991	259	MS	STONE
SWSL	—	<u>S. McDaniel</u>	<u>30900</u>	1990	250	MS	WAYNE
NCSC	63411	<u>A.E. Blair</u>	<u>1275</u>	1966	282	NC	BEAUFORT
DUKE	128731	<u>W.B. Fox</u>	<u>4092</u>	1950	214	NC	BERTIE
NCU	6459	<u>A.E. Radford</u>	<u>3013</u>	1947	221	NC	BLADEN

NCSC	748	<u>R.K. Godfrey</u>	<u>4858</u>	1938	188	NC	BRUNSWICK
DUKE	9899	<u>H.L. Blomquist</u>	<u>10351</u>	1938	191	NC	CARTERET
NCU	6462	<u>H.E. Ahles</u>	<u>47988</u>	1958	211	NC	CHOWAN
DUKE	344948	<u>R.L. Wilbur</u>	<u>49995</u>	1988	229	NC	COLUMBUS
DUKE	343698	<u>R.L. Wilbur</u>	<u>56191</u>	1990	206	NC	CRAVEN
NCSC	42757	<u>W.B. Fox</u>	<u>5623</u>	1951	290	NC	CUMBERLAND
DUKE	128738	<u>W.B. Fox</u>	<u>4166</u>	1950	216	NC	CURRITUCK
NCU	6467	<u>P.J. Crutchfield</u>	<u>4614</u>	1959	238	NC	DARE
NCU	6471	<u>H.E. Ahles</u>	<u>33086</u>	1957	214	NC	DUPLIN
NCU	6472	<u>A.E. Radford</u>	<u>36700</u>	1958	195	NC	EDGECOMBE
NCU	6473	<u>H.E. Ahles</u>	<u>54546</u>	1958	289	NC	GATES
NCU	6475	<u>A.E. Radford</u>	<u>36529</u>	1958	192	NC	GREEN
MICH	—	<u>A.E. Radford</u>	<u>36377</u>	1958	192	NC	GREENE
NCU	6476	<u>H.E. Ahles</u>	<u>17097</u>	1956	200	NC	HALIFAX
FLAS	28440	<u>R.K. Godfrey</u>	<u>—</u>	1938	217	NC	HARNETT
NCSC	20986	<u>W.B. Fox &amp; R.K. Godfrey</u>	<u>2802</u>	1949	204	NC	HERTFORD
NCU	6479	<u>H.E. Ahles</u>	<u>29617</u>	1957	177	NC	HOKE
NCU	6481	<u>A.E. Radford</u>	<u>39070</u>	1958	218	NC	HYDE
DUKE	159876	<u>H.L. Blomquist</u>	<u>16883</u>	1955	301	NC	JOHNSTON
DUKE	343696	<u>R.L. Wilbur</u>	<u>56171</u>	1990	200	NC	JONES
DUKE	343700	<u>R.L. Wilbur</u>	<u>54097</u>	1989	215	NC	LENOIR
DUKE	128727	<u>W.B. Fox</u>	<u>4088</u>	1950	214	NC	MARTIN
NCU	6488	<u>A.E. Radford</u>	<u>19555</u>	1956	282	NC	MONTGOMERY
US	1767882	<u>R.K. Godfrey</u>	<u>5111</u>	1938	195	NC	MOORE

DUKE	34203	<u>R.K. Godfrey</u>	<u>5124</u>	1938	199	NC	NASH
NCU	6490	<u>D.J. Sieren</u>	<u>1672</u>	1978	201	NC	NEW HANOVER
US	1768186	<u>R.K. Godfrey</u>	<u>5819</u>	1938	218	NC	ONslow
NCSC	14653	<u>R.K. Godfrey</u>	<u>48336</u>	1948	202	NC	PAMLICO
NCSC	844	<u>J.C. Rabb</u>	<u>—</u>	1940	224	NC	PASQUOTANK
NCU	6495	<u>H.E. Ahles</u>	<u>32339</u>	1957	207	NC	PENDER
DUKE	128697	<u>W.B. Fox</u>	<u>4168</u>	1950	217	NC	PERQUIMANS
DUKE	53516	<u>H.L. Blomquist</u>	<u>6456</u>	1931	208	NC	PITT
NCU	6497	<u>A.E. Radford</u>	<u>14397</u>	1956	205	NC	RICHMOND
NCU	6498	<u>R. Kral</u>	<u>3417</u>	1956	248	NC	ROBESON
DUKE	37023	<u>R.K. Godfrey</u>	<u>5713</u>	1938	217	NC	SAMPSON
DUKE	361680	<u>J.W. Horn</u>	<u>1366</u>	1998	232	NC	SCOTLAND
DUKE	128696	<u>W.B. Fox</u>	<u>4107</u>	1950	215	NC	TYRELL
DUKE	141167	<u>W.B. Fox</u>	<u>1832</u>	1948	230	NC	WAKE
US	2265533	<u>W.B. Fox</u>	<u>4105</u>	1950	215	NC	WASHINGTON
NCU	6505	<u>W.W. Ashe</u>	<u>—</u>	1898	319	NC	WASHINGTON
NCU	6506	<u>A.E. Radford</u>	<u>37990</u>	1958	209	NC	WILSON
NCU	6511	<u>W.T. Batson</u>	<u>—</u>	1951	267	SC	AIKEN
NCU	6512	<u>C.R. Bell</u>	<u>3989</u>	1956	181	SC	ALLENDALe
NCU	6513	<u>H.E. Ahles</u>	<u>30469</u>	1957	185	SC	BAMBERG
MICH	—	<u>P.E. Hyatt</u>	<u>5753</u>	1993	259	SC	BARNWELL
DUKE	280077	<u>J.M. MacDougal</u>	<u>1497</u>	1981	217	SC	BERKELEY
NLU	101065	<u>T. Daggy</u>	<u>5147</u>	1968	246	SC	CHESTERFIELD
NCU	6520	<u>C.R. Bell</u>	<u>3696</u>	1956	177	SC	COLLETON

MICH	—	<u>J.B. Nelson</u>	<u>9608</u>	1990	228	SC	DARLINGTON
NCU	6524	<u>H.E. Ahles</u>	<u>32178</u>	1957	206	SC	DILLON
NCU	6525	<u>H.E. Ahles</u>	<u>31822</u>	1957	201	SC	DORCHESTER
NCU	6527	<u>A.E. Radford</u>	<u>44262</u>	1961	236	SC	FAIRFIELD
NCU	6528	<u>C.R. Bell</u>	<u>10895</u>	1957	291	SC	FLORENCE
NCU	6530	<u>H.E. Ahles</u>	<u>15725</u>	1956	180	SC	HAMPTON
NCU	6531	<u>H.E. Ahles</u>	<u>15517</u>	1956	177	SC	JASPER
UNA	9067	<u>C.N. Horn</u>	<u>7630</u>	1993	232	SC	KERSHAW
US	585326	<u>H.D. House</u>	<u>2613</u>	1906	207	SC	LAUCASLER
NCU	6532	<u>A.E. Radford</u>	<u>27323</u>	1957	207	SC	LEE
NCU	6533	<u>A.E. Radford</u>	<u>27211</u>	1957	190	SC	LEXINGTON
NCU	6534	<u>C.R. Bell</u>	<u>—</u>	1958	183	SC	MARION
NCU	6535	<u>A.E. Radford</u>	<u>15601</u>	1956	222	SC	MARLBORO
NCU	6536	<u>H.E. Ahles</u>	<u>31682</u>	1957	200	SC	ORANGEBURG
MICH	—	<u>J.B. Nelson</u>	<u>11628</u>	1991	274	SC	RICHLAND
NCU	6538	<u>A.E. Radford</u>	<u>26827</u>	1957	189	SC	SALUDA
NCU	6540	<u>A.E. Radford</u>	<u>27415</u>	1957	208	SC	SUMTER
US	1837291	<u>R.K. Godfrey</u>	<u>398</u>	1939	191	SC	WILLIAMSBURG
SWSL	—	<u>V.E. McNeilus</u>	<u>90907</u>	1990	251	TN	BLEDSON
US	2132993	<u>W.B. Fox</u>	<u>5623</u>	1951	280	TN	CUMBERLAND
US	1814699	<u>H.K. Svenson</u>	<u>9183</u>	1938	227	TN	GRUNDY
NLU	384064	<u>J.A. Churchill</u>	<u>95121</u>	1995	237	TN	VAN BUREN
LL	292965	<u>E.L. Bridges</u>	<u>4500</u>	1986	205	TX	ANGELINA
LL	—	<u>P. Fryxell</u>	<u>3016</u>	1978	204	TX	HARDIN

LL	—	<u>E.L. Bridges</u>	<u>5674</u>	1987	224	TX	JASPER
US	2297824	<u>C.L. Lundell</u>	<u>14092</u>	1945	199	TX	NEWTON
LL	292967	<u>B.C. Tharp</u>	<u>43282</u>	1941	102	TX	POLK
LL	292966	<u>E.L. Bridges</u>	<u>5659</u>	1987	224	TX	SABINE
TAES	158793	<u>F.W. Gould</u>	<u>6523</u>	1952	262	TX	SAN AUGUSTINE
SWSL	—	<u>S.D. Jones</u>	<u>3707</u>	1989	229	TX	TYLER
NLU	383621	<u>W.J. Hayden</u>	<u>2880</u>	1990	252	VA	CHESTERFIELD
NCU	6591	<u>A. Carter</u>	<u>15</u>	1974	263	VA	CUMBERLAND
NCU	6592	<u>A.M. Harvill</u>	<u>17613</u>	1967	215	VA	HENRICO
SWSL	—	<u>L.L. Whitmarsh</u>	<u>469</u>	1979	211	VA	ISLE OF WIGHT
NCU	6593	<u>R. Kral</u>	<u>9510</u>	1959	247	VA	NANSEMOND
US	356208	<u>H. Kearney</u>	<u>1516</u>	1898	190	VA	NORFOLK
NCU	6589	<u>E.B. Neyes</u>	<u>—</u>	—	—	VA	PORTSMOLLH
FLAS	125141	<u>D.M.E. Ware</u>	<u>5600</u>	1974	210	VA	PRINCE GEORGE
MICH	—	<u>J.M. Fogg</u>	<u>9746</u>	1935	255	VA	PRINCESS ANNE
US	3380826	<u>M.T. Strong</u>	<u>2167</u>	1999	253	VA	SUFFOLK
SWSL	—	<u>B. Mikula</u>	<u>7631</u>	1950	226	VA	SURRY
NCU	6596	<u>R. Kral</u>	<u>11368</u>	1960	241	VA	SUSSEX
SWSL	—	<u>V.E. McNeilus</u>	<u>95577</u>	1995	238	VA	VAN BUREN

## APPENDIX E

REPRESENTATIVE ANNOTATED HERBARIUM SPECIMENS USED TO CREATE  
*CAREX JOORII* DISTRIBUTION MAPS

HERBARIUM	ACCESION NUMBER	COLLECTOR	COLLECTION NUMBER	COLLECTION YEAR	JULIAN COLLECTION DATE*	STATE	COUNTY/ PARISH
UNA	948	<u>D.L. Lentz</u>	<u>70</u>	1980	242	AL	AUTAUGA
MICH	—	<u>B. Keener</u>	<u>1464</u>	1998	226	AL	BLOUNT
UNA	914	<u>M. Birk</u>	<u>447</u>	1995	264	AL	DALLAS
SWSL	—	<u>R. Kral</u>	<u>56408</u>	1975	224	AL	FAYETTE
UNA	904	<u>M. Birk</u>	<u>115</u>	1994	230	AL	HALE
SWSL	—	<u>J.R. MacDonald</u>	<u>5083</u>	1992	203	AL	HOUSTON
NCU	6439	<u>D. Isely</u>	<u>4893</u>	1946	216	AL	LIMESTONE
SWSL	—	<u>C.T. Bryson</u>	<u>3099</u>	1980	220	AL	MADISON
UNA	964	<u>J.H. Wiersema</u>	<u>1961</u>	1980	185	AL	MARENGO
UNA	999	<u>R.R. Haynes</u>	<u>8194</u>	1980	218	AL	PERRY
SWSL	—	<u>S. McDaniel</u>	<u>9769</u>	1967	259	AL	PICKENS
SWSL	—	<u>R. Kral</u>	<u>47958</u>	1972	211	AL	PIKE
DUKE	42039	<u>D.S. Correll</u>	<u>8325</u>	1937	251	AL	SAINT CLAIR
US	3164922	<u>D.S. Correll</u>	<u>8325</u>	1937	251	AL	ST. CLAIR
UNA	944	<u>S. McDaniel</u>	<u>9917</u>	1967	280	AL	WASHINGTON
SWSL	—	<u>P.O. DeWitt</u>	<u>68691</u>	1974	183	AR	ARKANSAS
NLU	265332	<u>R.D. Thomas</u>	<u>97424</u>	1986	198	AR	ASHLEY
NLU	255652	<u>S. Leslie</u>	<u>1184</u>	1985	231	AR	BRADLEY
NLU	287878	<u>E. Sundell</u>	<u>7879</u>	1987	246	AR	CALHOUN
NLU	311881	<u>E. Sundell</u>	<u>9120</u>	1989	259	AR	CLEVELAND
MICH	—	<u>D. Demaree</u>	<u>13698</u>	1933	262	AR	DREW
UNA	969	<u>S. Alley</u>	<u>87720</u>	1977	216	AR	FAULKNER
NCU	6440	<u>G.E. Tucker</u>	<u>6544</u>	1967	252	AR	GRANT

NCU	6456	<u>G.E. Tucker</u>	<u>6552</u>	1967	252	AR	HOT SPRING
MICH	—	<u>S.L. Orzell</u>	<u>3586</u>	1985	296	AR	JACKSON
SFRP	—	<u>P.E. Hyatt</u>	<u>7758</u>	1997	257	AR	LAFAYETTE
MICH	—	<u>S.L. Orzell</u>	<u>3600</u>	1985	297	AR	LAWERENCE
MICH	—	<u>S.L. Orzell</u>	<u>3270</u>	1985	280	AR	MILLER
MICH	—	<u>S.L. Orzell</u>	<u>3675</u>	1985	313	AR	MONROE
SFRP	—	<u>P.E. Hyatt</u>	<u>7765</u>	1997	257	AR	NEVADA
MICH	—	<u>H.E. Hasse</u>	<u>—</u>	1886	275	AR	PULASKI
NLU	92010	<u>D. Lawson</u>	<u>2071</u>	1974	204	AR	SEVIER
SWSL	—	<u>R.D. Thomas</u>	<u>111768</u>	1989	216	AR	UNION
MICH	—	<u>S.L. Orzell</u>	<u>3561</u>	1985	296	AR	WOODRUFF
FLAS	180878	<u>W.J. Dunn</u>	<u>803</u>	1979	184	FL	ALACHUA
FSU	147232	<u>S.W. Leonard</u>	<u>6367</u>	1976	191	FL	CALHOUN
FLAS	154283	<u>J.R. Burkhalter</u>	<u>9029</u>	1983	267	FL	ESCAMBIA
FSU	170546	<u>L.C. Anderson</u>	<u>7774</u>	1984	319	FL	FRANKLIN
FSU	16291	<u>R. Kral</u>	<u>1638</u>	1955	287	FL	GADSDEN
FSU	41867	<u>R.K. Godfrey</u>	<u>57587</u>	1958	246	FL	GULF
NCU	106675	<u>R.K. Godfrey</u>	<u>54257</u>	1955	294	FL	JACKSON
FSU	186078	<u>L.C. Anderson</u>	<u>14518</u>	1993	228	FL	LEON
MICH	—	<u>E.L. Bridges</u>	<u>14469</u>	1990	214	FL	LEVY
FSU	135523	<u>R.K. Godfrey</u>	<u>71631</u>	1972	204	FL	LIBERTY
FSU	147432	<u>R.K. Godfrey</u>	<u>75363</u>	1976	231	FL	MADISON
NCU	6442	<u>S.L. Orzell</u>	<u>12507</u>	1989	264	FL	OKALOOSA
FSU	170885	<u>J.R. Burkhalter</u>	<u>9724</u>	1984	290	FL	SANTA ROSA



FLAS	107048	<u>D.B. Ward</u>	<u>6687</u>	1968	197	FL	WALTON
FLAS	109541	<u>W.H. Duncan</u>	<u>12587</u>	1951	170	GA	BARTOW
MICH	—	<u>R. Carter</u>	<u>6093</u>	1987	225	GA	BEN HILL
SWSL	—	<u>R. Carter</u>	<u>4432</u>	1985	236	GA	BROOKS
TAES	207541	<u>R. Carter</u>	<u>10269</u>	1992	216	GA	BRYAN
MICH	—	<u>A.W. Cusick</u>	<u>6531</u>	1967	256	GA	BURKE
NCU	6563	<u>J.R. Bozeman</u>	<u>10501</u>	1967	177	GA	CAMDEN
SWSL	—	<u>V.E. McNeilus</u>	<u>—</u>	1986	216	GA	CLINCH
US	384434	<u>R.M. Harper</u>	<u>563</u>	1900	246	GA	DOOLY
US	1929636	<u>R.F. Thorne</u>	<u>6677</u>	1947	259	GA	EARLY
SWSL	—	<u>V.E. McNeilus</u>	<u>—</u>	1985	215	GA	ECHOLS
NCU	6601	<u>D.E. Boufford</u>	<u>21688</u>	1979	240	GA	EMANUEL
US	511667	<u>A.J. Pieters</u>	<u>504</u>	1899	227	GA	GLYNN
SWSL	—	<u>V.E. McNeilus</u>	<u>—</u>	1986	217	GA	LANIER
NCU	6602	<u>J.R. Bozeman</u>	<u>1699</u>	1962	279	GA	LONG
MICH	—	<u>J.R. Manhart</u>	<u>143</u>	1980	243	GA	MUSCOGEE
NCSC	103044	<u>N.C. Coile</u>	<u>2298</u>	1981	230	GA	WALKER
MICH	—	<u>R. Kral</u>	<u>85414</u>	1995	221	GA	WHEELER
NLU	331079	<u>C.M. Allen</u>	<u>11021</u>	1981	179	LA	ALLEN
TAES	188984	<u>S. &amp; G. Jones</u>	<u>3803</u>	1989	230	LA	BEAUREGARDE
NLU	—	<u>W.C. Holmes</u>	<u>3675</u>	1979	244	LA	BIENVILLE
DUKE	77974	<u>D.S. Correll</u>	<u>10115</u>	1938	219	LA	BOSSIER
UNA	949	<u>R.R. Haynes</u>	<u>5340</u>	1975	249	LA	CADDO
NLU	337445	<u>L.E. Brown</u>	<u>14942</u>	1990	287	LA	CALCASIEU

SWSL	—	<u>W.C. Holmes</u>	<u>5394</u>	1991	242	LA	CLAIBORNE
NLU	163377	<u>R.D. Thomas</u>	<u>67940</u>	1979	266	LA	DESOTO
NLU	159108	<u>K. Cormier</u>	<u>1156</u>	1978	274	LA	EVANGELINE
NLU	191966	<u>R.D. Thomas</u>	<u>77802</u>	1981	214	LA	FRANKLIN
NLU	73971	<u>C.E. DePoe</u>	<u>64522</u>	1966	228	LA	GRANT
NLU	90037	<u>R.D. Thomas</u>	<u>40213</u>	1974	203	LA	JACKSON
NLU	127439	<u>P. Laird</u>	<u>1488</u>	1975	215	LA	LASALLE
SFRP	5328	<u>R. Smith &amp; H. Grelen</u>	<u>1134</u>	1975	273	LA	LINCOLN
NLU	252549	<u>G.F. Joye</u>	<u>190</u>	1984	272	LA	LIVINGSTON
NLU	28717	<u>R.D. Thomas</u>	<u>16986</u>	1969	277	LA	MOREHOUSE
MICH	—	<u>P.E. Hyatt</u>	<u>6693</u>	1995	199	LA	NATCHITOCHES
NLU	377453	<u>C.M. Allen</u>	<u>18084</u>	1994	254	LA	OUACHITA
NLU	—	<u>R.D. Thomas</u>	<u>5449</u>	1967	276	LA	RAPIDES
NLU	347573	<u>A. Parker</u>	<u>=</u>	1992	232	LA	RED RIVER
NCU	6443	<u>D. Demaree</u>	<u>48375</u>	1963	190	LA	SABINE
FLAS	121564	<u>V.I. Sullivan</u>	<u>1590</u>	1973	227	LA	ST. HELENA
US	1032803	<u>G. Arsene</u>	<u>11500</u>	1919	—	LA	ST. TAMMANY
SWSL	—	<u>F.M. Givens</u>	<u>4413</u>	1985	319	LA	TANGIPAHOA
NLU	321925	<u>R.D. Thomas</u>	<u>119900</u>	1990	214	LA	UNION
NLU	73969	<u>C.E. DePoe</u>	<u>57673</u>	1966	201	LA	VERNON
NLU	225162	<u>D.D. Taylor</u>	<u>5834</u>	1983	261	LA	WASHINGTON
NLU	176803	<u>R.D. Thomas</u>	<u>72135</u>	1980	204	LA	WEBSTER
NLU	149782	<u>R.D. Thomas</u>	<u>61111</u>	1978	263	LA	WEST CARROLL
NLU	193946	<u>K.H. Kessler</u>	<u>1719</u>	1981	249	LA	WINN

MICH	—	<u>C.T. Frye</u>	<u>2080</u>	1999	221	MD	CAROLINE
MICH	—	<u>D. Samson</u>	<u>—</u>	1993	244	MD	DORCHESTER
US	1815716	<u>A. Chase</u>	<u>12631</u>	1940	286	MD	WORCESTER
US	671934	<u>B.F. Bush</u>	<u>6327</u>	1910	255	MO	DUNKLIN
DUKE	358747	<u>M.H. Alford</u>	<u>73</u>	1997	284	MS	AMITE
NCU	6603	<u>J.D. Ray</u>	<u>7134</u>	1956	206	MS	FORREST
SWSL	—	<u>W. Morris</u>	<u>2355</u>	1986	203	MS	GRENADA
SWSL	—	<u>C.T. Bryson</u>	<u>10841</u>	1991	238	MS	HANCOCK
MICH	—	<u>C.T. Bryson</u>	<u>6672</u>	1987	206	MS	ITAWAMBA
SWSL	—	<u>C.T. Bryson</u>	<u>12851</u>	1993	244	MS	JACKSON
NCU	6604	<u>J.G. Teer</u>	<u>—</u>	1963	185	MS	JONES
SWSL	—	<u>S. McDaniel</u>	<u>24552</u>	1980	268	MS	KEMPER
SWSL	—	<u>C.T. Bryson</u>	<u>14198</u>	1994	227	MS	LAMAR
SWSL	—	<u>C.T. Bryson</u>	<u>14966</u>	1995	236	MS	LEFLORE
SWSL	—	<u>C.T. Bryson</u>	<u>10145</u>	1990	224	MS	LOWNDES
US	1923025	<u>W.B. McDougall</u>	<u>1479</u>	1947	253	MS	MADISON
TAES	235371	<u>C.T. Bryson</u>	<u>10152</u>	1990	224	MS	MONROE
SWSL	—	<u>C.T. Bryson</u>	<u>305</u>	1973	231	MS	NEWTON
FLAS	132042	<u>S. McDaniel</u>	<u>12492</u>	1969	300	MS	NOXUBEE
NLU	269764	<u>C.T. Bryson</u>	<u>3440</u>	1982	261	MS	OKTIBBEHA
US	2068063	<u>R.L. Wilbur</u>	<u>3341</u>	1950	193	MS	SCOTT
SWSL	—	<u>C.T. Bryson</u>	<u>12553</u>	1993	215	MS	SHARKEY
TAES	5700	<u>S.M. Tracy</u>	<u>8475</u>	1903	213	MS	SIMPSON
FLAS	76314	<u>J.D. Ray</u>	<u>7490</u>	1956	220	MS	TISHOMINGO

SWSL	—	<u>S. McDaniel</u>	<u>9793</u>	1967	263	MS	WINSTON
NCU	6400	<u>G.S. Ramseur</u>	<u>2917</u>	1956	307	NC	ALAMANCE
NCU	6401	<u>H.E. Ahles</u>	<u>52121</u>	1958	301	NC	BERTIE
DUKE	311103	<u>R.L. Wilbur</u>	<u>38136</u>	1985	230	NC	CHATHAM
NCU	6404	<u>A.E. Radford</u>	<u>40252</u>	1958	253	NC	CRAVEN
DUKE	368481	<u>R.L. Wilbur</u>	<u>69421</u>	1997	270	NC	DURHAM
NCU	6408	<u>A.E. Radford</u>	<u>44476</u>	1961	281	NC	GATES
DUKE	19800	<u>W.T. Batson</u>	<u>554</u>	1951	292	NC	GRANVILLE
NCSC	13456	<u>S. Boyce</u>	<u>2119</u>	1948	297	NC	HERTFORD
NCU	6412	<u>L.S. Beard</u>	<u>983</u>	1955	200	NC	LEE
NCU	6413	<u>A.E. Radford</u>	<u>39431</u>	1958	221	NC	MARTIN
NCU	6414	<u>E.F. Wells</u>	<u>2357</u>	1969	292	NC	MONTGOMERY
UNA	929	<u>A.E. Radford</u>	<u>44342</u>	1961	252	NC	NASH
MICH	—	<u>K.L. McIntosh</u>	<u>12071</u>	1994	246	NC	PENDER
NCU	6421	<u>C.R. Bell</u>	<u>—</u>	1958	224	NC	PERSON
NCSC	14685	<u>W.B. Fox &amp; R.K. Godfrey</u>	<u>2150</u>	1948	302	NC	SAMPSON
NCSC	120066	<u>R. Ingle</u>	<u>735</u>	1992	223	NC	WAKE
NCSC	53197	<u>J.P. Gillespie</u>	<u>20</u>	1955	194	NC	WAYNE
NLU	146420	<u>J. Taylor</u>	<u>23279</u>	1976	241	OK	MCCURTAIN
NCU	6424	<u>W.T. Batson</u>	<u>—</u>	1951	206	SC	AIKEN
NCU	6426	<u>W.T. Batson</u>	<u>—</u>	1952	286	SC	BARNWELL
US	1837755	<u>R.K. Godfrey</u>	<u>1005</u>	1939	212	SC	CLARENDON
NCU	6428	<u>C.R. Bell</u>	<u>5389</u>	1956	285	SC	COLLETON
NCU	6429	<u>C.R. Bell</u>	<u>10809</u>	1957	291	SC	FLORENCE

DUKE	359454	<u>J.B. Nelson</u>	<u>18742</u>	1997	269	SC	GEORGETOWN
NCU	6431	<u>C.R. Bell</u>	<u>4982</u>	1956	252	SC	HAMPTON
NCU	6432	<u>J.A. Duke</u>	<u>94</u>	1957	292	SC	HORRY
NCU	6435	<u>H.E. Ahles</u>	<u>18188</u>	1956	251	SC	JASPER
NCU	6436	<u>C.R. Bell</u>	<u>11049</u>	1957	293	SC	MARION
NCU	6437	<u>A.E. Radford</u>	<u>27506</u>	1957	208	SC	SUMTER
MICH	—	<u>V.E. McNeilus</u>	<u>98901</u>	1998	257	TN	BLEDSON
UNA	994	<u>R. Kral</u>	<u>40685</u>	1970	229	TN	COFFEE
MICH	—	<u>V.E. McNeilus</u>	<u>95676</u>	1995	264	TN	COOK
MICH	—	<u>K.E. Rogers</u>	<u>40999</u>	1966	287	TN	CUMBERLAND
NLU	194323	<u>T.S. Patrick</u>	<u>2698</u>	1979	243	TN	FENTRESS
LL	—	<u>T. Smith</u>	<u>253</u>	1982	210	TN	FRANKLIN
NLU	408412	<u>V.E. McNeilus</u>	<u>98917</u>	1998	257	TN	HAMILTON
NCU	6446	<u>M. Pyne</u>	<u>93277</u>	1993	276	TN	LEWIS
US	3164923	<u>H.K. Svenson</u>	<u>9519</u>	1938	197	TN	MARION
MICH	—	<u>V.E. McNeilus</u>	<u>931724</u>	1993	223	TN	OVERTON
MICH	—	<u>H.K. Svenson</u>	<u>9658</u>	1938	229	TN	SEQUATOCHIE
US	3164924	<u>B.C. Tharp</u>	<u>43268</u>	1944	288	TX	ANGELINA
US	333292	<u>A.A. Heller</u>	<u>4106</u>	1898	227	TX	BOWIE
TAES	188838	<u>S. &amp; G. Jones</u>	<u>3859</u>	1989	294	TX	CASS
TAES	38325	<u>C.L. York</u>	<u>47132</u>	1939	201	TX	GREGG
DUKE	110468	<u>B.C. Tharp</u>	<u>43322</u>	1937	253	TX	HARDIN
LL	293021	<u>J.W. Kessler</u>	<u>5100</u>	1981	261	TX	HARRIS
NCU	6599	<u>R.J. Fleetwood</u>	<u>12231</u>	1976	203	TX	HARRISON

TAES	188692	<u>S. &amp; G. Jones</u>	<u>3827</u>	1989	267	TX	HOUSTON
TAES	77672	<u>F.W. Gould</u>	<u>7318</u>	1956	297	TX	Jasper
LL	293022	<u>B. Ferguson</u>	<u>770</u>	1907	300	TX	LIBERTY
LL	—	<u>D.S. Correll</u>	<u>26408</u>	1962	291	TX	MARION
LL	293020	<u>B. Ferguson</u>	<u>763</u>	1907	299	TX	MONTGOMERY
TAES	10667	<u>H.B. Parks</u>	<u>25929</u>	1937	283	TX	NACOGDOCHES
SWSL	—	<u>S.D. Jones</u>	<u>5669</u>	1990	209	TX	NEWTON
LL	—	<u>S.L. Orzell</u>	<u>12507</u>	1989	264	TX	OKALOOSA
NLU	391506	<u>L.E. Brown</u>	<u>—</u>	1991	0	TX	ORANGE
NCU	6457	<u>E.S. Nixon</u>	<u>5419</u>	1972	276	TX	SAN JACINTO
TAES	221907	<u>E.S. Nixon</u>	<u>16988</u>	1988	189	TX	TRINITY
SWSL	—	<u>T.F. Wieboldt</u>	<u>3182</u>	1977	253	VA	CAMPBELL
NCU	6448	<u>D.M.E. Ware</u>	<u>6485</u>	1975	261	VA	CHARLES CITY
MICH	—	<u>K.L. McIntosh</u>	<u>13021</u>	1996	285	VA	DINWIDDIE
FLAS	141379	<u>J.M. Greaves</u>	<u>1452</u>	1976	237	VA	GLOUCESTER
US	1682819	<u>M.L. Fernald</u>	<u>6551</u>	1936	231	VA	GREENSVILLE
MICH	—	<u>H.H. Ittis</u>	<u>30329</u>	1990	217	VA	ISLE OF WIGHT
MICH	—	<u>T.F. Wieboldt</u>	<u>8787</u>	1996	252	VA	MECKLENBURG
NCU	6451	<u>P.K. Appler</u>	<u>346</u>	1972	287	VA	NEWPORT NEWS
SWSL	—	<u>R. Kral</u>	<u>56511</u>	1975	248	VA	PRINCE GEORGE
SWSL	—	<u>B. Mikula</u>	<u>8045</u>	1950	241	VA	SOUTHAMPTON
US	1682820	<u>M.L. Fernald</u>	<u>6552</u>	1936	232	VA	SUSSEX

## APPENDIX F

REPRESENTATIVE ANNOTATED HERBARIUM SPECIMENS USED TO CREATE  
*CAREX VERRUCOSA* DISTRIBUTION MAPS

HERBARIUM	ACCESION NUMBER	COLLECTOR	COLLECTION NUMBER	COLLECTION YEAR	JULIAN COLLECTION DATE*	STATE	COUNTY/ PARISH
NLU	363603	<u>J.R. MacDonald</u>	<u>5570</u>	1992	262	AL	HOUSTON
NLU	104987	<u>J. Taylor</u>	<u>16480</u>	1974	186	AL	MOBILE
FLAS	94919	<u>I.L. Wiggins</u>	<u>19737</u>	1965	109	FL	ALACHUA
FLAS	38653	<u>W.A. Murrill</u>	<u>=</u>	1941	144	FL	BAKER
FLAS	194985	<u>E.L. Bridges</u>	<u>21274</u>	1993	79	FL	CLAY
FLAS	77783	<u>L.J. Brass</u>	<u>32731</u>	1960	286	FL	COLLIER
FLAS	47130	<u>A.H. Curtiss</u>	<u>3249</u>	—	196	FL	DUVAL
FLAS	32392	<u>L.E. Arnold</u>	<u>=</u>	1940	108	FL	FLAGLER
FLAS	92367	<u>S. McDaniel</u>	<u>6130</u>	1965	124	FL	FRANKLIN
FLAS	102332	<u>S. McDaniel</u>	<u>8576</u>	1967	77	FL	GADSDEN
FSU	39853	<u>R. Kral</u>	<u>6413</u>	1958	110	FL	HAMILTON
MICH	—	<u>W.P. Stoutamire</u>	<u>1742</u>	1955	320	FL	HIGHLANDS
FLAS	85759	<u>J.A. Lassiter</u>	<u>25076</u>	1962	140	FL	HILLSBOROUGH
DUKE	165719	<u>R. Kral</u>	<u>2874</u>	1956	201	FL	HOLMES
FLAS	38654	<u>W.A. Murrill</u>	<u>=</u>	1941	112	FL	JEFFERSON
US	1028731	<u>P.C. Standley</u>	<u>18952</u>	1919	351	FL	LEE
DUKE	160469	<u>R.K. Godfrey</u>	<u>53496</u>	1955	161	FL	LEON
FLAS	4582	<u>E. West</u>	<u>=</u>	1934	101	FL	LEVY
FLAS	92366	<u>S. McDaniel</u>	<u>6146</u>	1965	124	FL	LIBERTY
FLAS	113471	<u>J.T. Youree</u>	<u>143</u>	1972	113	FL	NASSAU
MICH	—	<u>E.L. Bridges</u>	<u>16723</u>	1991	139	FL	ORANGE
MICH	—	<u>J.B. McFarlin</u>	<u>5662</u>	1931	156	FL	OSCEOLA
UNA	1698	<u>A.G. Shuey</u>	<u>2390</u>	1979	295	FL	PASCO



FLAS	159700	<u>E. Wheeler</u>	<u>—</u>	1985	327	FL	POLK
NLU	413355	<u>C. Slaughter</u>	<u>10862</u>	1999	98	FL	PUTNAM
FSU	98174	<u>S. McDaniel</u>	<u>6102</u>	1965	122	FL	SUWANNEE
DUKE	138831	<u>R. Kral</u>	<u>2427</u>	1956	171	FL	WAKULLA
FLAS	101262	<u>R.R. Smith</u>	<u>1516</u>	1967	171	FL	WALTON
UNA	1617	<u>R.M. Harper</u>	<u>3548</u>	1936	57	GA	EARLY
SWSL	—	<u>T.M. Zebryk</u>	<u>132</u>	1992	104	GA	EVANS
NCSC	51372	<u>R.L. Wilbur</u>	<u>3356</u>	1953	143	GA	IRWIN
TAES	207534	<u>R. Carter</u>	<u>10347</u>	1992	224	GA	LIBERTY
SWSL	—	<u>C.T. Bryson</u>	<u>8021</u>	1988	160	GA	LOWNDES
MICH	—	<u>W.P. Adams</u>	<u>19995</u>	1956	105	GA	MCINTOSH
FLAS	182371	<u>K.A. Williges</u>	<u>46</u>	1993	195	GA	WARE
NLU	189783	<u>C.M. Allen</u>	<u>10684</u>	1981	110	LA	ALLEN
LL	—	<u>S.L. Orzell</u>	<u>7116</u>	1988	151	LA	CALCASIEU
NLU	229034	<u>R.D. Thomas</u>	<u>88087</u>	1984	109	LA	CAMERON
LL	—	<u>J.W. Thieret</u>	<u>25921</u>	1967	119	LA	ST. TAMMANY
MICH	—	<u>S.M. Tracy</u>	<u>—</u>	1898	72	MS	HARRISON
MICH	—	<u>C.T. Bryson</u>	<u>4172</u>	1986	105	MS	JACKSON
SWSL	—	<u>C.T. Bryson</u>	<u>3440</u>	1982	261	MS	OKTIBBEHA
SWSL	—	<u>C.T. Bryson</u>	<u>7065</u>	1987	247	MS	SHARKEY
NCSC	17507	<u>R.K. Godfrey</u>	<u>49189</u>	1949	141	NC	BRUNSWICK
NCU	6396	<u>H.E. Ahles</u>	<u>26225</u>	1957	147	SC	DORCHESTER
DUKE	60312	<u>D.S. Correll</u>	<u>5310</u>	1936	165	SC	GEORGETOWN
DUKE	22834	<u>C.A. Weatherby</u>	<u>16438</u>	1932	112	SC	HORRY

NCU	6398	<u>A.E. Radford</u>	<u>21396</u>	1957	111	SC	WILLIAMSBURG
LL	—	<u>E.L. Bridges</u>	<u>5064</u>	1987	106	TX	HARDIN
TAES	209301	<u>K. Northrup</u>	<u>=</u>	1992	116	TX	HARRIS
TAES	186959	<u>S. &amp; G. Jones</u>	<u>2466</u>	1989	152	TX	JASPER
MICH	—	<u>E.L. Bridges</u>	<u>6458</u>	1988	111	TX	SAN JACINTO

## APPENDIX G

## JULIAN DATES WITH CORRESPONDING GREGORIAN DATES

JULIAN	GREGORIAN	JULIAN	GREGORIAN	JULIAN	GREGORIAN	JULIAN	GREGORIAN
1	1-Jan	49	18-Feb	97	7-Apr	145	25-May
2	2-Jan	50	19-Feb	98	8-Apr	146	26-May
3	3-Jan	51	20-Feb	99	9-Apr	147	27-May
4	4-Jan	52	21-Feb	100	10-Apr	148	28-May
5	5-Jan	53	22-Feb	101	11-Apr	149	29-May
6	6-Jan	54	23-Feb	102	12-Apr	150	30-May
7	7-Jan	55	24-Feb	103	13-Apr	151	31-May
8	8-Jan	56	25-Feb	104	14-Apr	152	1-Jun
9	9-Jan	57	26-Feb	105	15-Apr	153	2-Jun
10	10-Jan	58	27-Feb	106	16-Apr	154	3-Jun
11	11-Jan	59	28-Feb	107	17-Apr	155	4-Jun
12	12-Jan	60	1-Mar	108	18-Apr	156	5-Jun
13	13-Jan	61	2-Mar	109	19-Apr	157	6-Jun
14	14-Jan	62	3-Mar	110	20-Apr	158	7-Jun
15	15-Jan	63	4-Mar	111	21-Apr	159	8-Jun
16	16-Jan	64	5-Mar	112	22-Apr	160	9-Jun
17	17-Jan	65	6-Mar	113	23-Apr	161	10-Jun
18	18-Jan	66	7-Mar	114	24-Apr	162	11-Jun
19	19-Jan	67	8-Mar	115	25-Apr	163	12-Jun
20	20-Jan	68	9-Mar	116	26-Apr	164	13-Jun
21	21-Jan	69	10-Mar	117	27-Apr	165	14-Jun
22	22-Jan	70	11-Mar	118	28-Apr	166	15-Jun
23	23-Jan	71	12-Mar	119	29-Apr	167	16-Jun
24	24-Jan	72	13-Mar	120	30-Apr	168	17-Jun
25	25-Jan	73	14-Mar	121	1-May	169	18-Jun
26	26-Jan	74	15-Mar	122	2-May	170	19-Jun
27	27-Jan	75	16-Mar	123	3-May	171	20-Jun
28	28-Jan	76	17-Mar	124	4-May	172	21-Jun
29	29-Jan	77	18-Mar	125	5-May	173	22-Jun
30	30-Jan	78	19-Mar	126	6-May	174	23-Jun
31	31-Jan	79	20-Mar	127	7-May	175	24-Jun
32	1-Feb	80	21-Mar	128	8-May	176	25-Jun
33	2-Feb	81	22-Mar	129	9-May	177	26-Jun
34	3-Feb	82	23-Mar	130	10-May	178	27-Jun
35	4-Feb	83	24-Mar	131	11-May	179	28-Jun
36	5-Feb	84	25-Mar	132	12-May	180	29-Jun
37	6-Feb	85	26-Mar	133	13-May	181	30-Jun
38	7-Feb	86	27-Mar	134	14-May	182	1-Jul
39	8-Feb	87	28-Mar	135	15-May	183	2-Jul
40	9-Feb	88	29-Mar	136	16-May	184	3-Jul
41	10-Feb	89	30-Mar	137	17-May	185	4-Jul
42	11-Feb	90	31-Mar	138	18-May	186	5-Jul
43	12-Feb	91	1-Apr	139	19-May	187	6-Jul
44	13-Feb	92	2-Apr	140	20-May	188	7-Jul
45	14-Feb	93	3-Apr	141	21-May	189	8-Jul
46	15-Feb	94	4-Apr	142	22-May	190	9-Jul
47	16-Feb	95	5-Apr	143	23-May	191	10-Jul
48	17-Feb	96	6-Apr	144	24-May	192	11-Jul

193 ..... 12-Jul	242 ..... 30-Aug	291 ..... 18-Oct	340 ..... 6-Dec
194 ..... 13-Jul	243 ..... 31-Aug	292 ..... 19-Oct	341 ..... 7-Dec
195 ..... 14-Jul	244 ..... 1-Sep	293 ..... 20-Oct	342 ..... 8-Dec
196 ..... 15-Jul	245 ..... 2-Sep	294 ..... 21-Oct	343 ..... 9-Dec
197 ..... 16-Jul	246 ..... 3-Sep	295 ..... 22-Oct	344 ..... 10-Dec
198 ..... 17-Jul	247 ..... 4-Sep	296 ..... 23-Oct	345 ..... 11-Dec
199 ..... 18-Jul	248 ..... 5-Sep	297 ..... 24-Oct	346 ..... 12-Dec
200 ..... 19-Jul	249 ..... 6-Sep	298 ..... 25-Oct	347 ..... 13-Dec
201 ..... 20-Jul	250 ..... 7-Sep	299 ..... 26-Oct	348 ..... 14-Dec
202 ..... 21-Jul	251 ..... 8-Sep	300 ..... 27-Oct	349 ..... 15-Dec
203 ..... 22-Jul	252 ..... 9-Sep	301 ..... 28-Oct	350 ..... 16-Dec
204 ..... 23-Jul	253 ..... 10-Sep	302 ..... 29-Oct	351 ..... 17-Dec
205 ..... 24-Jul	254 ..... 11-Sep	303 ..... 30-Oct	352 ..... 18-Dec
206 ..... 25-Jul	255 ..... 12-Sep	304 ..... 31-Oct	353 ..... 19-Dec
207 ..... 26-Jul	256 ..... 13-Sep	305 ..... 1-Nov	354 ..... 20-Dec
208 ..... 27-Jul	257 ..... 14-Sep	306 ..... 2-Nov	355 ..... 21-Dec
209 ..... 28-Jul	258 ..... 15-Sep	307 ..... 3-Nov	356 ..... 22-Dec
210 ..... 29-Jul	259 ..... 16-Sep	308 ..... 4-Nov	357 ..... 23-Dec
211 ..... 30-Jul	260 ..... 17-Sep	309 ..... 5-Nov	358 ..... 24-Dec
212 ..... 31-Jul	261 ..... 18-Sep	310 ..... 6-Nov	359 ..... 25-Dec
213 ..... 1-Aug	262 ..... 19-Sep	311 ..... 7-Nov	360 ..... 26-Dec
214 ..... 2-Aug	263 ..... 20-Sep	312 ..... 8-Nov	361 ..... 27-Dec
215 ..... 3-Aug	264 ..... 21-Sep	313 ..... 9-Nov	362 ..... 28-Dec
216 ..... 4-Aug	265 ..... 22-Sep	314 ..... 10-Nov	363 ..... 29-Dec
217 ..... 5-Aug	266 ..... 23-Sep	315 ..... 11-Nov	364 ..... 30-Dec
218 ..... 6-Aug	267 ..... 24-Sep	316 ..... 12-Nov	365 ..... 31-Dec
219 ..... 7-Aug	268 ..... 25-Sep	317 ..... 13-Nov	
220 ..... 8-Aug	269 ..... 26-Sep	318 ..... 14-Nov	
221 ..... 9-Aug	270 ..... 27-Sep	319 ..... 15-Nov	
222 ..... 10-Aug	271 ..... 28-Sep	320 ..... 16-Nov	
223 ..... 11-Aug	272 ..... 29-Sep	321 ..... 17-Nov	
224 ..... 12-Aug	273 ..... 30-Sep	322 ..... 18-Nov	
225 ..... 13-Aug	274 ..... 1-Oct	323 ..... 19-Nov	
226 ..... 14-Aug	275 ..... 2-Oct	324 ..... 20-Nov	
227 ..... 15-Aug	276 ..... 3-Oct	325 ..... 21-Nov	
228 ..... 16-Aug	277 ..... 4-Oct	326 ..... 22-Nov	
229 ..... 17-Aug	278 ..... 5-Oct	327 ..... 23-Nov	
230 ..... 18-Aug	279 ..... 6-Oct	328 ..... 24-Nov	
231 ..... 19-Aug	280 ..... 7-Oct	329 ..... 25-Nov	
232 ..... 20-Aug	281 ..... 8-Oct	330 ..... 26-Nov	
233 ..... 21-Aug	282 ..... 9-Oct	331 ..... 27-Nov	
234 ..... 22-Aug	283 ..... 10-Oct	332 ..... 28-Nov	
235 ..... 23-Aug	284 ..... 11-Oct	333 ..... 29-Nov	
236 ..... 24-Aug	285 ..... 12-Oct	334 ..... 30-Nov	
237 ..... 25-Aug	286 ..... 13-Oct	335 ..... 1-Dec	
238 ..... 26-Aug	287 ..... 14-Oct	336 ..... 2-Dec	
239 ..... 27-Aug	288 ..... 15-Oct	337 ..... 3-Dec	
240 ..... 28-Aug	289 ..... 16-Oct	338 ..... 4-Dec	
241 ..... 29-Aug	290 ..... 17-Oct	339 ..... 5-Dec	

## VITA

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## EDUCATION

2000-2003	M.S.	Rangeland Ecology and Management Texas A&M University, College Station, Texas
1995-2000	B.S.	Horticulture Texas A&M University, College Station, Texas Emphasis in Ecology and Botany

## HERBARIUM EXPERIENCE

2000-2003	Assistant Curator, S.M. Tracy Herbarium (TAES) Department of Rangeland Ecology and Management Texas A&M University, College Station, Texas Plant identification, curation and management of collections
1999-2000	Herbarium Assistant, Biology Department Herbarium (TAMU) Texas A&M University, College Station, Texas Organize teaching herbarium, mount herbarium specimens

## TEACHING EXPERIENCE

2003	Educational Outreach Facilitator, S.M. Tracy Herbarium (TAES), Department of Rangeland Ecology and Management, Texas A&M University, Guide outdoor native plant hikes and lead indoor plant educational activities for area schools
2002-2003	Graduate Teaching Assistant, Department of Forest Science, Texas A&M University, Prepare and teach Dendrology Laboratory
2002	Graduate Teaching Assistant, Department of Botany, Texas A&M University, Prepare and teach Taxonomy of Flowering Plants Laboratory
2001	Graduate Teaching Assistant, Department of Rangeland Ecology and Management, Texas A&M University, Prepare and teach Plant Taxonomy Laboratory

## AWARDS &amp; ACTIVITIES

2002	Member of Society of Ecological Restoration, Texas Chapter Attended Joint ESA/SER annual meeting in Tucson, Arizona
2001	F. W. Gould Research Award in Plant Systematics, Research Grant
1998	International Institute of Tropical Agriculture in Ibadan, Nigeria

## PERSONAL

1977	Born February 9, 1977
2002	Married Darrel McLaughlin November 27, 2002

## ENJOY

Botanizing, Camping, Hiking, Fishing, Mountain biking and Sewing